

ORIGINAL ARTICLE

Biometric relationships between body size and otolith size in 15 demersal marine fish species from the northern Brazilian coast

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ABSTRACT

The relationships between fish size and sagitta otolith measurements were calculated for the first time for 15 species belonging to six families from the northern Brazilian coast. A total of 220 fish were sampled from the bycatch landed by the bottom-trawl industrial shrimp-fishing fleet between August and September 2016. All species had strong relationships between otolith measurements and fish total length with the coefficient of determination (r^2) ranging between 0.71 and 0.99. The variable most strongly related to fish total length was found to be the sagittal otolith length (OL) with 98% of the variability. These relationships are a useful tool to estimate length and mass of preyed fish from otoliths found in stomach contents of marine predators.

KEYWORDS: Amazon delta, bycatch, sagitta, Neotropics, Sciaenidae, stomach content

Relação biométrica entre tamanho corporal e tamanho do otólito em 15 espécies de peixes marinhos demersais da costa norte do Brasil

RESUMO

As relações entre o tamanho corporal do peixe e as medidas do otólito sagitta foram calculadas pela primeira vez para 15 espécies pertencentes a seis famílias de peixes demersais da costa norte do Brasil. Um total de 220 peixes foi amostrado da captura acidental realizada pelo arrasto de fundo da frota pesqueira industrial de camarão entre agosto e setembro de 2016. Todas as espécies apresentaram fortes relações entre as medidas do otólito e o comprimento total do peixe, com o coeficiente de determinação (r^2) variando entre 0,71 e 0,99. A variável mais fortemente relacionada ao comprimento total dos peixes foi o comprimento do otólito sagital (OL) com 98% da variabilidade. Essas relações representam uma ferramenta útil para estimar o comprimento e a massa de peixes predados a partir dos otólitos encontrados no conteúdo estomacal de predadores marinhos.

PALAVRAS-CHAVE: Delta do Amazonas, captura acidental, sagitta, Neotrópico, Scianidae, conteúdo estomacal

INTRODUCTION

Otoliths are structures composed mainly of calcium carbonate, located in the inner ear of Osteichthyes, which have body balancing and hearing as main functions (Campana 2004). These structures are arranged in pairs, called sagitta, asteriscus, and lapillus in bony fish, and vary widely in size and shape among species (Campana 2004; Popper *et al.* 2005). Due to its largest size in the majority of bone fishes, sagittae otoliths are the most suitable for systematic and ecological studies, for taxon identification, age estimation, and life history tracking (Harvey *et al.* 2000;

De La Cruz-Agüero *et al.* 2016; Assis *et al.* 2018). Their non-digestible calcified structure has allowed them to be widely used to identify fishes ingested by different predators such as aquatic mammals, seabirds and fishes (Battaglia *et al.* 2010; Tuset *et al.* 2010). Furthermore, relationships between fish size and otolith measures are useful to elucidate the feeding behavior of piscivorous fauna, providing subsidies for the management of these species (De Pierrepont *et al.* 2005; Lombarte *et al.* 2006; Battaglia *et al.* 2010).

Studies on otoliths in marine fish from the Western South Atlantic are scarce (Waessle *et al.* 2003; Assis *et al.*

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2018; Souza *et al.* 2019). The high nutrient and sediment load from the Amazon River, near the Northern Coast of Brazil favors the occurrence of abundant fishery stocks, mainly shrimp (Penaeidae) and several fish species (Isaac and Braga 1999; Marceniuk *et al.* 2019). The ichthyofauna is functionally important as an intermediate trophic level for many consumers, however, its importance is not well understood (Barletta *et al.* 2010).

The present study aimed to determine the relationship between fish size (i.e. length and weight) and sagittae otolith measurements (i.e. length, width, and weight) for the 15 most abundant demersal species captured along the northern Brazilian coast.

MATERIAL AND METHODS

Fish were captured between August and September 2016 in an area characterized by the estuarization of inshore waters, due to the proximity to the Amazonas River (5°02'21.6"N, 47°49'33.9"W; 0°51'02.0"N, 47°50'30.0"W northern and southern limits of the sampling area, respectively; Figure 1), and is inserted in the world's second largest mangrove (~700 000 ha), which is an important fishery area (Isaac and Braga 1999; Giarrizzo and Krumne 2008).

Samples were randomly collected from the bottom trawl of industrial shrimp trawlers using a 30 x 21 mm mesh bottom trawl of 22.4 m length towed at velocity of ~2.5 knots at 35 – 49 m depth. Bycatch were taken to the laboratory and kept frozen until processing. Sampled fish were identified to species level, and measured (standard

length, SL, and total length, TL, precision 0.01 cm) and weighed for total body weight (BW, 0.01 g). Sagittae otoliths were removed, cleaned and stored dry in coded microtubes. Each otolith was weighed (WO, 0.0001 g) using an analytical balance, and measured for maximum length (OL, 0.001 cm), as the horizontal distance between the anterior and posterior tips of the sagitta, and width (OW, 0.001 cm), as the greater distance between the dorsal and ventral margins of the otolith (Harvey *et al.* 2000; Battaglia *et al.* 2010).

