MUSEU PARAENSE EMÍLIO GOELDI

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PROGRAMA DE PÓS-GRADUAÇÃO EM ZOOLOGIA

CURSO DE MESTRADO EM ZOOLOGIA

Revisão taxonômica do grupo *rubripes* do gênero *Corinna* Koch, 1842 (Araneae; Corinnidae)

BRUNO VINICIUS BASTOS RODRIGUES

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Dissertação apresentada ao Programa de Pós-Graduação em Zoologia, Curso de Mestrado, do Museu Paraense Emílio Goeldi e Universidade Federal do Pará como requisito parcial para obtenção do grau de mestre em Zoologia.

Orientador: Dr. Alexandre Bragio Bonaldo

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SUMÁRIO

INTRODUÇÃO GERAL

A família Corinnidae Karsch, 1880 apresenta grande diversidade, compreendendo atualmente 1032 espécies em 87 gêneros (Platnick 2013), distribuídas em zonas temperadas e tropicais. O posicionamento filogenético de Corinnidae em relação a outras famílias de Dionycha ainda não está bem estabelecido. Alguns autores propuseram hipóteses para compreender essa relação. Lehtinen (1967) sugeriu que Corinnidae estaria mais relacionado com Zodariinae dentro da superfamília Zodarioidea, juntamente com outros 14 grupos, que compartilhariam uma tendência à redução das fiandeiras posteriores. Penniman (1985) estabeleceu um clado que inclui Gnaphosidae como grupo-irmão de Corinnidae devido a características morfológicas das fiandeiras médias posteriores. Coddington & Levi (1991) sugeriram uma hipótese de relação entre as famílias de Araneomorphae, baseados em um conjunto de informações disponíveis até então sobre o conhecimento sistemático de Araneae. A topologia proposta por esses autores inclui um grupo monofilético composto por Corinnidae e Liocranidae, dentro do clado Dionycha. Lehtinen (1996) sugeriu, com reservas, uma relação próxima entre essas duas famílias. Entretanto, até mesmo a monofilia de Corinnidae ainda é discutida. Atualmente, a família é dividida em quatro subfamílias: Corinninae Karsch, Castianeirinae Reiskind, Trachelinae Simon e Phrurolithinae Banks. Somente Corinninae e Castianeirinae são claramente estabelecidas. Segundo Ramírez (comunicação pessoal), a partir de uma análise filogenética de Dionycha, Trachelinae e Phrurolithinae provavelmente não serão mais incluídas em Corinnidae. A subfamília Corinninae Karsch (1880) é ainda o único grupo de Corinnidae cujos gêneros neotropicais foram recentemente caracterizados. Bonaldo (2000) reconheceu dezessete gêneros para esta subfamília.

O gênero *Corinna* foi proposto por C.L. Koch (1842), e como salientado por Bonaldo (1996), logo foi transformado em um depositário artificial de espécies, no sentido de que várias espécies de Corinninae sem características derivadas óbvias, descritas nos séculos XIX e XX, foram incluídas no gênero. Bonaldo (1996) propôs uma sinapomorfia putativa para *Corinna*, a presença de um condutor esclerotizado, indicando que todas as espécies com condutor hialino (um caráter provavelmente primitivo) deveriam ser transferidas para outros gêneros. Segundo Bonaldo (2000), *Corinna* é caracterizado pela combinação dos seguintes caracteres: carapaça sub-retangular, região cefálica bem diferenciada, com bordas laterais sinuosas, infladas e projetadas. Palpo do macho com apófise tibial retrolateral única; processo tegular triangular, virguliforme ou digitiforme; condutor esclerotinizado, com margem prolateral dobrada ventralmente, formando uma calha onde se aloja o êmbolo; êmbolo longo, filiforme ou achatado; epígino com uma abertura de copulação, geralmente anterior ao ducto de copulação. Atualmente, *Corinna* é o quarto gênero mais diverso de Corinnidae com 69 espécies válidas (Platnick 2013). Destas, 58 ocorrem na região Neotropical. Entretanto, Bonaldo (2000) indicou que as espécies africanas atualmente alocadas em *Corinna*, não pertencem a este gênero. Na América do Sul, o gênero está distribuído em quase todos os países, exceto Chile, Bolívia e Suriname. A ausência do gênero na Bolívia e Suriname, provavelmente é consequência de um baixo esforço amostral.

A descrição original da espécie-tipo, *Corinna rubripes* C. L. Koch, 1842, foi baseada somente em espécimes machos provenientes do estado da Bahia, Brasil. Bonaldo (1996) redescreveu o macho dessa espécie e descreveu a fêmea pela primeira vez. Bonaldo (2000) dividiu as espécies neotropicais do gênero em quatro grupos: grupo *rubripes*, grupo *capito*, grupo *kochi* e grupo *aenea*. O grupo c*apito* inclui as espécies *C.*

capito (Lucas 1857) *e C. colombo* Bonaldo 2000*;* o Grupo *kochi* as espécies *C. kochi* (Simon 1898) e *C. ducke* Bonaldo 2000; e o Grupo *aenea* as espécies *C. aenea* Simon 1896 e *C. recurva* Bonaldo 2000. O grupo *rubripes* atualmente compreende três espécies, *Corinna nitens* (Keyserling 1891) do sul e sudeste do Brasil, norte da Argentina e Paraguai, *C. mourai* Bonaldo 2000 do sul e sudeste do Brasil e a espécie-tipo *C. rubripes* do nordeste do Brasil e Guiana (Bonaldo 1996). Desses, somente o grupo *capito* é claramente monofilético, caracterizado por uma inserção embolar articulada, enquanto nos outros grupos de Corinninae o êmbolo é contínuo ao tégulo (Bonaldo 2000).

O grupo *rubripes* é diagnosticado pela combinação dos seguintes caracteres: carapaça e quelíceras com granulações finas, região cefálica alta com rebaixamento posterior abrupto; fila de olhos posteriores procurva. Palpo do macho com apófise tibial retrolateral robusta, sem processo ventral; tégulo amplo e ovóide com reservatório de orientação elipsóide; processo tegular triangular pouco desenvolvido ou ausente; conductor não estendido prolateralmente, alojando parcialmente ou não o êmbolo; êmbolo fusionado ao tégulo. Fêmeas apresentam uma placa vulvar posterior bem desenvolvida com dobras laterais cobrindo parcialmente as espermatecas primárias e secundárias e fenestras latero-mediana no nível das espermatecas primárias.

Como salientado por Bonaldo (1996), o gênero *Corinna*, serviu por muito tempo como depositário artificial de espécies. Apesar dos esforços de Bonaldo (2000), o gênero *Corinna* ainda não foi revisado e apenas nove espécies (do total de 69 do gênero) foram formalmente incluídas nos grupos de espécies. Assim, a maioria das espécies alocadas atualmente no gênero não foram incluídas nos grupos de espécies propostos e os grupos foram caracterizados apenas através de combinação de caracteres.

Portanto, é preciso que seja feita a revisão taxonômica dos grupos de espécies, definindo-os com base em hipóteses filogenéticas.

Neste trabalho apresentamos uma revisão taxonômica do grupo *rubripes* do gênero *Corinna* re-diagnosticando as três espécies do grupo. Um exame cuidadoso de evidências disponíveis sugere que nenhuma das demais espécies atualmente em *Corinna*, pertencem ao grupo *rubripes*. Entretanto, a diversidade do grupo é grande e são propostas aqui dezessete espécies novas, todas do Brasil. Além disso, novos registros de distribuição do grupo para o Peru e o primeiro registro do gênero para a Bolívia são apresentados.

As espécies deste grupo distribuem-se principalmente pelo sul e sudeste da Mata Atlântica brasileira. Dessas espécies, *Corinna nitens* apresenta a maior distribuição, do norte da Argentina ao nordeste do Brasil, com registros isolados para o Peru, o primeiro registro do grupo para o país. Além disso, um macho e uma fêmea foram registrados para a Bolívia, o primeiro do gênero para o país. Um macho da espéci-tipo, *C. rubripes*, foi registrado para a Guiana por Bonaldo (1996), entretanto esse material não foi examinado neste trabalho. Sete espécies são registradas para o nordeste brasileiro. Entre essas, *C. caatinga* sp. n. foi a única registrada somente na Caatinga, tipo de vegetação exclusiva do Brasil.

O grupo *rubripes* apresenta um conjunto de sinapomorfias putativas, que inclui o palpo do macho com uma apófise tibial retrolateral robusta sem processo ventral (Figs. 47, 50, 63, 78), processo tegular de *Corinna* triangular pouco desenvolvido (Figs. 43, 45, 54, 74, 83, 87) ou ausente (Figs. 90, 94); fêmeas com uma placa vulvar posterior dobrada lateralmente, cobrindo parcialmente as espermatecas primárias com fenestras laterais (Figs. 53, 66, 77, 81, 86, 97, 110). Nos outros grupos de *Corinna* a apófise tibial retrolateral é menos desenvolvida, com ou sem processo ventral. Esse processo,

segundo Bonaldo (2000), provavelmente é uma simplesiomorfia em Corinninae, presente em *Ianduba* Bonaldo, um gênero considerado Corinnidae *insertae sedis*. Assim, a ausência desse processo nas espécies do grupo *rubripes* pode indicar uma perda sinapomórfica, relacionada ou não com a robustez da apófise tibial retrolateral. O processo tegular de *Corinna* é pouco desenvolvido e triangular na maioria das espécies, diferentemente dos outros grupos que apresentam esse processo bem desenvolvido, digitiforme (grupo *capito*) ou virguliforme (grupo *aenea* e *kochi*). Além disso, a posição desse processo pode ser considerada filogeneticamente informativa, pois no grupo *rubripes* geralmente é prolateral, enquanto nos outros grupos esse processo é inserido apicalmente em relação à base do condutor. Entretanto, não está claro se esse processo no grupo *rubripes* representa uma redução de uma estrutura bem desenvolvida. As espécies *C. demersa* sp. n e *C. maracas* sp. n. compartilham a ausência desse processo. As fêmeas do grupo *rubripes* apresentam uma placa vulvar posterior bem desenvolvida cobrindo parcialmente as espermatecas primárias com suas dobras laterais, além das fenestras laterais e da borda posterior dessa placa acima das espermatecas primárias, enquanto os grupos *capito* e *kochi* apresentam essa placa pouco desenvolvida e sua borda fica ao nível da base das espermatecas primárias. No grupo *aenea* essa placa é ausente. Segundo Bonaldo (2000), essa placa esclerotizada é um caráter comum em Corinninae.

Algumas características são promissoras como informação para a definição das relações filogenéticas entre as espécies do grupo *rubripes*, como a presença, em algumas espécies, de um processo mediano na superfície prolateral da apófise tibial retrolateral. Esse processo é pouco evidente em *C. balacobaco* sp. n., *C. telecoteco* sp. n. e *C. ziriguidum* sp. n. Entretanto, em *C. aechmea* sp. n. e *C. jecatatu* sp. n. é comprido e largo, enquanto em *C. zecarioca* sp. n. esse processo é largo, robusto e

subquadrado. A presença dessa estrutura pode suportar um grupo composto pelas espécies citadas acima, enquanto o maior comprimento e largura pode ser considerado uma sinapomorfia para as últimas três espécies. Outra estrutura presente somente em algumas espécies do grupo é uma projeção tegular geralmente retrolateral, mas apical em *C. aechmea* sp. n. Essa projeção é pouco desenvolvida em *C. rubripes* e *C. nitens*, enquanto em *C. maracas* sp. n., *C. vesperata* sp. n. e *C. vilanovae* sp. n. é bem desenvolvida. As espécies *C. demersa* sp. n. e *C. maracas* sp. n. além de compartilharem a ausência de processo tegular, também compartilham uma incisão ventro-apical na apófise tibial retrolateral e uma área não esclerotizada bem delimitada no ápice do condutor. Esses caracteres indicam uma forte relação filogenética entre essas espécies. As espécies *C. vesperata* sp. n. e *C. hyalina* sp. n. compartilham a apófise tibial retrolateral fortemente arrendondada retrolateralmente. A maioria das fêmeas do grupo compartilha um ducto copulatório em forma de "T" invertido, ligando a abertura copulatória média às espermatecas localizadas lateralmente. Na maioria dessas espécies, a abertura é anterior e a placa epiginal não é projetada posteriormente. Entretanto, as espécies *C. rubripes*, *C. nitens* e *C. kuryi* sp. n. apresentam o epígino projetado além do sulco epigástrico e a abertura é posterior, fazendo com que o ducto copulatório não assuma um formato de "T" invertido. Esse caráter é considerado uma sinapomorfia putativa para o grupo formado por essas três espécies.