Vouchers of each species were fixed in 10% formalin after processing, then preserved in 70% alcohol and deposited in the ichthyological collection of the Grupo de Ecologia Aquática (GEA) at Universidade Federal do Pará (UFPA). Potential differences between the dimensions of the right and left sagittae otoliths were tested using a paired Student's t-test per species (Park *et al.* 2018). The length-length relationship (LLR) was determined by the method of least squares to fit a simple linear regression model: $TL = a + bSL$. The length-weight relationship (LWR) was determined as: $W = aSL^b$, and was fitted to the data using a linear regression of the \log_{10} -transformed data. Morphometric relationships between TL and otolith dimensions were calculated using linear ($Y = a + bX$) and linearizable ($Y = aX^b$) regression models, which best fit the data. When present, outliers were removed by graphical inspection of the plot before performing the regression analyses (Froese *et al.* 2011). The coefficient of determination (Pearson *r*-squared, r^2) was used as indicator of regression quality and to check if fish growth (b) was statistically different from isometric growth, a t-test ($H_0: b = 3$) (Froese *et al.* 2011). A significance level of $\alpha < 0.05$ (confidence level $\pm 95\%$) was routinely adopted.

RESULTS

The analyses were performed using 220 specimens from 15 species (see Figure 2 for otoliths) and six families. The most representative family was Sciaenidae with nine species, followed by Haemulidae, with two species, and the remaining four families, with only one species each (Table 1). Body weight ranged between 6.7 to 911.1 g, SL between 7.1 to 69.5 cm, and TL from 8.9 to 72.1 cm. Paired t-tests did not detect differences between left and right sagittae otoliths for OL, OW, and WO ($p > 0.05$ for all species). Hence, all further analyses were standardized by using only the left otolith measures.

Differences between the b values estimated by the different regression models were species specific. However, for species of the same family with similar body shape (e.g. Sciaenidae and Haemulidae), values were similar (Figure 3). Length-weight relationships (LWR) were highly significant ($p < 0.001$) only for six species: *Ctenosciaena gracilicirrhus*, *Macrodon ancylodon*, *Menticirrhus americanus* (all Sciaenidae), *Haemulon steindachneri* (Haemulidae),

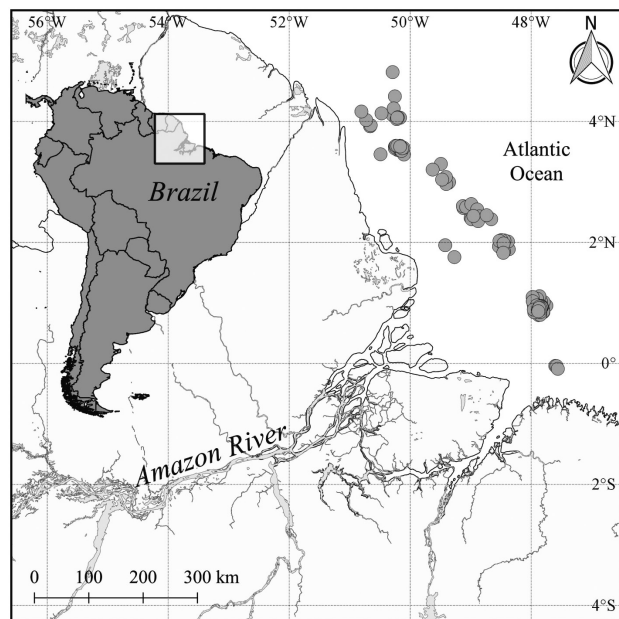


Figure 1. Sampling locations of the study on the Northern Brazilian Coast. Samples of demersal fish were collected from bottom trawling of industrial shrimp trawlers along the coast of Pará and Amapá states in Brazil (grey dots).

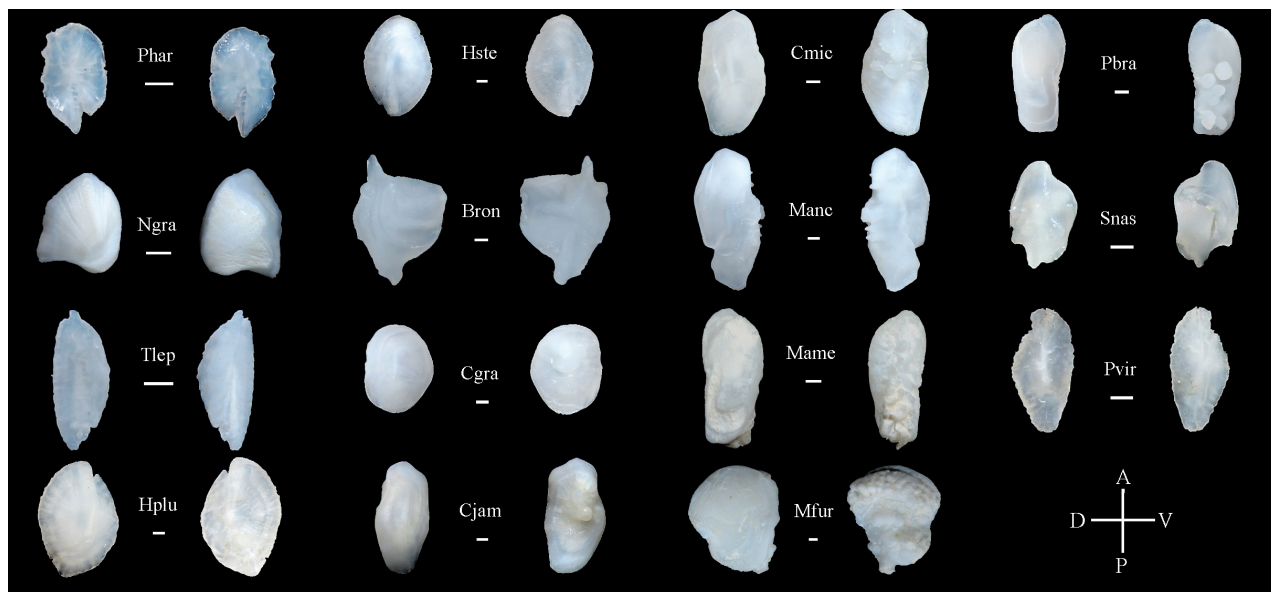


Figure 2. Left sagitta otolith morphology of the 15 fish species captured by bottom trawling along the northern Brazilian coast. Phar: *Pellona harroweri*, Ngra: *Notarius grandicassis*, Tlep: *Trichiurus lepturus*, Hplu: *Haemulon plumierii*, Hste: *Haemulon steindachneri*, Bron: *Bairdiella ronchus*, Cgra: *Ctenosciaena gracilicirrhus*, Cjam: *Cynoscion jamaicensis*, Cmic: *Cynoscion microlepidotus*, Manc: *Macrodon ancylodon*, Mame: *Menticirrhus americanus*, Mfur: *Micropogonias furnieri*, Pbra: *Paralonchurus brasiliensis*, Snas: *Stellifer naso*, Pvir: *Polydactylus virginicus*, D: dorsal, V: ventral, A: anterior, P: posterior. Scale bars = 1 mm. This figure is in color in the electronic version.

Table 1. Sample size (N) and ranges of the standard length (SL), total length (TL), weight (W), otolith length (OL) and otolith width (OW) of 15 demersal fish species captured by bottom trawl in the Northern Brazilian coast in 2016.

Family/Species	Voucher	N	SL (cm)	TL (cm)	W (g)	OL (cm)	OW (cm)	WO (g)
Pristigasteridae								
<i>Pellona harroweri</i> (Fowler, 1917)	GEA4157	9	10.5-13.2	13.8-14.6	23.98-28.35	0.131-0.445	0.079-0.271	0.0068-0.0099
Ariidae								
<i>Notarius grandicassis</i> (Valenciennes, 1840)	GEA4154	10	14.7-42.3	17.5-46.5	50.38-911.12	0.111-0.233	0.088-0.193	0.1242-1.2638
Trichiuridae								
<i>Trichiurus lepturus</i> Linnaeus, 1758		10	43.3-69.5	47.7-72.1	63.33-232.92	0.371-0.540	0.156-0.204	0.0035-0.0103
Haemulidae								
<i>Haemulon plumierii</i> (Lacepède, 1801)	GEA4164	14	14.5-19.0	17.5-20.9	94.75-159.98	0.097-0.170	0.080-0.125	0.0619-0.1715
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	GEA4662	13	14.2-18.2	19.3-22.1	105.3-168.03	0.141-0.196	0.095-0.120	0.1041-0.1223
Sciaenidae								
<i>Bairdiella ronchus</i> (Cuvier, 1830)		8	19.2-31.4	23.4-37.6	207.76-756.00	0.145-0.254	0.095-0.210	0.0124-2.6202
<i>Ctenosciaena gracilicirrhus</i> (Metzelaar, 1919)	GEA4166	26	7.1-13.4	8.9-16.4	9.29-73.74	0.446-0.735	0.380-0.582	0.0242-0.1091
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1883)		20	12.3-21.5	16.3-23.0	47.91-160.14	0.880-1.221	0.446-0.613	0.0613-0.2203
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)		13	14.5-34.5	17.0-38.5	39.70-625.60	0.139-0.260	0.074-0.140	0.0747-0.3090
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	GEA3702	31	14.4-25.3	18.2-30.9	47.20-242.20	0.943-1.341	0.431-0.709	0.0618-0.1979
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	GEA4311	16	9.5-14.0	12.0-17.0	22.73-82.02	0.088-0.120	0.075-0.092	0.0468-0.1079
<i>Micropogonias furnieri</i> (Desmarest, 1823)	GEA4299	14	12.1-27.8	17.8-33.9	51.32-448.70	0.150-0.247	0.060-0.187	0.0518-0.7530
<i>Paralonchurus brasiliensis</i> (Steindachner, 1875)	GEA4914	10	11.0-18.5	15.6-22.0	38.20-129.00	0.140-0.193	0.067-0.093	0.0587-0.0913
<i>Stellifer naso</i> (Jordan, 1889)	GEA4715	15	7.7-8.1	9.4-11.2	6.70-13.12	0.386-0.433	0.243-0.296	0.0118-0.0133
Polynemidae								
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	GEA4834	11	11.9-19.0	17.0-25.3	42.39-177.99	0.415-0.574	0.226-0.287	0.0052-0.0128