Dados de história natural referente às espécies aqui abordadas são escassos. Entretato, dados de etiquetas e observações pessoais revelam que algumas espécies pertencem à guilda de caçadoras aéreas ou terrestres (Dias *et. al.* 2010). *Corinna nitens* e *C. mourai* podem ser encontradas em pequenas aberturas de troncos vivos (A.B. Bonaldo observação pessoal). Alguns indivíduos de duas espécies foram coletas em bromélias, *C. demersa* sp. n. e *C. aechmea* sp. n.. Esse tipo de planta apresenta uma

arquitetura que permite criar um microhábitat que fornece às espécies uma área de forrageamento, proteção contra predadores e berçários (Romero & Vasconcellos-Neto 2004, 2005). Espécimes de *C. aechmea* sp. n. foram coletados em bromélias da espécie *Aechmea distinchantha* Lem. em paredões rochosos de arenito, onde essa espécie geralmente ocorre, enquanto que vários indivíduos de *C. demersa* sp. n. foram coletados em bromélias da espécies *Quesnelia arvensis* (Vell.) Mez, sendo que neste caso foi registrado que esta espécie é capaz de mergulhar no reservatório da planta (Piccoli, 2011). Ambas as espécies apresentam uma alta densidade de pelos plumosos no abdômen, os quais poderiam auxiliar na manutenção de uma bolha de ar entre o abdômen e as pernas IV, de modo similar ao registrado em *Argyroneta aquatica* (Clerck, 1757), espécie que vive submersa na água (Ehlers 1939). Entretanto, *C. demersa* sp. n. visivelmente apresenta uma quantidade desse pelos mais significativa que *C. aechmea* sp. n., permitindo essa espécie submergir em fitotelmata de bromélias, provavelmente para escapar de seus predadores (Piccoli, 2011). Entre outros caracteres morfológicos, a presença desses pelos em grande densidade no abdômen de *C. demersa* sp. n. é compartilhado com *C. maracas* sp. n., indicando que possivelmente essa espécie também habita bromélias.

A seguir são apresentados os resultados da revisão taxonômica do grupo *rubripes* do gênero *Corinna*, em formato de artigo científico a ser submetido ao periódico Zootaxa.

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Taxonomic revision of the group *rubripes* of *Corinna* Koch, 1842 (Araneae; Corinnidae)*

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Abstract

The species of the group *rubripes*, which harbors the type species of the genus *Corinna*, are revised, including 20 Neotropical species. Three previously known species were re-diagnosed: *Corinna rubripes* C. L. Koch, 1842, *Corinna nitens* (Keyserling, 1891) and *Corinna mourai* Bonaldo, 2000. New records of *Corinna nitens* are provided, including the first ones from Peru and Bolivia. Seventeen new species, all from Brazil, were described: *C. aechmea* n. sp., *C. balacobaco* n. sp., *C. caatinga* n. sp., *C. demersa* n. sp., *C. escalvada* n. sp., *C. hyalina* n. sp., *C. jecatatu* n. sp., *C. kuryi* n. sp., *C. loiolai* n. sp., *C. maracas* n. sp., *C. regi* n. sp., *C. telecoteco* n. sp., *C. tranquilla* n. sp., *C. vesperata* n. sp., *C. vilanovae* n. sp., *C. zecarioca* n. sp. and *C. ziriguidum* n. sp. A key for all twenty species of the group is presented.

Key words: New species, Neotropical Region.

Introduction

The genus *Corinna* was proposed by C. L. Koch (1842) and was soon transformed into a "dump genus", since most species of Corinninae described in the 19th and 20th centuries were included there. Bonaldo (1996) presented the first modern redescription of the type species *Corinna rubripes* C. L. Koch, proposing a putative synapomorphy for the genus, the presence of a sclerotized conductor. Bonaldo (2000) excluded from *Corinna* several species presenting hyaline conductors, but still the genus is the fourth more diverse in Corinnidae, with 69 valid species (Platnick 2013), 58 of which from the Neotropical region. Bonaldo (2000) indicated that the African species currently allocated in *Corinna* do not belong there. The genus is recorded from

all South American countries, except in Chile, Bolivia and Suriname. These absences, at least in Bolivia and Suriname, are probably undersampling artifacts.

The original description of the type species**,** *Corinna rubripes* C. L. Koch, 1842, was based only on a male specimen from the state of Bahia, Brazil. The description was poorly illustrated, making the recognition of the species impossible without examining the types. Bonaldo (1996) provided illustrations of the male and described the female for first time, but a more general view of the genus diversity was provided by Bonaldo (2000) who proposed four informal species groups defined by combinations of characters, most of them from the male palp: group *rubripes*, group *capito*, group *kochi* and group *aenea*. Bonaldo (2000) included only a few species in each group, without attempting to allocate in these groups all other species he did not transferred to other genera. Besides, he did not propose putative sinampomorphies for three of his species groups, stating that only the group *capito* appeared to be clearly monophyletic since it gathers species with a uniquely articulated embolar insertion, while the embolus is continuous to the tegulum in all other Corinninae.

The group *rubripes* currently includes three species, *Corinna nitens* (Keyserling 1891) from Southeastem and Southem Brazil, Northem Argentina and Paraguay, *C. mourai* Bonaldo 2000, from Southeastem and Southem Brazil and the type species *C. rubripes*, known from Northeastem Brazil and Guyana. In this paper we present a taxonomic review of the group *rubripes*, accessing a larger sample of the group's diversity and rediagnosing the three species studied by Bonaldo (1996, 2000). Carefull examination of the available evidence based in original descriptions, illustrations and type material suggests that none of the species currently in *Corinna*, other than these three species, belongs to the group *rubripes*. However, the richness of the group is greater than expected, especially in the Brazilian Atlantic Forest. As a result, in this

paper we describe seventeen new species, all from Brazil. *Corinna nitens* have a wide distribution, occuring from Northern Argentina to Northeastern Brazil. This species is newly recorded from Peru, which represents the first record of the group *rubripes* for that country. Besides, one male and one female were recorded from Bolivia, representing the first record of the entire genus to that country. Most species described below occurs in Brazilian Atlantic Forest, including six species recorded from Northeastern Brazil, which appear to be related to relicts of this forest formation. However, at least one additional Northeastern species, *C. caatinga* n. sp., is apparently restricted to the semi-arid phytophysionomy called Caatinga.

Additionally, we propose a set of putative synapomorphies for the group *rubripes*, which must be tested in a future formal phylogenetic analysis of the subfamily. The male palp of all species here addressed presents an extremely robust retrolateral tibial apophysis (Figs. 47, 50, 63, 78), which is devoided of a ventral process, and a poorly developed or even absent tegular process of *Corinna* (Figs. 43, 45, 54, 74, 83, 87, 90, 94). In females, the posterior vulval plate is folded, enveloping most of the spermathecae surface, but is fenestrated laterally, revealing partially the primary spermathecae and the fertilization ducts (Figs. 53, 66, 77, 81, 86, 97, 110). In other groups of *Corinna,* the retrolateral tibial apophysis is not as developed as in the group *rubripes* and the ventral process is recognizable in several species (Bonaldo, 2000: figs. 127, 145, 149). According to Bonaldo (2000), the presence of the ventral process is probably a symplesiomorphy for Corinninae, present at least in *Ianduba* Bonaldo, a genus of Corinnidae *insertae sedis*. Thus, its absence in all known species of the group may indicate an instance of synapomorphic loss, connected or not with the event of enlargement of the retrolateral tibial apophysis itself, which also may constitute evidence of the group's monophyly. The tegular process of *Corinna* was hypotethized

by Bonaldo (2000) as a synapomorphy of *Corinna*. In all species groups but the group *rubripes*, this process is well developed, being digitiform (group *capito*) or virguliform (group *aenea* and *kochi*) and is loosely inserted on the tegulum at an apical position. In the group *rubripes*, when present, it is small, triangular, fused to the tegulum and is generally prolaterally located instead of apically inserted in relation to the conductor base. It is not clear at this time whether this state represents a reduction of a well-developed structure, and therefore a putative synapomorphy for the group *rubripes*, or merely a symplesiomorphy shared with other groups, such as *Abapeba* Bonaldo.

According to Bonaldo (2000), most Corinninae present a sclerotized dorsal plate in the vulva. In *Corinna*, this character is extremely variable, from poorly developed (groups *capito* and *kochi*) or even absent (group *aenea*) to sclerotized, well developed in the group *rubripes*. In this group the vulvar plate is uniquely modified, and its lateral folds and latero-median fenestrae are here hypothetized as synapomorphies for the group.

As for the relationships between the species of the group, a few characters from the male palp and female epigynum with potential phylogenetic information can be discussed here. The prolateral surface of the retrolateral tibial apophysis has a median process of which size and shape may be informative. This process is small in *C. balacobaco* n. sp., *C. telecoteco* n. sp. and *C. ziriguidum* n. sp., but in *C. aechmea* n. sp. and *C. jecatatu* n. sp. it is large and long while in *C. zecarioca* n. sp. this process is also large, but stout and sub-squared. The presence of this structure can support a group composed by all species listed above. The two species that share the absence of the tegular process (*C. maracas* n. sp. and *C. demersa* n. sp.) also share a sub-apical notch on the prolateral margin of retrolateral tibial apophysis and a sharply delimited unsclerotized area in the apex of the conductor. These three features indicate a possible

sister-species relationship between these two species. Two additional species, *C. vesperata* n.sp. and *C. hyalina* n. sp., may be united by a unique modification of the retrolateral tibial apophysis, which is almost rounded, inflated retrolaterally. The majority of known females in the group share an inverted "T" shaped copulatory duct, linking the single median copulatory openig to the laterally located pair of spermathecae. In most of these species, the copulatory opening is anteriorly located and the epigynal plate is not projected posteriorly. This configuration is common in those Corinninae with a single copulatory opening. However, in a group of three species (*C. rubripes*, *C. nitens* and *C. kuryi* n. sp.), the epigynum is projected far beyond the epigastric furrow and the copulatory opening is shift posteriorly, causing the copulatory duct to assume a non-inverted "T"shaped configuration (compare figs. 108 and 110). This character is here considered as a putative synapomorphy of the group formed by these three species.

The natural history data for most of the species treated below are scarse or inexistent. Data from labels and reports from collectors indicated that at least some species of the group are trunk-drilling ambushers, belonging to the guild of aereal or ground hunters (Dias *et. al.* 2010). At least *Corinna nitens* and *Corinna mourai* can be found occupying burrows in hardpan bounds or living trunks (A. B. Bonaldo personal observation). However, recent observations of two species here described as new, *C. demersa* n. sp. and *C. aechmea* n. sp., indicated that the natural history of the group is much more diverse. Several individuals of *C. demersa* n. sp. and *C. aechmea* n. sp. were collected inside bromeliads, and at least *C. demersa* n. sp. was reported to be able to submerge into the bromeliad's phytotelmata and stay there for several minutes (Piccoli, 2011). Specimens of *C. aechmea* n. sp. were collected in *Aechmea distinchantha* Lem., on sandstone rock formations in a floodplain in the State of Paraná, while several

individuals of *C. demersa* n. sp. were collected in *Quesnelia arvensis* (Vell.) Mez at Ilha do Cardoso State Park, State of São Paulo. Both species have highly dense covering of feathery hairs, along with long single hairs, on the abdomen, which could represent an adaptation to retain air boobles. The density of such hairs appears to be greater in *C. demersa* n. sp. than in *C. aechmea* n. sp. On the other hand, the presence of highly dense abdominal hair covering is also shared by *C. maracas* n. sp., the putative sister species of *C. demersa* n. sp., indicating that this species could have the same capabilities that were observed in *C. demersa* n. sp.