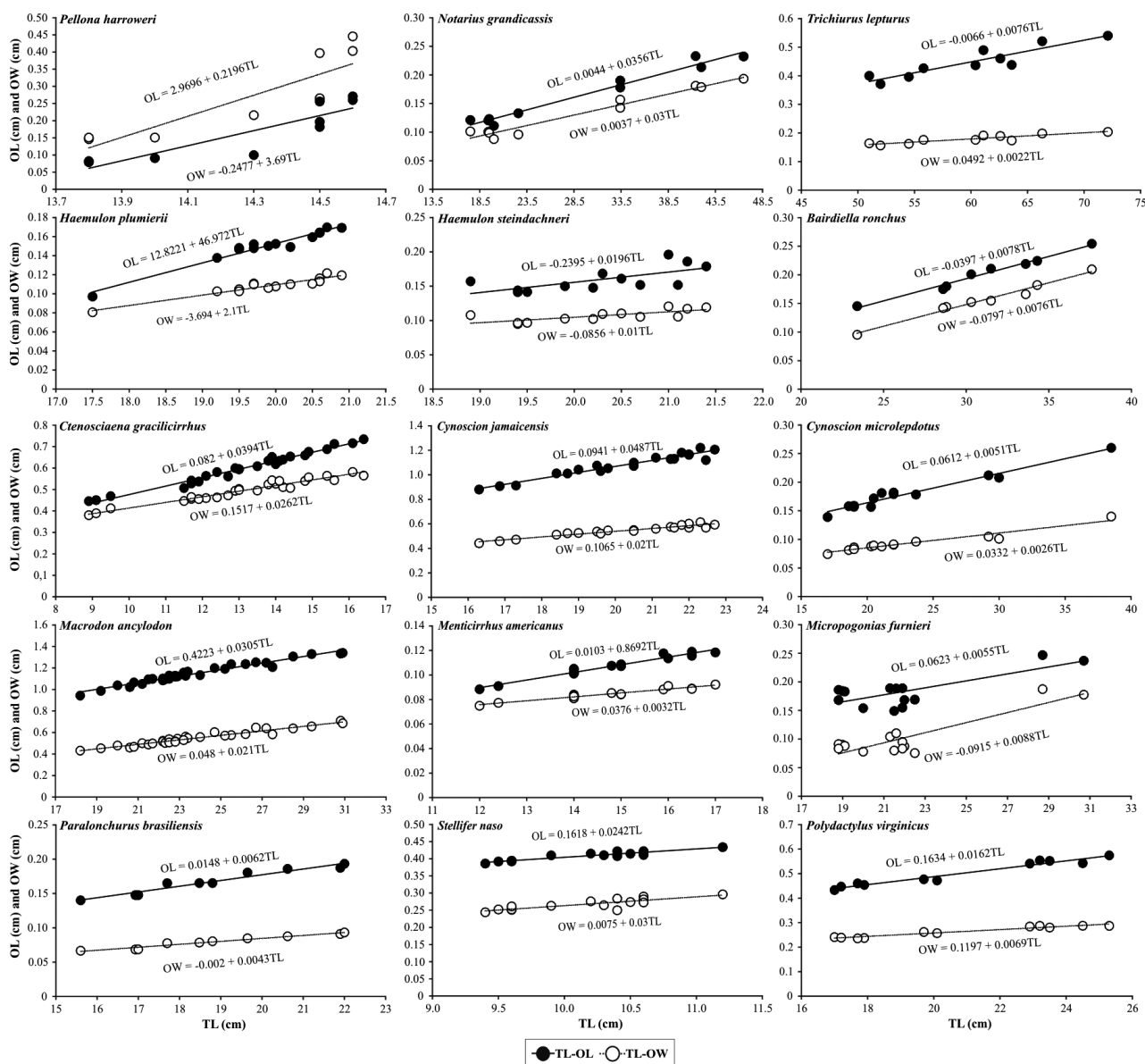


Figure 3. Relationship of otolith length (OL) and width (OW) with total length (TL) of 15 fish species captured by bottom trawling along the northern Brazilian coast.

Pellona harroweri (Pristigasteridae), and *Polydactylus virginicus* (Polynemidae) (Table 2). However, all r^2 values were greater than 0.91 in LWRs. The slope b of the LWR ranged from 2.75 for *Notarius grandicassis* to 3.54 for *H. steindachneri*. Otolith–fish body biometric relationships (i.e. BW-OL, BW-WO, BW-OW, TL-OL, TL-OW, WO-TL, WO-OL, and WO-OW) were highly correlated with r^2 , ranging from 0.713 in TL-OW for *Stellifer naso* to 0.980 in TL-OL for *Bairdiella ronchus*, and most of the relationships were significant ($p < 0.01$) (Table 2).

DISCUSSION

Body size and mass relationships are important tools for the functional understanding of a species at specific locations

(Froese *et al.* 2011). Generally the length–weight relationships (LWRs) are used for converting lengths into fish mass and vice versa (Froese 2006; Froese *et al.* 2011), and length–length relationships (LLRs) are used to convert one length into another (e.g. standard length to total length). Additionally, assuming that otolith size is closely correlated to fish size, and its shape is species specific (Campana 2004), it is suggested that otolith analysis is a feasible and reliable method to identify fish species and to estimate fish size and weight (Battaglia *et al.* 2010; Park *et al.* 2018). Froese (2006) suggests that the angular coefficient does not differ from isometry when $b = 3$. Accordingly our b values of the BW-TL relationship were isometric for sciaenids, which, in addition, had coefficients of determination (r^2) higher than 0.90 (see Figure 3). However,

Table 2. Estimated parameters of the relationship between measurements of fish and *sagitta* otoliths in 15 demersal fish species captured by bottom trawl along the northern Brazilian coast in 2016.