Material and methods

The material examined belong to the following institutions (acronyms and curators in parentheses): American Museum of Natural History, New York (AMNH, N. I. Platnick); California Academy of Sciences, San Francisco (CAS, C.E. Griswold); Instituto de Biología Neotropical, Tucumán (IBN, P. Globoff); Instituto Butantan, São Paulo (IBSP, D.M. Barros Battesti); Museu de Ciências e Tecnologia da Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre (MCTP, A.A. Lise); Museu Nacional do Rio de Janeiro, Rio de Janeiro (MNRJ, A.B. Kury); Museu Paraense Emílio Goeldi, Belém (MPEG, A.B. Bonaldo); Museu de Zoologia da Universidade de São Paulo, São Paulo (MZSP, R. Pinto da Rocha); Universidade de Brasília, Brasília (UNB, P.C. Motta); Coleção de História Natural de Universidade do Piauí, Piauí (CHNUFPI, L.S. Carvalho); Museu de Ciências Naturais, Fundação Zoobotânica do Rio Grande do Sul, Porto Alegre (MCN, R. Otti); Museu de História Natural "Capão da Imbuia", Curitiba (MHCI, M. Arzua); Universidade Federal de Minas Gerais, Belo Horizonte (UFMG, A.J. Santos); Staatliches Museum für Naturkunde, Karlsruhe

(SMNK, H. Höfer); Museo de Historia Natural, Universidad Nacional de San Marcos, Lima (MUSM, D. Silva).

The material was examined submersed in alcohol 80%, on a ZEISS Discovery V8 stereomicroscope. Measurements are expressed in millimeters and were obtained with the use of an ocular with micrometric ruler. The format of the descriptions and morphological terms follows Bonaldo (2000). All illustrations were made in the same equipment, with the aid of a *camera lucida*. The left palps of males were illustrated in ventral and retrolateral positions. Illustrations of female genitalia were made in ventral and dorsal positions. For visualization of internal structures of the female genitalia, the epigynum was detached from the body, mecanically cleaned with pins and immersed in clove oil (Levi 1965). The number and arrangement of spines were expressed according Petrunkevitch (1925). Scanning eletron micrographs were taken with a Zeiss Leo 1450 VP SEM from the Laboratório Institucional de Microscopia Eletrônica de Varredura (MPEG) and a Zeiss Leo 1430 from the Laboratório de Microscopia Eletrônica de Varredura from the Instituto de Geociências (UFPA). Distribution maps were made for all species, using the Geographic Information System (GIS) software program Quantum Gis version 1.8.0. Records without coordinates were georeferenced approximatively (for example, only municipalities) through Google Earth. All geographic information not extracted from vial labels was listed in brackets. Abbreviations used in the text followed Bonaldo (1996, 2000) and Santos-Souza & Bonaldo (2007) and were translated into English when necessary: **ALE**—Anterior lateral eyes; **AME**—Anterior median eyes; **C—**Conductor; **CD**—Copulatory ducts **CO**—Copulatory opening; **Cpr**—Cymbial process; **d—**Dorsal; **E—**Embolus; **FD**—Fertilization ducts; **FR1—**First fold of reservoir; **MOQ**—Median Ocular Quadrangle; **p—**Prolateral; **PLE**—Posterior lateral eyes; **PME**—Posterior median eyes; **ppRTA—**Prolateral process of Retrolateral tibial

apophysis; **PS**—Primary spermathecae; **PVP—**Posterior vulvar plate; **r—**Retrolateral; **RTA—**Retrolateral tibial apophysis; **SS**—Secondary spermathecae; **T—**Tegulum; **TPC—**Tegular process of *Corinna*; **TPr—**Tegular projection; **v—**Ventral; **vp—**Ventral prolateral; **vr**—Ventral retrolateral.

Taxonomy

Corinna C. L. Koch, 1842

Corinna C. L. Koch, 1842: 17. Type species by original designation, *Corinna rubripes* C. L. Koch, 1842; Bonaldo, 2000: 37; Bosselaers & Jocqué, 2002: 250.

Diestus Simon, 1898: 199. Type species by original designation, *Diestus kochi* Simon, 1898; Synonymized with *Corinna* by Bonaldo, 2000: 37.

Lausus Simon, 1898: 199. Type species by original designation, *Corinna aenea* Simon, 1896; Synonymized with *Corinna* by Bonaldo, 2000: 37.

Tranquilinus Mello-Leitão, 1915:140. Type species by original designation and monotypy, *Tranquilinus benefaciens* Mello-Leitão, 1915; Synonymized with *Corinna* by Mello-Leitão, 1925: 445.

Diagnosis and Description. See Bonaldo (2000).

Note. Bonaldo (1996, 2000) proposed a single putative synapomorphy for the entire genus, the sclerotized conductor in the male palp. This hypothesis was not put to phylogenetic test yet. While there is little dispute that this condition really reffers to a modification of the hyaline conductor, it is not clear if this state is the result of a single

event or if the various shapes of the sclerotized conductor across the *Corinna* species groups represent different instances of transformation. The solution to this problem requires a formal phylogenetic analysis that may lead to the redefinition of the genus to include fewer groups of species or even only the species here addressed.

Group *rubripes*

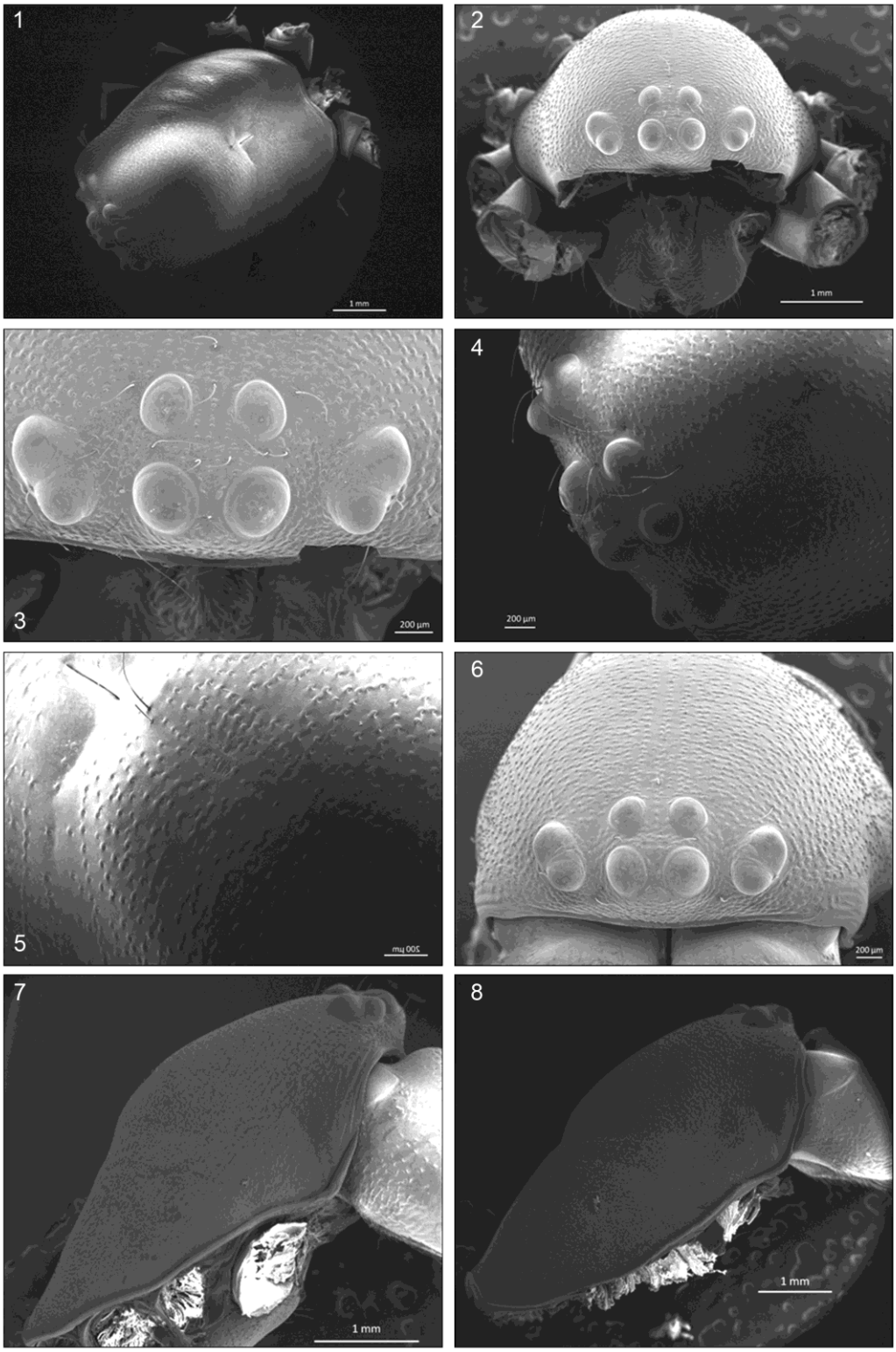
Diagnosis. Species of the group *rubripes* of *Corinna* can be recognized by the finely granulated carapace and chelicerae (Figs. 4–6, 9–10), high cephalic region, abruptly depressed posteriorly (Figs. 7–8, 37–40); posterior eye row procurved (Figs. 2–3, 6). Male palp with retrolateral tibial apophysis robust, without ventral process (Figs. 47, 50, 68, 74, 99); tegulum wide and ovoid (Figs. 45, 54, 78, 87); reservoir with ellipsoid orientation (Figs. 43, 58, 63, 90); tegular process triangular, poorly developed or absent (Figs. 43, 54, 74, 83, 94); conductor not extended prolaterally, generally accommodating the distal third of the embolus (Figs. 47, 58, 69, 90); embolus fused to tegulum (Figs. 45, 63, 70, 94). Posterior vulval plate well developed, with lateral folds embracing the primary and secondary spermatecae and latero-median fenestrae at the level of primary spermatecae (Figs. 53, 66, 77, 81, 86, 97, 110).

Description. Total length (males and females) 7.25–16.5. Carapace suboval in dorsal view, with thin granulations (Figs. 1, 4–5, 33–36), longer than wide, widest at coxae II, cephalic area high (Figs. 7–8, 37–40), well delimited, specially in *C. caatinga* (Fig. 40); thoracic area with abrupt posterior depression, thoracic groove short; clypeus relatively low (variation 0.27–0.65); posterior and anterior eye rows procuverd in frontal view (Figs. 2–3, 6); MOQ wider than long (*C. regi* n. sp. and *C. ziriguidum* n. sp., longer than wide); eyes circular (Figs. 3, 6), AME slightly larger than others. Interdistances: AME–