Species	SL-TL							BW-TL						
	a	SE a	b	SE b	r ²	G	Model	a	SE a	b	SE b	r ²	G	Model
<i>Pellona harroweri</i>	0.2838	0.025	7.77	0.362	0.95	+	L	0.0104	1.0308	2.9513	0.0114	0.99	-	NL
<i>Notarius grandicassis</i>	1.03	0.032	2.81	0.899	0.99	l	L	2.2292	1.0617	0.6588	0.0596	0.96	-	NL
<i>Trichiurus lepturus</i>	0.7969	0.023	16	1.281	0.99	+	L	0.0002	3.5751	3.209	0.3127	0.93	l	NL
<i>Haemulon plumierii</i>	0.6535	0.089	8.96	1.498	0.82	+	L	0.0176	1.4708	2.9973	0.129	0.98	l	NL
<i>Haemulon steindachneri</i>	0.6209	0.022	10.1	0.365	0.99	+	L	0.0029	1.1984	3.5416	0.06	1	+	NL
<i>Bairdiella ronchus</i>	1.142	0.144	1.28	3.778	0.91	l	L	0.0275	1.6226	2.8464	0.1412	0.99	l	NL
<i>Ctenoscaena gracilicirrus</i>	1.0738	0.044	1.77	0.475	0.96	-	L	0.0042	1.3198	3.4748	0.108	0.98	+	NL
<i>Cynoscion jamaicensis</i>	0.8012	0.041	6.11	0.722	0.96	+	L	0.0055	1.8035	3.2789	0.1965	0.94	l	NL
<i>Cynoscion microlepidotus</i>	1.0567	0.034	2.48	0.697	0.99	l	L	0.0043	2.4226	3.2398	0.2833	0.92	l	NL
<i>Macrodon ancylodon</i>	1.0541	0.055	3.74	1.053	0.93	l	L	0.0026	1.5049	3.3368	0.1291	0.96	+	NL
<i>Menticirrus americanus</i>	1.107	0.043	1.39	0.518	0.98	-	L	0.0038	1.5617	3.5124	0.166	0.97	+	NL
<i>Micropogonias furnieri</i>	0.7445	0.029	9.31	0.513	0.98	+	L	0.0041	2.5242	3.2587	0.3014	0.91	l	NL
<i>Paralichthys brasiliensis</i>	0.8536	0.093	6.68	1.351	0.91	+	L	0.0067	2.5334	3.1352	0.3168	0.92	l	NL
<i>Stellifer naso</i>	0.2287	0.004	5.59	0.042	1	+	L	0.0038	1.8594	3.3745	0.2676	0.92	l	NL
<i>Polydactylus virginicus</i>	1.2467	0.081	1.39	1.277	0.96	l	L	0.0028	1.3329	3.4219	0.0949	0.99	+	NL
Species	BW-OL							BW-OW						
	a	SE a	b	SE b	r ²	G	Model	a	SE a	b	SE b	r ²	G	Model
<i>Pellona harroweri</i>	-7.1025	0.7331	9.462	1.0914	0.93	+	NL	-0.1292	0.014	1.31	1E-04	0.92	-	NL
<i>Notarius grandicassis</i>	2228.98	2.664	4.9264	0.3438	0.89	+	NL	1571.8	2.748	5.11	0.396	0.87	+	NL
<i>Trichiurus lepturus</i>	1523.7	1.4599	3.2232	0.4627	0.86	l	NL	0.0706	1.142	0.2	0.028	0.86	-	NL
<i>Haemulon plumierii</i>	823.392	1.1803	0.942	0.087	0.91	-	NL	2700.3	1.274	1.34	0.109	0.93	-	NL
<i>Haemulon steindachneri</i>	0.0491	0.0095	0.0009	0.0001	0.93	-	L	0.0207	1.276	0.35	0.05	0.81	-	NL
<i>Bairdiella ronchus</i>	0.1077	0.0075	0.0002	1E-05	0.97	-	L	0.0053	1.329	0.55	0.046	0.96	-	NL
<i>Ctenoscaena gracilicirrus</i>	0.2616	1.0262	0.2388	0.0074	0.98	-	NL	0.2535	1.028	0.19	0.008	0.96	-	NL
<i>Cynoscion jamaicensis</i>	0.3527	1.0978	0.24	0.0201	0.89	-	NL	0.1906	1.098	0.23	0.02	0.87	-	NL
<i>Cynoscion microlepidotus</i>	0.0712	1.1087	0.1972	0.0219	0.88	-	NL	0.0398	1.123	0.18	0.025	0.83	-	NL
<i>Macrodon ancylodon</i>	0.4867	1.0457	0.1866	0.0097	0.93	-	NL	0.1662	1.089	0.26	0.018	0.87	-	NL
<i>Menticirrus americanus</i>	0.042	1.0605	0.2412	0.0152	0.95	-	NL	0.0468	1.047	0.15	0.012	0.92	-	NL
<i>Micropogonias furnieri</i>	4.5867	0.6869	5.3525	0.261	0.73	-	NL	1.9403	0.287	3.96	0.09	0.74	-	NL
<i>Paralichthys brasiliensis</i>	0.1699	2.4426	3.3286	0.498	0.85	l	NL	0.257	1.166	0.28	0.037	0.88	-	NL
<i>Stellifer naso</i>	0.2826	1.0533	0.1632	0.023	0.8	-	NL	0.1954	0.015	0.01	0.002	0.78	-	L
<i>Polydactylus virginicus</i>	0.2068	1.043	0.1971	0.0094	0.98	-	NL	0.1294	1.065	0.16	0.014	0.93	-	NL
Species	BW-WO							TL-OL						
	a	SE a	b	SE b	r ²	G	Model	a	SE a	b	SE b	r ²	G	Model
<i>Pellona harroweri</i>	15.0327	1.9515	1364.5	227.826	0.84	+	L	-2.9696	0.5966	0.2196	0.0417	0.83	-	L
<i>Notarius grandicassis</i>	13.247	1.4257	2.8074	0.0945	0.87	+	NL	0.0044	0.0003	0.0356	0.0086	0.97	-	L
<i>Trichiurus lepturus</i>	0.0008	0.0006	4E-05	5E-06	0.89	l	NL	-0.0066	0.0695	0.0076	0.0012	0.95	-	L
<i>Haemulon plumierii</i>	426.234	1.0712	0.5443	0.0328	0.96	-	NL	12.8221	0.4434	46.972	2.9262	0.96	+	L
<i>Haemulon steindachneri</i>	0.0735	0.0055	0.0003	0.00004	0.82	-	L	-0.2395	0.0334	0.0196	0.0016	0.93	-	L
<i>Bairdiella ronchus</i>	3.00E-11	9.2668	3.789	0.3609	0.95	+	NL	-0.0397	0.0141	0.0078	0.0005	0.98	-	L
<i>Ctenoscaena gracilicirrus</i>	0.0049	1.073	0.7016	0.0202	0.98	-	NL	0.082	0.0162	0.0394	0.0012	0.98	-	L
<i>Cynoscion jamaicensis</i>	0.0025	1.4058	0.8477	0.0732	0.88	-	NL	0.0941	0.0595	0.0487	0.0029	0.97	-	L
<i>Cynoscion microlepidotus</i>	0.016	1.3556	0.4604	0.0647	0.82	-	NL	0.0612	0.0077	0.0051	0.0003	0.96	-	L
<i>Macrodon ancylodon</i>	0.0052	1.1351	0.6595	0.0275	0.95	-	NL	0.4223	0.0288	0.0305	0.0012	0.96	-	L
<i>Menticirrus americanus</i>	0.006	1.1943	0.6593	0.046	0.94	-	NL	0.0103	1.1219	0.8692	0.0428	0.97	-	NL
<i>Micropogonias furnieri</i>	0.8819	0.1071	2.7139	0.0096	0.82	-	NL	0.0623	0.0096	0.0055	0.0004	0.93	-	L
<i>Paralichthys brasiliensis</i>	0.0439	0.0036	0.0004	0.00005	0.9	-	L	0.0148	0.0049	0.0062	0.0003	0.96	-	L
<i>Stellifer naso</i>	0.0089	1.0689	0.1532	0.0295	0.76	-	NL	0.1618	0.0247	0.0242	0.0024	0.89	-	L
<i>Polydactylus virginicus</i>	0.0007	1.3338	0.5665	0.0642	0.9	-	NL	0.1634	0.0237	0.0162	0.0011	0.96	-	L