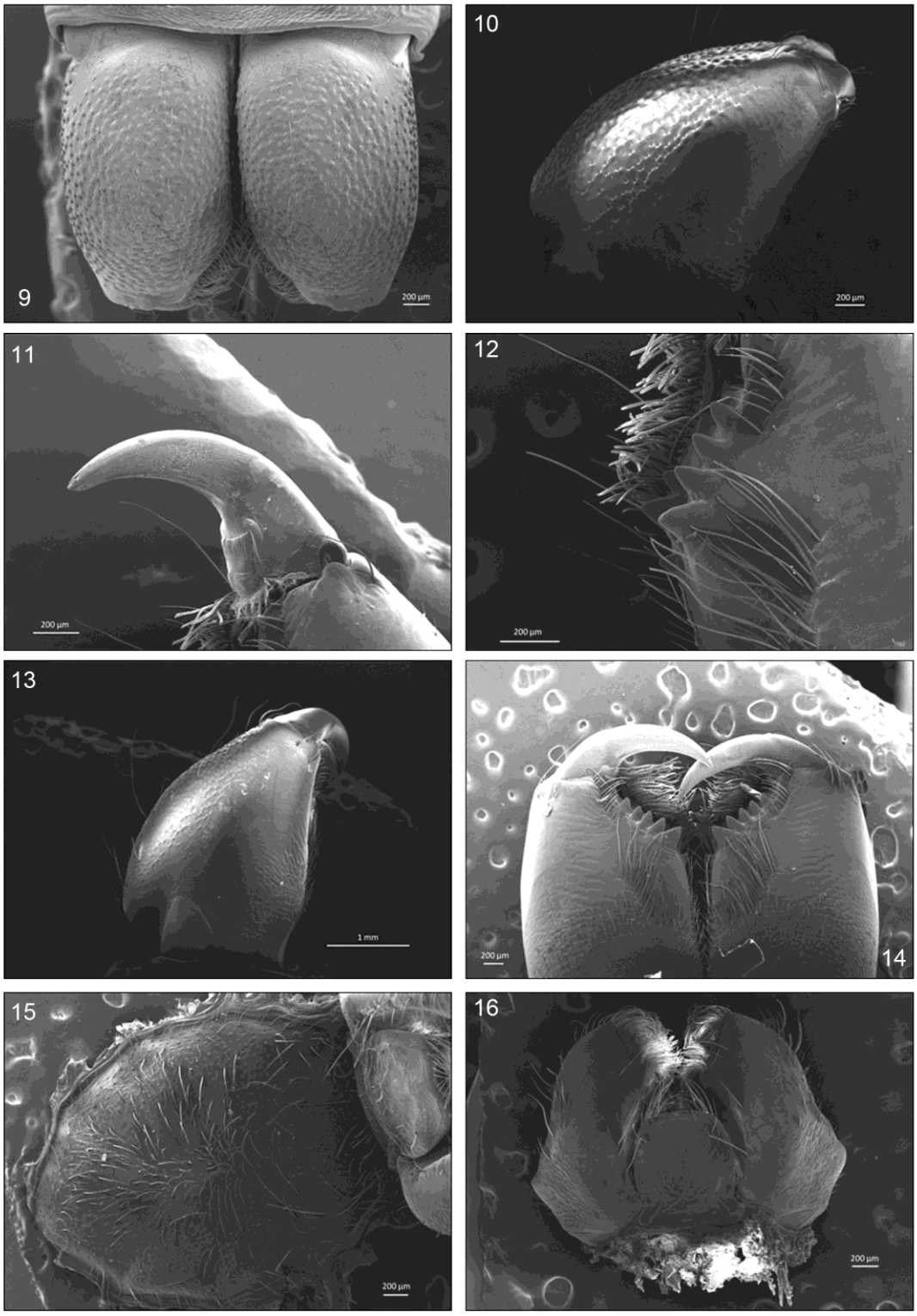
AME separated by approximately an AME diameter; AME–ALE by one to two AME diameter (in *C. loiolai* n. sp. separated by more two times the AME diameter); PME–PME by one to two PME diameter (females of *C. balacobaco* n. sp. and *C. caatinga* n. sp. separated by more two times the PME diameter); PME–PLE by two to four PME diameter; ALE–PLE separated by approximately a PLE diameter. Chilum glabrous and entire (reduced in *C. tranquilla* n. sp.). Chelicerae voluminous, strongly geniculate, with thin granulations and conspicuous basal condylus (Figs. 9–10, 13, 37–40); cheliceral promargin with 3 teeth, retromargin mostly with 4 teeth (Fig. 12), *C. demersa* n. sp., *C. maracas* n. sp. and *C. loiolai* n. sp. with 5 teeth (Fig. 14) and *C. caatinga* n. sp. with 6; fang strong (Fig. 11), cheliceral length approximately equal to height of carapace (*C. caatinga* sp. n., females of *C. zecarioca* n. sp and males of *C. maracas* n. sp., approximately twice times or more the carapace height). Endites convergent, promargin concave, retromargin with discrete internal excavation (Fig. 16), serrula in single row; labium longer than wide, with laterally projected posterior margin (Fig. 16); sternum longer than wide, entirely rebordered, especially on anterior margin (Fig. 15). Legs long, Leg formula variable, generally I, IV, II, III; in *C. rubripes*, *C. demersa* n. sp., *C. maracas* n. sp., *C. caatinga* n. sp., *C. tranquilla* n. sp., males of *C. vesperata* n. sp., females of *C. jecatatu* n. sp. and *C. mourai*, IV, I, II, III; femur I and II generally with 2 ventral spines and 1 prolateral; metatarsus I and II generally with 2 pairs of ventral spines; dense scopula on all tarsi (Figs. 23, 27) and metatarsi I and II (covering the entire segment, the segment distal half or distal third); distal metatarsi III and IV with ventral dense clusters of stout hairs (Fig. 24); claws pectinate, with 12–13 teeth (Fig. 28); claw tufts dense (Figs. 26–27); thricobothria irregularly distributed on dorsal surface of tarsi (Figs. 17, 21–22), thricobothrial base rebordered (Figs. 18–19); tarsal organ subapical, capsulated, with rounded aperture (Fig. 20); female palpal tarsus with

one pectinate claw (Fig. 25). Abdomen generally oval, with long sparse simple and feathery hairs, except in *C. aechmea* n. sp., *C. demersa* n. sp. and *C. maracas* n. sp. with highly dense hair covering (Figs. 29–32); male dorsal scutum placed on anterior distal third or anterior half of abdomen, generally elongate (Figs. 33–36); scutum absent in females; tracheal tubercle absent; colulus inconspicuous, with few hairs.

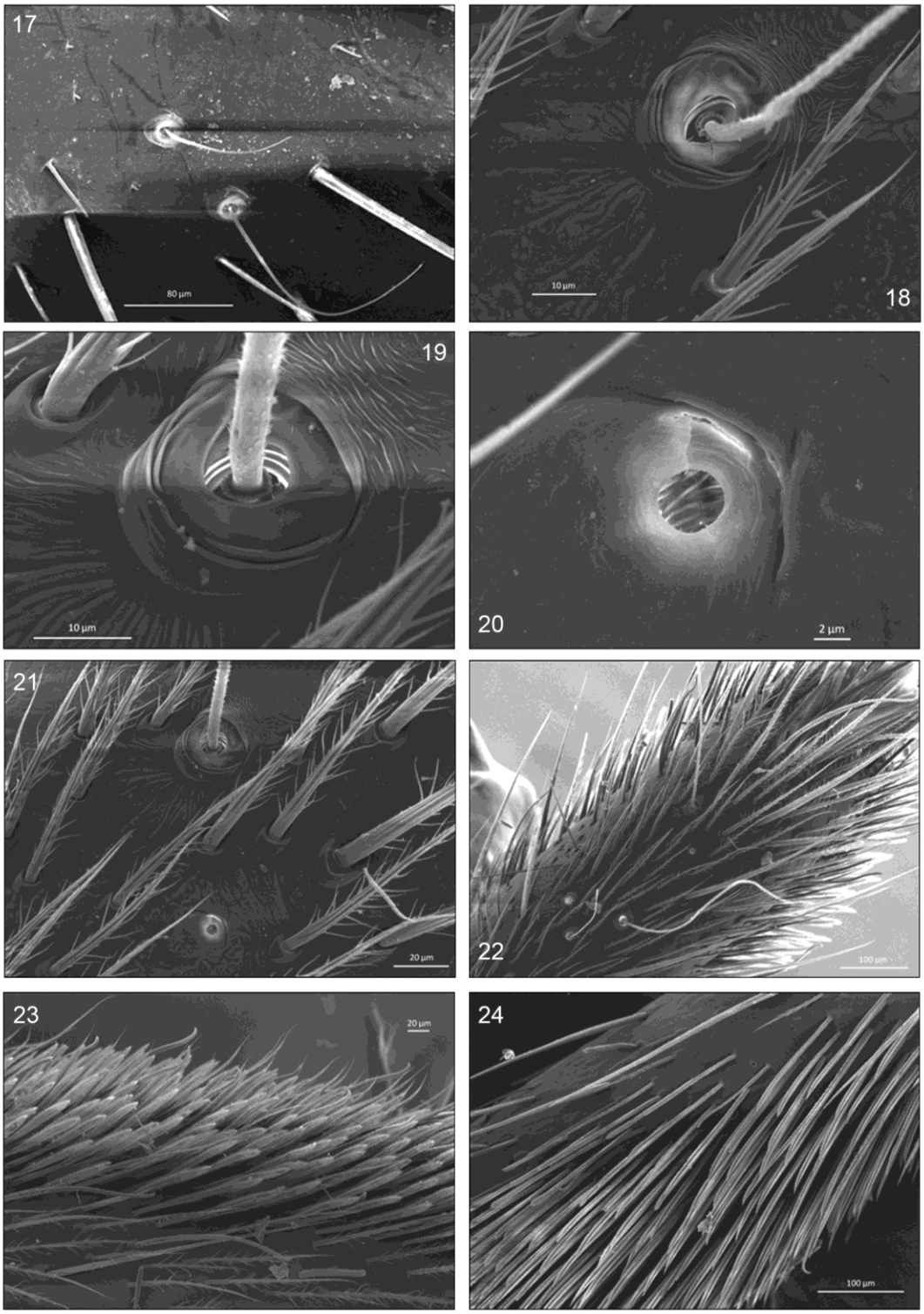
Male palp: femur unmodified, generally with two posterior dorsal spines (*C. jecatatu* n. sp. with one; *C. balacobaco* n. sp. and *C. zacarioca* n. sp. with three spines). Patella unmodified. Tibia with single RTA, robust, without ventral process (Figs. 47, 54, 63, 78, 90); ppRTA small in *C. telecoteco* n. sp., *C. ziriguidum* n. sp. and *C. balacobaco* n. sp. (Figs. 43, 45, 83), large in *C. aechmea* n. sp., *C. jecatatu* n. sp. and *C. zecarioca* n. sp. (Figs. 50, 54, 58); *C. demersa* n. sp. and *C. maracas* n. sp. with ventro-apical incision (Figs. 91, 95); *C. vesperata* n. sp. and *C. hyalina* n. sp. with RTA retrolaterally enlarged (Figs. 75, 79). Cymbium with distal dorsal scopulae; in *C. caatinga* n. sp., cymbium with a large spurn-like basal dorsal projection, tapering towards the dorsum of tibia (Figs. 64, 68). T wide and ovoid, reservoir with ellipsoid orientation, with four folds visible ventrally (Figs. 47, 50, 63, 78); TPC triangular, fused to tegulum, pooly developed, generally inserted prolaterally (Figs. 43, 58, 74, 87), except in *C. caatinga* n. sp. inserted retrolaterally (Figs. 63, 70), absent in *C. demersa* n. sp. and *C. maracas* n. sp. (Figs. 90, 94); TPr inserted retroapically in *C. vesperata* n. sp., *C. vilanovae* n. sp. and *C. maracas* n. sp. (Figs. 74, 87, 94), inserted apically in *C. aechmea* n. sp. (Fig. 50). C sclerotized, not extended prolaterally, with retrolateral margin forming a groove which accommodates the distal third of embolus (Figs. 47, 58, 69, 90); groove absent in *C. vilanovae* n. sp. and *C. maracas* n. sp. (Figs. 87, 94). E fused to tegulum, curved retrolaterally, generally filiform (Figs. 45, 63, 78, 90); in *C. vilanovae* n. sp., flattened, with wide tip (Fig. 87).



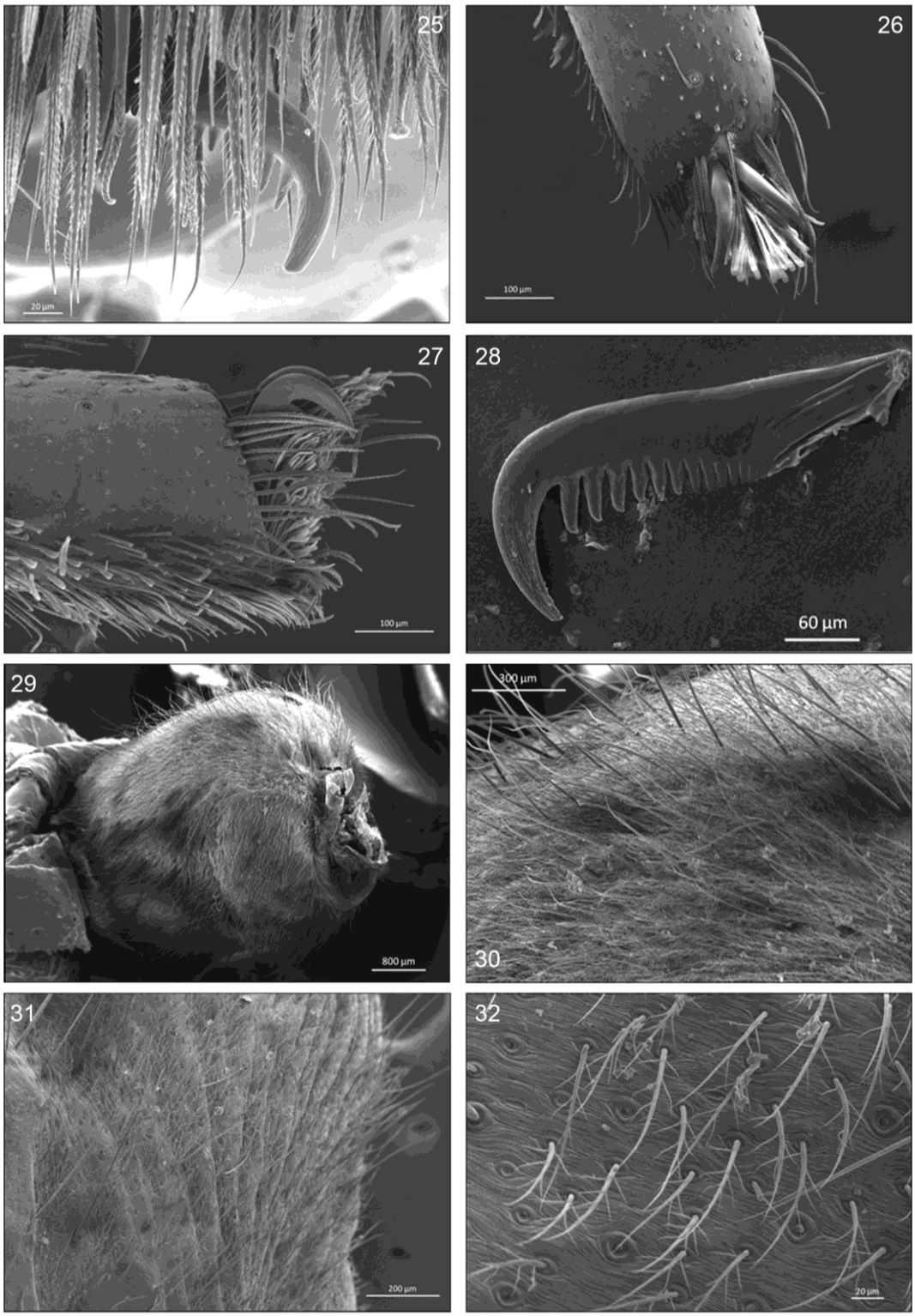
FIGURES 1–8. *Corinna aechmea* n. sp., male, carapace: 1) dorsal; 2) frontal; 3) eyes, frontal; 4) eyes, dorsal; 5) cephalic region granulations, dorsal. *Corinna ziriguidum* n. sp., male, carapace: 6) frontal 7) lateral. *Corinna maracas* n. sp., female, carapace: 8) lateral.