Table 2. Continued.

Species	TL-OW							WO-TL						
	a	SE a	b	SE b	r ²	G	Model	a	SE a	b	SE b	r ²	G	Model
<i>Pellona harroweri</i>	-0.2477	0.026	3.69	0.376	0.93	I	L	255.881	41.48	12.116	0.3553	0.85	+	L
<i>Notarius grandicassis</i>	0.0037	3.00E-04	0.03	0.008	0.96	-	L	2.5465	1.1177	1.6438	0.0296	0.9	-	L
<i>Trichiurus lepturus</i>	0.0492	0.0224	0.0022	0.0004	0.94	-	L	0.8843	1.0286	0.0333	0.0069	0.74	-	NL
<i>Haemulon plumierii</i>	-3.6941	0.221	2.1	0.17	0.93	-	L	1.00E-08	2.1111	5.3811	0.2499	0.98	+	NL
<i>Haemulon steindachneri</i>	-0.0856	0.016	0.01	8E-04	0.93	-	L	-0.0284	0.0191	0.0069	0.0009	0.83	-	L
<i>Bairdiella ronchus</i>	-0.0797	0.0151	0.0076	0.0005	0.97	-	L	3.61E-17	50.065	10.8	1.1415	0.94	+	NL
<i>Ctenoscaena gracilicirrhus</i>	0.1517	0.0166	0.0262	0.0013	0.95	-	L	0.0001	1.3115	2.436	0.1056	0.96	-	NL
<i>Cynoscion jamaicensis</i>	0.1065	0.019	0.02	9E-04	0.97	-	L	0.00002	1.8332	2.9321	0.202	0.92	I	NL
<i>Cynoscion microlepidotus</i>	0.0332	0.005	0.0026	0.0002	0.93	-	L	0.0009	1.7738	1.6016	0.1835	0.87	-	NL
<i>Macrodon ancylodon</i>	0.048	0.0234	0.021	0.001	0.94	-	L	0.0001	1.3917	2.234	0.1044	0.94	-	NL
<i>Menticirrhus americanus</i>	0.0376	0.0032	0.0032	0.0002	0.94	-	L	0.0001	1.5065	2.3625	0.1527	0.95	-	NL
<i>Micropogonias furnieri</i>	-0.0915	0.0298	0.0088	0.0013	0.92	-	L	1E-06	2.4738	3.7761	0.2948	0.93	+	NL
<i>Paralichthys brasiliensis</i>	-0.002	0.0061	0.0043	0.0003	0.97	-	L	0.0022	2.0483	1.1875	0.2444	0.75	-	NL
<i>Stellifer naso</i>	0.0075	0.046	0.03	0.005	0.71	-	L	0.0034	1.2313	0.5693	0.0898	0.76	-	NL
<i>Polydactylus virginicus</i>	0.1197	0.0119	0.0069	0.0006	0.95	-	L	0.00002	2.002	1.9361	0.2292	0.89	-	NL