FIGURES 9–16. *Corinna ziriguidum* n. sp., male: 9) chelicerae, frontal; 10) chelicerae, lateral. *Corinna aechmea* n. sp., male: 11) chelicerae, fang; 12) chelicerae, teeth. *Corinna maracas* n. sp., female: 13) chelicerae, lateral; 14) chelicerae, ventral; 15) sternum, ventral. *Corinna ziriguidum* n. sp., male: 16) endites and labium, ventral.



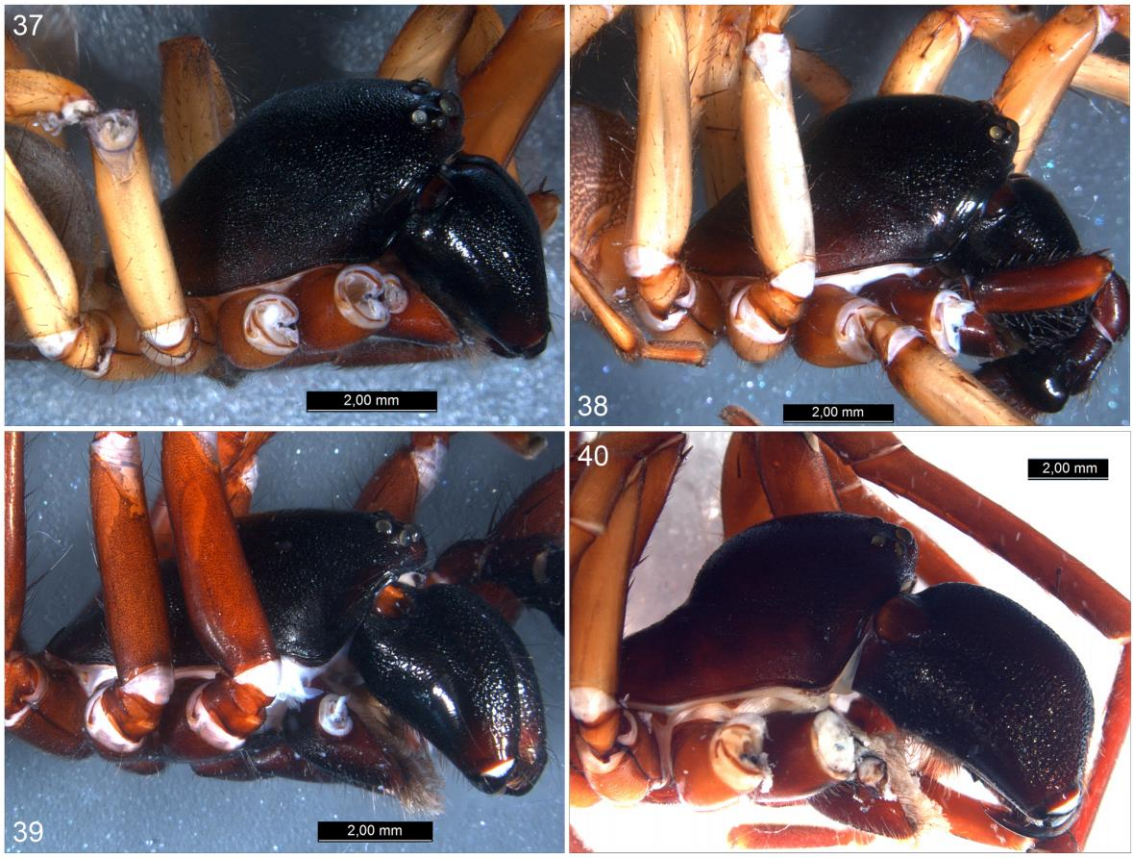
FIGURES 17–24. *Corinna demersa* n. sp., male: 17) thricobothria on RTA, retrolateral. *Corinna aechmea* n. sp., male, tarsus I: 18) thricobothria; 19) thricobothria base; 20) tarsal organ; 21) thricobothria and tarsal organ; 22) thricobotrhial cluster; 23) scopula. *Corinna ziriguidum* n. sp., male, metatarsus IV: 24) ventral distal stout hairs cluster.



FIGURES 25–32. *Corinna maracas* n. sp., female: 25) palpal claw. *Corinna ziriguidum* n. sp., male, tarsus I: 26) claws, dorsal; 27) claws, lateral. *Corinna demersa* n. sp., female: 28) tarsus I, claw, lateral; 29) abdomen, lateral; 30) abdominal hairs, lateral. *Corinna aechmea* n. sp., male, abdomen 31) lateral hairs; 32) feathery hairs.

Epigynum: ventral plate generally not projected posteriorly (Figs. 56, 60, 76, 85, 96, 105); moderatelly projected in *C. rubripes* and *C. kuri* n. sp. (Fig. 109), strongly projected in *C. nitens* and *C. caatinga* n. sp. (Fig. 65). CO generally median (Figs. 52, 80, 107), anteriorly placed in *C. demersa* n. sp. and *C. maracas* n. sp. (Figs. 92, 96, 103), posteriorly placed in *C. rubripes*, *C. nitens* and *C. kuryi* n. sp. (Fig. 109). CD ventrally visible in *C. rubripes*, *C. demersa* n. sp. and *C. caatinga* n. sp. (Figs. 65, 92). SS ventrally visible, except in *C. caatinga* n. sp. (Fig. 65). PS and SS partially covered by PVP (Figs. 53, 77, 86, 106), in *C. caatinga* n. sp. PVP covers only primary spermathecae (Fig. 66); PVP well developed. SS generally globular, with unsclerotized apex; ovoid in *C. tranquilla* n. sp. (Fig. 81) and in irregular *C. caatinga* n. sp. (Fig. 66); PS globular, smalller than SS, dorsally visible through PVP fenestrae (Figs. 61, 77, 81, 108). FD small, generally curved inward toward anterior end of epigynum (Figs. 53, 77, 97, 110).

FIGURES 33–36. *Corinna* spp., males, dorsal view: 33) *Corinna aechmea* n. sp.; 34) *Corinna escalvada* n. sp.; 35) *Corinna telecoteco* n. sp.; 36) *Corinna caatinga* n. sp.



FIGURES 37–40. *Corinna* spp., males, carapace and chelicerae, lateral view: 37) *Corinna balacobaco* n. sp.; 38) *Corinna vesperata* n. sp.; 39) *Corinna demersa* n. sp.; 40) *Corinna caatinga* n. sp.

Key to species of the group *rubripes*

1 – Males.....2

- Females 17

2(1) – Tegular process absent (Figs. 90, 94, 101, 102); RTA with a ventro-apical incision (Figs. 91, 95, 99) 3

- Tegular process present (Figs. 43, 63, 70, 78), RTA otherwise 4

3(2) – Tegular apical margin with a pointed retrolateral projection; embolus not covered by conductor (Fig. 94) ..... *C. maracas* **n. sp.**

- Tegular apical margin without retrolateral projection; embolus partially covered by an apical groove of conductor (Figs. 90, 102) .....*C. demersa* **n. sp.**

4(2) – Cymbium with tapered basal projection (Fig. 64, 68)..... *C. caatinga* **n. sp.**

- Cymbial projection absent 5

5(4) – Prolateral surface of RTA with a median projection (Figs. 43, 45, 50, 54, 58, 83). 6

- RTA without such projection.....11

6(5) – RTA median projection large (Figs. 50, 54, 58). 7

- RTA median projection small (Figs. 43, 45, 83). 9

7(6) – Tegulum with a rounded apical projection (Fig. 50).....*C. aechmea* **n. sp.**

- Tegulum otherwise. 8

8(7) – Median process in the prolateral surface of RTA stout, sub-squared (Fig. 59); tegular process close to embolar base (Fig. 58). .....*C. zecarioca* **n. sp.**

- Median process in the prolateral surface of RTA longer than wide (Fig. 55); tegular process far from embolar base (Fig. 54)……..*C. jecatatu* **n. sp.**

9(6) – Embolus short, bent in straight angle (Fig. 83)….....*C. balacobaco* **n. sp.**

- Embolus longer, gently bent 10

10(9) – Tegular process curved prolaterally, FR1 restricted to midlle of tegulum (Fig. 43) .....*C. telecoteco* **n. sp.**

- Tegular process directed apically, FR1 large, expanded to retro-apical sector of tegulum (Fig. 45) .....*C.ziriguidum* **n. sp.**

11(5) – Apex of conductor extend retrolaterally (Bonaldo, 1996: fig. 16; 2000: fig. 119) 12

- Apex of conductor not extend retrolaterally 13

12(11) – Apex of RTA pointed, conductor with a quadrangular basal process (Bonaldo, 1996: fig. 16).....*C. rubripes*

- Apex of RTA obtuse, conductor without basal process (Bonaldo 2000: fig. 119) .....*C. nitens*

13(11) – Embolus stout and spatulated, not covered by conductor (Fig. 87) .....*C. vilanovae* **n. sp.**

- Embolus filiform, partially cover by groove of conductor 14

14(13) – RTA enlarged retrolaterally (Figs. 75, 79). 15

- RTA otherwise.....16

15(14) – Apex of RTA not projected ventrally, with small hyaline process, tegular process inseted closely to embolar base (Figs. 78, 79) ..... *C. hyalina* **n. sp.**

- Apex of RTA strongly projected ventrally, without hyaline process (Fig. 75); tegular process inseted far from embolar base (Fig. 74). .....*C. vesperata* **n. sp.**

16(14) – Apex RTA rounded, directed prolaterally (Bonaldo, 2000: fig. 123).....*C. mourai*

- Apex of RTA pointed, directed apically (Fig. 47). ..... *C. escalvada* **n. sp.**

17(1) – Epigynal plate projected posteriolly, beyond epigastric groove (Figs. 65, 109)..... 18

- Epigynal plate not projected posteriolly. 21

18(17) – Copulatory opening medially placed (Fig. 65)..... *C. caatinga* **n. sp.**

- Copulatory opening posteriorly placed.....19

19(18) – Copulatory ducts visible ventrally (Bonaldo, 1996: fig. 16).....*C. rubripes*

- Copulatory ducts not visible ventrally 20

20(19) – Copulatory opening placed in a small posterior atrium; posterior epigynal margin medially rounded (Bonaldo, 2000: fig. 121) .....*C. nitens*

-Copulatory opening placed in undifferentiated area; posterior epigynal margin medially truncated (Fig. 109).....*C. kuryi* **n. sp.**

21(17) – Copulatory opening inserted in a ventral triangular epigynal projection (Fig. 85) .....*C. balacobaco* **n. sp.**

- Triangular epigynal projection absent.....22

22(21) – Copulatory opening small, "v"-shaped and anteriorly placed, in the level of secondary spermatecae (Figs. 92, 96, 103) 23

- Copulatory opening otherwise (Figs. 52, 56, 60, 76, 80, 105, 107)..... 24

23(22) – Copulatory ducts long, inverted T-shaped and visible ventrally, epigynal surface not grooved (Fig. 92).....*C. demersa* **n. sp.**

- Copulatory ducts not visible ventrally, epigynal surface with wide longitudinal median groove (Fig. 96)..... *C. maracas* **n. sp.**

24(22) – Copulatory opening defined both anteriorly and posteriorly by well defined margins (Figs. 105, 107) 25

- Copulatory opening defined only posteriorly by a well defined margin 26

25(24) – Secondary spermatecae separated from each other (Fig. 106); posterior margin of copulatory opening nearly straight (Fig. 105) .....*C. loiolai* **n. sp.**

- Secondary spermatecae touching each other (Fig. 108); posterior margin of copulatory opening procurve (Fig. 107) .....*C. regi* **n. sp.**

26(24) – Posterior margin of copulatory opening large, "V"- shaped (Figs. 76) .....*C. vesperata* **n. sp.**

- Posterior margin of copulatory opening otherwise.....27

27(26) – Secondary spermatecae separated from each other apically by more than one diameter (Figs. 61, 81) 28

- Secondary spermatecae touching each other (Bonaldo, 2000: fig. 125) or separated from each other by less than one diameter (Figs. 53, 57) 29

28(27) – Secondary spermatecae globular (Fig. 60, 61) .....*C. zecarioca* **n. sp.**

- Secondary spermatecae oblong (Fig. 80, 81) ..... *C. tranquilla* **n. sp.**

29(27) – Secondary spermatecae touching each other .....*C. mourai*

- Secondary spermatecae separated from each other 30

30(29) – Posterior margin of copulatory opening "u"- shaped (Fig. 52); PVP with lateral rounded protuberances in the internal margin (Fig. 53) .....*C. aechmea* **n. sp.**

- Posterior margin of copulatory opening straigth (Fig. 56); PVP without lateral protuberances (Fig. 57) ..... *C. jecatatu* **n. sp.**

***Corinna rubripes* C. L. Koch, 1842**

Fig. 41

Corinna rubripes C. L. Koch, 1842: 17, pr. 293, fig. 702 (male holotype from Bahia, Brazil, Gomez col., ZMB 2134, not re-examined); Karsch, 1880: 375, est. 12, fig. 1; Simon, 1898: 198; Petrunkevitch, 1911: 469; 1928: 177; Roewer, 1954: 600; Bonnet, 1956: 1216; Moritz & Fischer, 1988: 137; Bonaldo, 1996: 80 (figs. 1–19); Bonaldo, 2000: 40 (figs. 21, 22).

Sparassus rubripes; Walckenaer, 1847: 561.

Diagnosis. Males of *Corinna rubripes* resemble those of *C. nitens* by the tip of conductor extend retrolaterally but differ by the apex of RTA projected ventrally, by a large proapical projection hidden the embolar base in ventral view and base of conductor with a quadrangular projection (Bonaldo, 1996: figs. 16–17); females resemble *C. nitens* and *C. kuryi* n. sp. by the copulatory opening posteriorly placed but

can be recognized by the copulatory duct visible ventrally and epigynal plate only slightly projected posteriorly (Bonaldo, 1996: figs. 18–19).

Description. See Bonaldo, 1996: figs. 1–19.

Distribution. Guyana and northeastern Brazil.

New records: BRAZIL**. Bahia**: Jequié (Brejo Novo) [13º56'41"S 40º06'33.9"W], 1 female, 6-7.VI.2009, Chagas Jr, A. Kury, D. Pedroso, A. Giupponi and V. Dill (MNRJ 6123); Uruçuca (Fazenda Santa Tereza), 2 males, 3.VI.1970 (MNRJ 13192); (Fazenda Santo Antonio), 1 female, 24.X.1978, J.S. Santos (CPDC-4225); Ilhéus (Reserva Zoobotânica) 14°46'22.07"S 39° 13'13.8"W, 2 males, 8-9.XII.2010, G.H.F. Azevedo and A.J. Santos (UFMG 9442); Camacan (RPPN Serra Bonita) [15°23'25" S 39°34'05"W], 1 female, 11-13.VI.2009, Chagas Jr, A. Kury, D. Pedroso, A. Giupponi and V. Dill (MNRJ 6120); (Fazenda Esperança), 1 male, 18.VI.1969 (MNRJ 13202); Itapebi [15°57'9.18"S 39°32'1.69"W], 1 female, T. Bernabé (MNRJ 6234); Porto Seguro [16°27'4.04"S 39° 3'52.79"W], 1 female (MNRJ 6444); (Fazenda Nossa Senhora da Conceição), 1 female, 21.III.1971 (MNRJ 1899); Itamarajú (Fazenda Pau Brasil) [17°04'S 39°31'W], 1 male, 8.VII.1968 (MNRJ 3304); 1 female, 10.I.1969 (MNRJ 13237); **Minas Gerais**: Águas Vermelhas [15°44'50.92"S 41°27'39.37"W], 1 male (MNRJ 2051); Marliéria (Parque Estadual do Rio Doce) 19°42'58.62" S 42°44'1.69" W, 1 male, 5-6.I.2010, W. H. Thomassen et. al. (UFMG 7512).

***Corinna nitens* (Keyserling, 1891)**

Fig. 41

Hypsinotus nitens Keyserling, 1891: 57, est. 2, fig. 30 (lectotype and paralectotype from Blumenau, Santa Catarina, Brazil, BMNH; not re-examined).

Corinna nitens; Simon, 1897: 192; Petrunkevitch, 1911: 468; Mello-Leitão, 1923: 54; 1927: 398; Roewer, 1954: 598; Bonnet, 1956; 1214. Bonaldo, 2000 (figs. 119–122): 41; Bosselaers & Jocqué, 2002: 250.

Diestus altifrons Mello-Leitão, 1945: 260 (holotype from Pindapoy, Misiones, Argentina, II.1942, Bridarolli col., MLP 16575, not re-examined); Roewer, 1954: 602; Arrozpide, 1986: 17. Synonymized with *Corinna nitens* by Bonaldo, 2000.

Diagnosis. Males of *Corinna nitens* differs from those of *C. rubripes* by the obtuse apex of RTA and by the absence of both the proapical tegular projection and the quadrangular process in conductor base (Bonaldo, 2000: figs. 119–120); females differ from those of *C. rubripes* and *C. kuryi* n. sp. by the epigynal plate extremely projected posteriorly, copulatory ducts not visible ventrally and copulatory opening placed in a small posterior atrium (Bonaldo, 2000: figs. 121–122).

Description. See Bonaldo, 2000: figs. 13, 27, 33, 57, 67, 74, 77–82, 89, 119–122, 153.

Distribuition. Peru, Bolívia, Paraguay, Northern Argentina, Southeastern and Southern Brazil.

New records: BRAZIL. **Góias:** Cidade Ocidental [16° 6'51.79"S 47°56'13.75"W] (área rural), 1 female, 20.III.2001, L. Fonseca (UNB 1110); Catalão [18° 9'40.37"S 47°56'41.61"W], 1 male and 2 females, Blaser (UNB 690); **Distrito Federal:** Brasília [15°49'36.09"S 47°55'18.55"W], 1 male, 1.X.2002, D. Diniz (UNB 1889); 2 males, a 20.III.2001 (UNB 1112); 1 male, 14.XI.2001 (UNB 1553); (Jardim Botânico) [15°52'45.92"S 47°50'32.16"W], 1 male, 19.VIII.2002, D. Diniz (UNB 1924); 1 male, 20.IV.2002, P. C. Motta (UNB 1885); 1 male, 12.X.2001, D. Diniz, F. S. P. Godoi (UNB 1376); (Fazenda Água Limpa) [15°56'45"S 47°56'8"W], 1 female, 22.V.1998,

P. C. Motta and R. Bertani (UNB 326); 1 male, 25.XI.1995, P. C. Motta (UNB 134); **Minas Gerais**: Morro do Pilar [19°12'49.12"S 43°23'8.38"W], 1 female, 10.IV.2011, P. Henrique (UFMG 7807); Santa Bárbara (RPPN Santuário do Caraça, Pico do Sol), 20°03'31.85"S 43°30'19.72"W, 1 male, 7.XI.2009, L.N. Perillo (UFMG 6651); Simonésia (RPPN Estação Biológica da Mata do Sossego) 20°04'25"S 42°04'13"W, 1 male, 28-30.XI.2010, M.T.T. Santos (UFMG 4741); Viçosa [20°45'15.32"S 42°52'56.55"W] (Escola Superior de Agricultura de Viçosa), 5 females, 21.II.1938, 21.II.1938 (MNRJ 3123); Tiradentes (Serra de São José) [21° 6'11.79"S 44°10'44.10"W], 1 female, 22.VIII.1990, R.J.V. Alves (MNRJ); Lavras (Escola Superior de Agricultura de Lavras), 1 female, 25.II.1993, R.L.C. Baptista (MNRJ 3125); Juiz de Fora (Reserva Biológica Municipal Poço das Antas) 21°45'37.8"S 43°19'10.2"W, 2–3.IV.2011, G.H.F. Azevedo and A.J. Santos (UFMG 5293); Itajúba (Horto Florestal Anhumas) 22°25'36"S 45° 28'10"W, 1 male, I.2010, C.G.G. Castro (UFMG 4254); 1 male, IV.2010 (UFMG 4255); Rio Preto [22°48'0.00"S 45°45'60.00"W], 3 males and 2 females, 12–20.V.2002, Exp. Arachné (MNRJ 6442); **Espírito Santo:** Santa Teresa (Estação Biológica de Santa Lúcia) [19º 56' 10" S 40º 36' 06" W], 1 female, 29–31.VII.2009, Equipe Myriapodologia GS (MNRJ 6008); (Museu Mello-Leitão) [19°56'10.92"S 40°36'0.39"W], 2 males, 14–18.V.2004, A. Giupponi (MNRJ 6432); **Rio de Janeiro**: Santa Maria Madalena (Parque Estadual do Desengano, Morumbecá) [21°53'34.41"S 41°54'48.31"W], 1 female, 13–17.V.2008, A. Kury, C. Sampaio and T. Moreira (MNRJ 13962); Nova Friburgo (Amparo) [22°15'8"S 42°27'36"W], 1 female, 20.XII.1993, A. Kury (MNRJ 13223); Itatiaia [22°29'44.63"S 44°33'39.41"W], 1 female, 15–17.XI.2007, C. Mello-Patir (MNRJ 6450); 1 male (MNRJ 6434); (Parque Nacional do Itatiaia) [22°27'17"S 44°36'29.08"W], 1 female, 15–22.II.2011, G.H.F. Azevedo et. al. (UFMG 9700); 1 male (UFMG 9701);

Teresópolis (Parque Nacional da Serra dos Orgãos) [22°27'23.4"S 42°59'41.4"W], 1 male, X.2006, Exp. Arachné (MNRJ 6439); Jacarepaguá (Represa do Cigano) [22°56'24.84"S 43°17'55.12"W], 1 female, 12.VII.89, A. Kury (MNRJ 6433); Rio de Janeiro (Parque Nacional da Tijuca) [22°57'40.93"S 43°16'30.83"W], 1 female, 20.I.2001, R.L.C. Baptista (MNRJ 3943); 1 female, XI.1985 (MNRJ 3301); (Parque Nacional da Tijuca, Serra da Carioca) [22°57'38.79"S 43°14'45.25"W], 1 male, 18.XI.2001, E.H. Wienskoski (MNRJ 6448); (Parque Nacional da Tijuca, Mesa do Imperador) [22°58'13"S 43°15'28"W], 1 male, 9.V.1988, R.L.C. Baptista (MNRJ 3297); Niterói (Itaipú) [22°56'59.19"S 43° 2'2.62"W], 1 male, 9.X.1996, C.C. Ratto (MNRJ 6445); **São Paulo:** Botucatu [22°53'53"S 48°29'29"W], 1 female, X.2002, (MNRJ 6443); 1 female, VI.2001, E.H. Wienskoski (MNRJ 6430); 1 female, 25.V.1988, R.L.C. Baptista (MNRJ 3298); **Paraná**: Fernandes Pinheiro (Floresta Nacional de Irati), 25°23'2.22"S 50°34'55.07"W, 1 male, 7.IV.2012, J.Ricetti (MPEG 20261); Candói (Usina Hidrelétrica de Fundão, Rio Jordão), 25°43'30.43"S 52° 1'32.19"W, 2 males and 12 females, V.2005, J. Ricetti (MPEG 20262);**Santa Catarina:** (Reserva Biológica Marinha, Ilha do Arvoredo) [27°16'60''S 48°22'W], 1 female, 3–4.V.1996, A.A. Lise et. al. (MCTP 10493); **Rio Grande do Sul:** Vicente Dutra [27°10'0.33"S 53°24'19.65"W], 1 male, II.2006, T. Freitas (MCTP 19919); Gramado [29°22'7.65"S 50°52'43.12"W], 1 male, VII.2008, A.A. Lise (MCTP 22572); São Francisco de Paula [29°26'48.23"S 50°34'48.33"W] (Potreiro Velho), 1 male, 02.VI.2000, A.A. Lise (MCTP 14633); Sapiranga [29°38'39.71"S 51° 0'34.83"W] (Picada Verão), 1 female, 27.XII.1983, M.N. Strieder (MCTP 18077); Taquara [29°38'29.61"S 50°47'43.95"W], 1 male and 1 female, 18.VI.1988, F.C. Quadros (MCTP 26098); (Santa Cruz do Pinhal) [29°45'17.53"S 50°48'26.67"W], 1 male, 8.VIII.1993, L. Buckup (MCTP 4333). PERU. **Cusco:** Rio Camisea (Cashiari),

11°52'57"S 72°39'02"W, 1 male, 29.XI.1997, S. Cordova (MUSM 504355); **Puno**: Sandia [14°14'41.93"S 69°25'51.93"W], 1 female, 9.XII.1997, J.A. Ochoa (MUSM 504195). BOLIVIA. **La Paz**: Nor Yungas (Coroico) [16°11'39.04"S 67°43'43.72"W], 1 female, 1.III.1990, E. Penaranda (IBN); 1 male, 24.II.1990 (IBN).