Species	WO-OL							WO-OW						
	a	SE a	b	SE b	r ²	G	Model	a	SE a	b	SE b	r ²	G	Model
<i>Pellona harroweri</i>	-95.2863	20.063	1.0512	0.1719	0.76	-	L	0.6918	0.104	64	12.17	0.8	+	L
<i>Notarius grandicassis</i>	677.397	1.9976	1.7663	0.2428	0.92	-	NL	459.89	2.281	1.88	0.323	0.87	-	NL
<i>Trichiurus lepturus</i>	0.0444	1.3164	2.6697	0.3361	0.89	I	NL	2.4086	2.378	3.57	0.502	0.86	I	NL
<i>Haemulon plumierii</i>	3.3557	1.2519	1.7313	0.1179	0.95	-	NL	25.986	1.574	2.4	0.204	0.92	-	NL
<i>Haemulon steindachneri</i>	0.0565	0.0063	0.3489	0.0391	0.88	-	L	0.0365	0.01	0.71	0.094	0.84	-	L
<i>Bairdiella ronchus</i>	617102	6.4754	8.7848	1.1499	0.91	+	NL	187418	2.238	6.92	0.425	0.98	+	NL
<i>Ctenoscaena gracilicirrhus</i>	0.2456	1.0538	2.8943	0.0976	0.97	I	NL	0.6417	1.137	3.49	0.18	0.94	+	NL
<i>Cynoscion jamaicensis</i>	0.1006	1.0254	3.4241	0.2167	0.93	+	NL	1.1632	1.167	3.6	0.249	0.92	+	NL
<i>Cynoscion microlepidotus</i>	7.6983	1.3784	2.3376	0.1851	0.94	-	NL	34.937	2.156	2.33	0.322	0.83	-	NL
<i>Macrodon ancylodon</i>	0.0675	1.0254	3.3869	0.1557	0.94	+	NL	0.4269	1.105	2.28	0.162	0.87	-	NL
<i>Menticirrhus americanus</i>	33.7587	1.2794	2.7191	0.1097	0.98	-	NL	1604.8	2.6	4.03	0.386	0.89	+	NL
<i>Micropogonias furnieri</i>	727.085	2.1282	5.0159	0.441	0.92	+	NL	23.157	1.476	2.15	0.162	0.94	-	NL
<i>Paralichthys brasiliensis</i>	-0.5903	0.0218	0.1707	0.1293	0.75	-	L	0.003	1.81	1.24	0.234	0.78	-	NL
<i>Stellifer naso</i>	-1.5281	0.039	0.953	0.1002	0.87	-	L	0.0057	6.00E-04	0.03	0.002	0.92	-	L
<i>Polydactylus virginicus</i>	0.0616	1.2884	2.8049	0.3601	0.87	I	NL	0.7464	1.957	3.33	0.501	0.83	I	NL

BW: Body weight, TL: Total length, OL: otolith length, OW: otolith width, WO: weight of the otolith, SE: standard error, G: growth type = -: negative allometric, +: positive allometric, I: isometric, NL: Nonlinear equation, L: linear equation. *b* values in bold are significant (t-test; $p < 0.05$).

despite the strong biometric relationships derived from our data, our estimated parameters should be used with caution, as our small sample sizes (mainly those ≤ 10 for *B. ronchus*, *P. harroweri*, *N. grandicassis*, *Trichiurus lepturus*, and *Paralichthys brasiliensis*) and a selective effect of the mesh size used by the shrimp trawlers may have caused the size distributions in our samples to be underrepresented.

The lack of statistical differences between left and right sagitta indicates that otoliths on either body side are indistinctly usable for fish-size estimations (Battaglia *et al.* 2010; Mehanna *et al.* 2016; Park *et al.* 2018; Yilmaz *et al.* 2015). The high coefficients of determination for the relationships between otolith measurements and fish size in all our species indicates that length or weight of fish can be

reliably estimated from otoliths found in stomach contents of predators. Our values of *b* varied considerably among the species, owing to the variable size and shape of the sagittae among the species. Yet, at family level (e.g. Scianidae), the *b* values tended to negative allometry in most relationships, as species within families are relatively more similar in shape. Most studies providing relationships between otolith and fish size have used only the width and length of the otolith (Giménez *et al.* 2016; Assis *et al.* 2018; Park *et al.* 2018). Considering the high values of correlation in our analyses, the inclusion of otolith weight in our estimations contributed to strengthen the relationship models.

The relationship between otoliths and fish size was estimated for species from different regions around the

world, such as Mexico (De La Cruz-Agüero *et al.* 2016), India (Aneesh Kumar *et al.* 2017), Turkey (Altin and Ayyildiz 2017), Montenegro (Kanjuh *et al.* 2018) and Australia (Park *et al.* 2018). However, studies on this subject are still incipient for the South Atlantic. An analysis of the otolith–fish size relationship for juveniles of four species of Scianidae in the De la Plata River estuary, in southern Uruguay (Waessle *et al.* 2003), included three of our species (*M. ancylodon*, *M. furnieri*, and *P. brasiliensis*), which showed similar parameter values to those estimated in our study. Assis *et al.* (2018) analyzed seven species of commercial importance from the coast of Bahia, in northeastern Brazil, and Souza *et al.* (2019) studied *Pomatomus saltatrix* from the coast of Rio de Janeiro in the southeast. Besides their economic importance, some of these fish species, such as *M. ancylodon*, *M. americanus*, *M. furnieri*, *P. harrowerri*, and *T. lepturus*, are common prey of the estuarine dolphin, *Sotalia guianensis* (Van Bénédén, 1864), that occurs along the Brazilian coast (Vieira 2014), further highlighting the importance of our biometric estimators for ecological studies.

CONCLUSIONS

This study is a contribution to the knowledge about the relationships between otolith and fish size in 15 fish species from the northern Brazilian coast for the reliable estimation of species-specific fish length or weight from otolith size. Our results form a baseline for future studies on trophic ecology and fish distribution, and will enable a more accurate evaluation of length and/or biomass of demersal fishes consumed by predators.

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