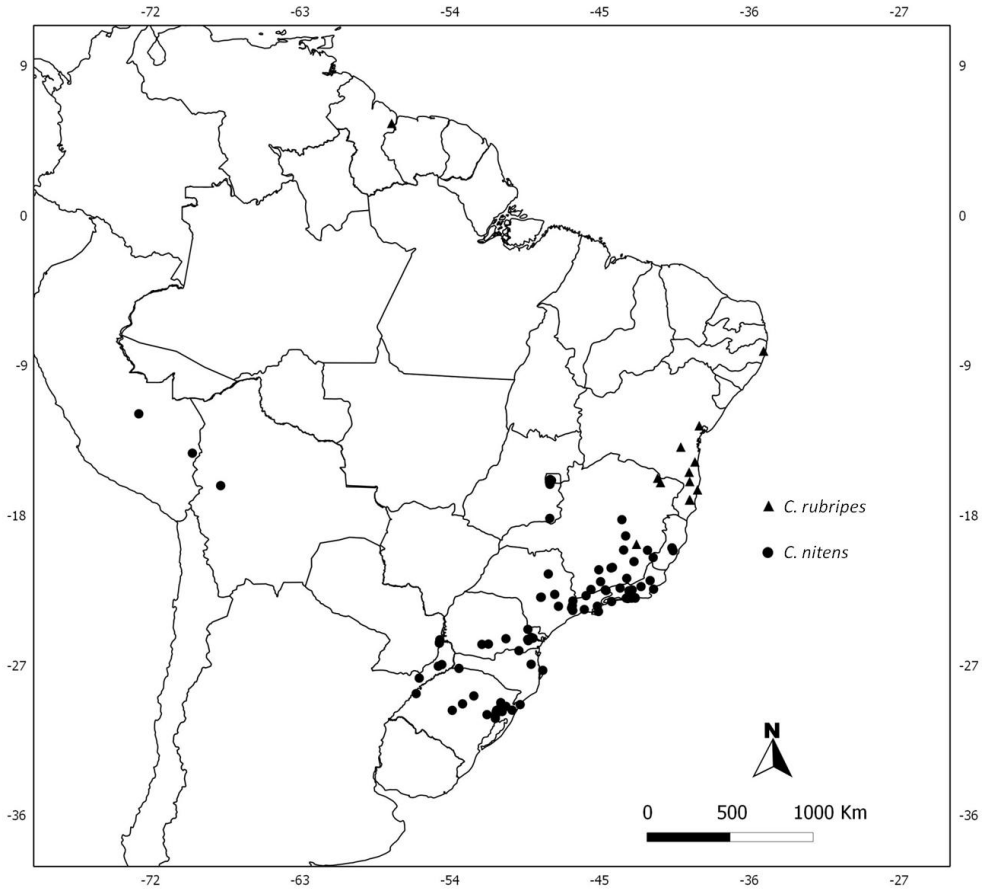


FIGURE 41. Known distribution of *Corinna rubripes* and *C. nitens.* Records from Bonaldo (1996, 2000) included.

***Corinna mourai* Bonaldo, 2000**

Fig. 42

Corinna mourai Bonaldo, 2000: 42, figs. 123–126 (holotype from Curitiba, Paraná, Brazil, A. Yamamoto, MCN 9290, not re-examined).

Diagnosis. Males of *Corinna mourai* resemble those of *C. escalvada* n. sp. and *C. balacobaco* n. sp. by the wide base of RTA, occupying most of ventral tibial surface; they differ from those of *C. balacobaco* n. sp. by the absence of a median process the RTA prolateral surface and from those of *C. escalvada* n. sp. by the apex of RTA rounded and directed prolaterally (Bonaldo 2000: figs. 123–124); females differ from those of other species with epigynal plate not projected posteriorly and with small, unconspicuous copulatory opening by the secondary spermathecae touching each other (Bonaldo, 2000: 125–126).

Description. See Bonaldo, 2000: figs. 123–126.

Distribution. Southeastern to southern Brazil.

New records: BRAZIL. **Minas Gerais:** Catas Altas (RPPN Parque Natural do Caraça) [20°05'51"S 43°29'18"W], 1 female (UFMG 10369); **Paraná:** Colombo [25°17'35.05"S 49°13'23.77"W], 1 male and 1 female, VI.2011, D. F. Candiani (MPEG 20274); Candói (Usina Hidrelétrica de Fundão, Rio Jordão), 25°43'30.43"S 52° 1'32.19"W, 6 females, V.2005, J. Ricetti (MPEG 20267); São João do Triunfo (Estação Experimental de São João do Tiunfo - UFPR), 25°41'18.96"S 50° 9'58.29"W, 1 male, 31.III.2012, J. Ricetti (MPEG 20272); 1 female (MPEG 20273) Fazenda Rio Grande (Fazenda Experimental Gralha Azul - PUCPR), 25°39'34.29"S 49°16'44.78"W, 2 males, 18.II.2012, J. Ricetti (MPEG 20269); 1 female, 4.IV.2011 J. Ricetti (MPEG 20270); 1 female (MPEG 20271); Tijucas do Sul (Tabatinga), 25°52'25.33"S 49° 8'53.83"W, 1 female, 10.V.2012, J.Ricetti and B.V.B. Rodrigues (MPEG 20264); (Haras Carlos dos Santos), 25°54'8.82"S 49° 8'17.65"W, 1 female, 8.V.2012, C.E. Conte (MPEG 20275); 4 females, J.Ricetti and B.V.B. Rodrigues (MPEG 20263); (Serra do Cabral), 25°57'23.08"S 49°14'48.63"W,1 female, 2.IV.2012, J. Ricetti (MPEG 20265); 1 female (MPEG 20266); **Rio Grande do Sul:** Gramado [29°22'7.65"S 50°52'43.12"W], 1

female, VII.2008, A.A. Lise (MCTP 22572); 1 female, 06.III.2011 (MCTP 29956); São Francisco de Paula [29°26'48.23"S 50°34'48.33"W], 1 male, 24.X.1996, R. Ott (MCTP 10668); (Barragem dos Bugres), 1 male and 2 females, L.A. Moura (MCN 30668); (Potreiro Velho), 2 female, 2–4.II.1999, A.A. Lise (MCTP 15841); 1 female, 02.VI.2000, A.A. Lise (MCTP 14633); 1 male, IV.2002, L.A. Bertoncello *et. al*. (MCTP 23104); 1 male (MCTP 23107); 1 male (MCTP 23099); 1 male (MCTP 23100); Triunfo (Parque COPESUL) [29°53'21.14"S 51°23'4.67"W], 1 female, 14–15.I.1997, L.A. Moura (MCN 28248); Eldorado do Sul (Parque Estadual do Delta do Jacuí), 2 males and 2 females, 5–7.I.2000, A.B. Bonaldo (MCN 31977).

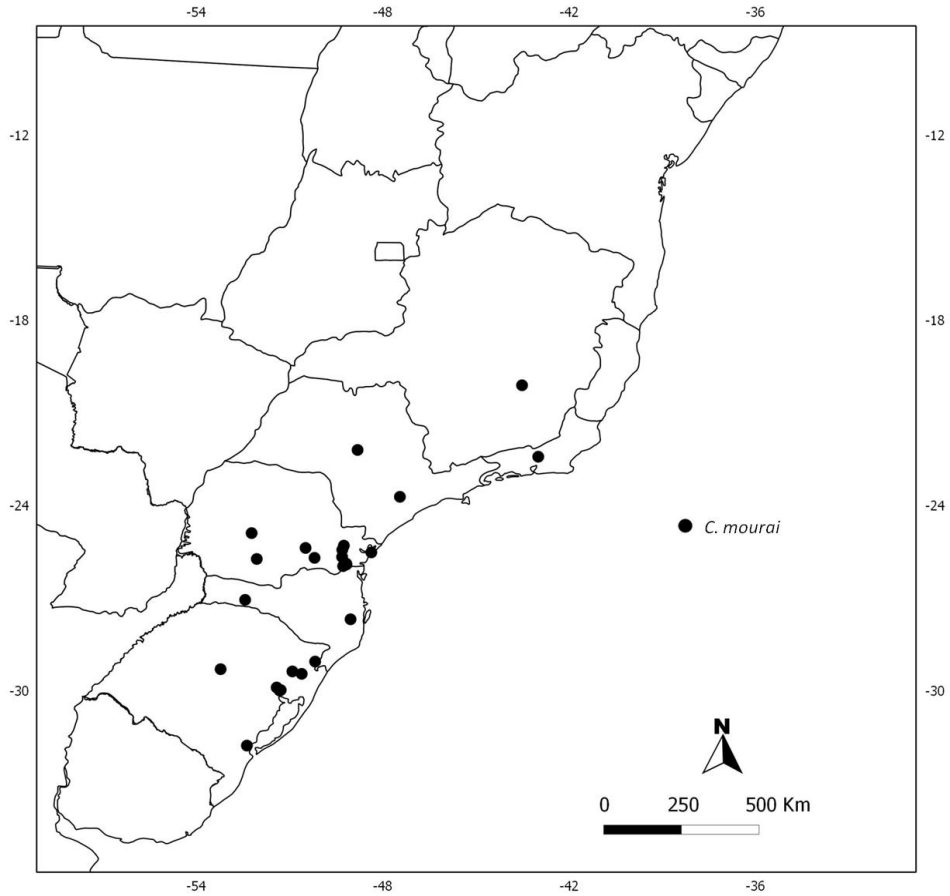


FIGURE 42. Known distribution records of *Corinna mourai* in Southeastern and Southern Brazil. Records from Bonaldo (2000) included.

***Corinna telecoteco* n. sp.**

Figs. 35, 43–44, 49

Type Material. Male holotype from Serra do Caraça [20°7'60.00"S 43°30'0.00"W], Minas Gerais, Brazil, 12–23.XI.1961, U. Martins-Lenko and R. Kloss, deposited in MZSP (6688). Paratypes: Brazil. Minas Gerais: (Serra do Cipó) [19°12'31.53"S 43°46'59.87"W], 1 male (MNRJ 1775).

Etymology. The specific name refers the classic samba tune "teleco-teco", interpreted by Cyro Monteiro in his Long Play "Sr. Samba" (Mr. Samba) released in 1961, the year in which the holotype was collected.

Diagnosis. Males of *Corinna telecoteco* n. sp. are similar to those of C. *ziriguidum* n. sp. by the combined presence of a small median process on prolateral surface of RTA and tegular process inserted closely to the embolar base; differ by presenting the median projection on prolateral surface of RTA directed apically and the tegular process curved prolaterally (Figs. 43–44).

Description. Male (holotype). Carapace reddish brown, chelicerae black, endites and labium brown, with lightened posterior end, sternum yellow, legs I dark yellow, legs II, III and IV yellow; abdomen light yellow with dorsal scutum in the anterior half of abdomen. Total length: 10.2. Carapace 4.6 long; 3.95 wide; 2.65 high. Clypeus 0.43 high. Abdomen 5.3 long; 3.5 wide. Eyes: anterior row 1.75, posterior row 2.1. MOQ: 0.63 long, 0.68 anterior wide, 0.68 posterior wide. Eyes diameters and interdistances: AME 0.23; ALE 0.2; PME 0.2; PLE 0.2; AME–AME 0.25; AME–ALE 0.35; PME–PME 0.3; PME–PLE 0.6; ALE–PLE 0.13. Chelicerae 2.35 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.35 long; 2.05 wide. Leg measurements: femur I 4.0/ patella 1.7/ tibia 3.5/ metatarsus 3.0/ tarsus 1.75/ total 13.95; II 3.65/ 1.7/ 3.0/ 2.75/ 1.4/

12.5; III 2.95/ 1.4/ 2.05/ 2.5/ 1.1/ 10.0; IV 3.8/ 1.5/ 3.25/ 3.3/ 1.2/ 13.05. Leg formula 1423. Leg spination: I – femur d1-0-0, p0-0-1; tibia v2-1r-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr-1-1-1; metatarsus v2-1p-1r. III – femur d1-1-0, p0-0-1, r0-1-0; tibia v2-2-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, vp1-1-0; metatarsus r0-1-0, v2-1p-1. Palp: femur with two posterior dorsal spines; t without projection; E filiform partially covered by the conductor apical fold (Figs. 43–44).

Female. Unknown.

Variation. Length (2 males): total 7.8–10.2; carapace 3.6–4.6; femur I 3.25–4.0.

Distribution. Minas Gerais, Brazil.

Other material examined. None

***Corinna ziriguidum* n. sp.**

Figs. 6–7, 9–10, 16, 24, 26–27, 45–46, 49

Type Material. Male holotype from Torre da Telepar, Curitiba, [25°25'42.08"S 49°16'23.71"W] Paraná, Brazil, 01.IV.1987, J.G. Kastelic, deposited in MCN (17198). Paratypes: Brazil. Rio de Janeiro: Petropólis [22°30'16.70"S 43°10'56.38"W], 1 male (MNRJ 704); Paraná: [Guaratuba, 25°43'35.67"S 48°56'56.19"W] (Usina Hidrelétrica de Guaricana), 1 male, 27–31.I.1986, S.R. Malkowiski (MHCI); Paranaguá (Praia Grande, Ilha do Mel), [25°30'42.00"S 48°20'20.00"W], 1 male, 08.I.1989, R. Dutra (MCN 20553).

Etymology. From Brazilian Portuguese, the specific name is an onomatopoeia imitating the sounds of percussion instruments used in samba. This world appers in the lirics of

several samba tunes and is also used to qualify the skills of samba players or dancers, as synonym of expertise.

Diagnosis. Males of *Corinna ziriguidum* n. sp. resemble those of *C. telecoteco* n. sp. by the presence of a small median projection on prolateral surface of RTA and tegular process inserted closely to the embolar base; differ by the course of reservoir in ventral view, with FR1 ample, reaching the retroapical region of tegulum and by the small median projection in the prolateral surface of the RTA directed prolaterally (Figs. 45–46).

Description. Male (holotype). Carapace dark reddish brown, chelicerae black, endites and labium red with posterior extremity lightened, sternum brown, leg I brown, legs II, III and IV dark yellow, posterior femora, tibiae and patellae with dark spots; dorsum of abdomen gray with dorsal scutum extending to the median region, ventrally light gray. Total length 8.8. Carapace 4.4 long; 3.5 wide; 2.5 high. Clypeus 0.4 high. Abdomen 4.25 long; 2.75 wide. Eyes: anterior row 1.7, posterior row 2.0. MOQ: 0.65 long, 0.55 anterior wide, 0.53 posterior wide. Eyes diameters and interdistances: AME 0.2; ALE 0.18; PME 0.18; PLE 0.15; AME–AME 0.25; AME–ALE 0.32; PME–PME 0.3; PME–PLE 0.55; ALE–PLE 0.16. Chelicerae 2.2 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.1 long; 1.8 wide. Leg measurements: femur I 3,5/ patella 1,5/ tibia 3,2/ metatarsus 2,4/ tarsus 1,5/ total 12,6; II 3,1/ 1,4/ 2,5/ 2,5/ 1,4/ 10,9; III 2,5/ 1,2/ 1,8/ 2,2/ 1,05/ 8,75; IV 3,3/ 1,4/ 3,1/ 3,0/ 1,0/ 11,8. Leg formula 1423. Leg spination: I – femur d1-0-1, p0-0-1; tibia v2-2-2-2; metatarsus vp1-1-0, vr1-0-1. II – femur d1-0-1, p0-0-1; tibia vr-1-1-1; metatarsus vp1-1-0, vr1-0-1. III – femur d1-1-1, p0-1-1, r0-0-1; tibia vp1-1-0, vr1-1-0, r1-0-1; metatarsus p0-1-0, r0-1-0, v2-1p-1. IV – femur d1-1-1, r0-0-1; tibia r1-0-1, vp1-1-0; metatarsus r0-1-0, v2-1p-1. Palp: femur with two posterior dorsal

spines; T without projection, course of reservoir in ventral view with a broad first loop; E filiform partly covered by apical groove of conductor (Figs. 45–46).

Female. Unknown.

Variation. Length (4 males): total 7.8–9.5, carapace 3.9–4.7, femur I 3.5–3.9.

Distribution. Rio de Janeiro and Paraná, Brazil.

Other material examined. None

***Corinna escalvada* n. sp.**

Figs. 34, 47–49

Type Material. Male holotype from Fazenda Escalvada, Mucuri [18°3'19.97"S 39°32'58.78"W], Bahia, Brazil, 11.IX.1979, A. C. Niella, deposited in SMNK.

Etymology. The specific name is a noun in apposition taken from the type locality. In Portuguese, escalvada is an adjective and means devoid of vegetation.

Diagnosis. Males of *Corinna escalvada* n. sp. differ from those of other species with tegular process inserted closely to the embolar base by the massive, wide base of RTA, without a median projection on its prolateral surface (Figs. 47–48).

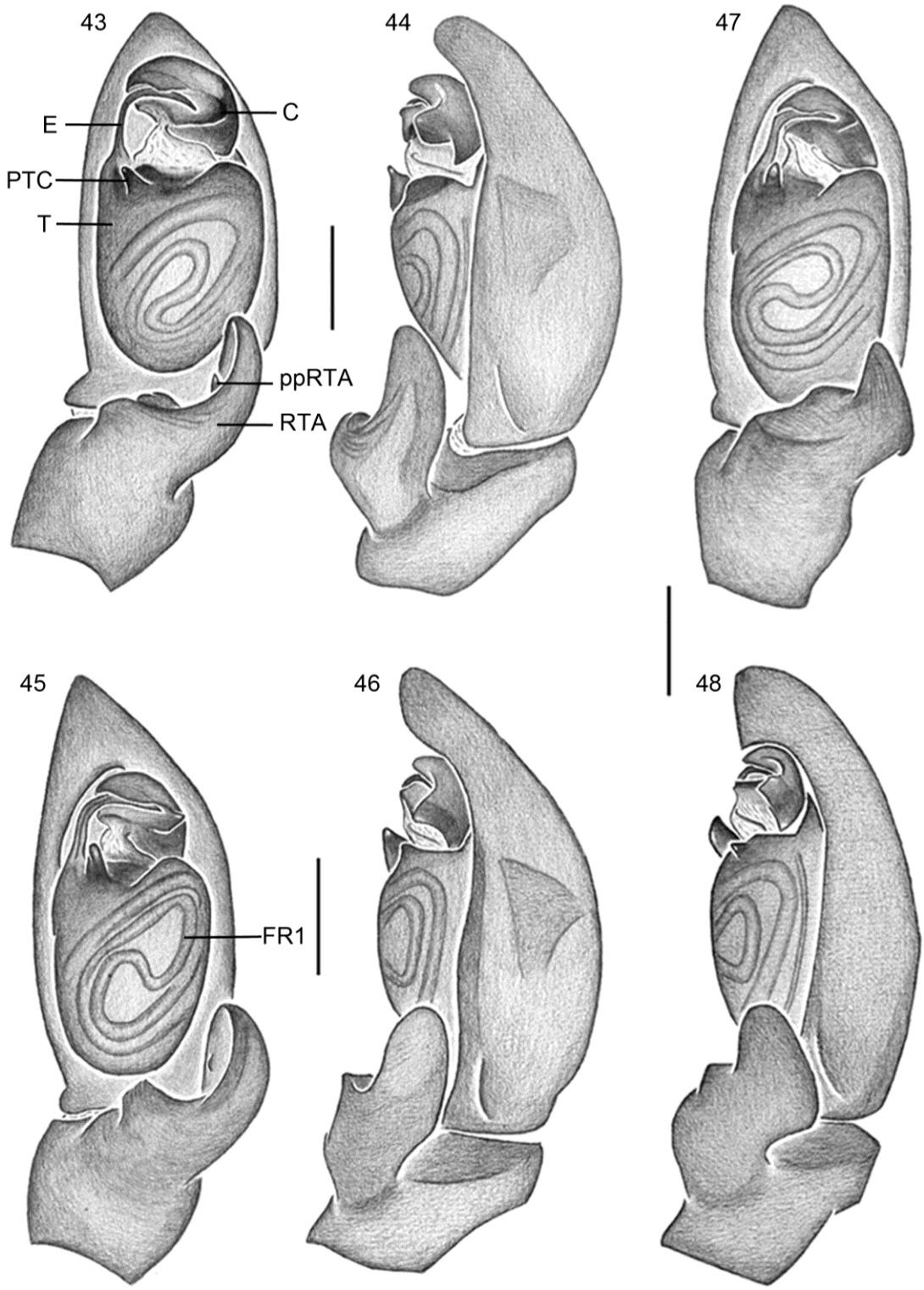
Description. Male (holotype). Carapace dark reddish brown with posterior end of thoracic region red and chelicerae black, endites and labium red with posterior extremity lightened, sternum brown, leg I with coxae, trochanter, femur and patella brown, tibia, metatarsus and tarsus dark red, legs II, III and IV dark yellow, except metatarsus and tarsus brown; dorsum of abdomen gray, with dorsal scutum on anterior third, ventrally light gray. Total length: 10.0. Carapace 5.0 long; 4.0 wide; 3.0 high. Clypeus 0.53 high. Abdomen 4.8 long; 4.3 wide. Eyes: anterior row 1.95, posterior row 2.1. MOQ: 0.8 long, 0.83 anterior wide, 0.78 posterior wide. Eyes diameters and

interdistances: AME 0.28; ALE 0.2; PME 0.2; PLE 0.2; AME–AME 0.33; AME–ALE 0.43; PME–PME 0.38; PME–PLE 0.6; ALE–PLE 0.25. Chelicerae 2.55 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.5 long; 2.1 wide. Leg measurements: femur I 4.05/ patella 2.0/ tibia 3.5/ metatarsus 3.0/ tarsus 1.75/ total 14.3; II 3.7/ 1.85/ 3.05/ 3.05/ 1.55/ 13.2; III 3.3/ 1.65/ 2.3/ 2.65/ 1.3/ 11.2; IV 4.0/ 1.75/ 3.25/ 3.5/ 1.35/ 13.85. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v1p-2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr-1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-1-1, r0-1-0; tibia v2-2-0; metatarsus p1-0-0, r0-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1; tibia r0-1-1, v2-1r-1p; metatarsus r0-1-0, v2-2-1. Palp: femur with two posterior dorsal spines; RTA basally wide, apex tapered; T without projection; E filiform partially covered by the distal groove of conductor (Figs. 47–48).

Female: Unknown.

Distribution. Known only from the type locality.

Other material examined. None.



FIGURES 43–48. *Corinna* spp., male palp. *Corinna telecoteco* n. sp., 43) ventral; 44) retrolateral. *Corinna ziriguidum* n. sp.: 45) ventral; 46) retrolateral. *Corinna escalvada* n. sp.e: 47) ventral; 48) retrolateral. Scale bar = 0.5 mm. Abbreviations: C–Conductor; E–Embolus; FR1- First fold of reservoir; ppRTA–Prolateral process of Retrolateral tibial apophysis; RTA–Retrolateral tibial apophysis; T–Tegulum; TPC–Tegular process of *Corinna*.

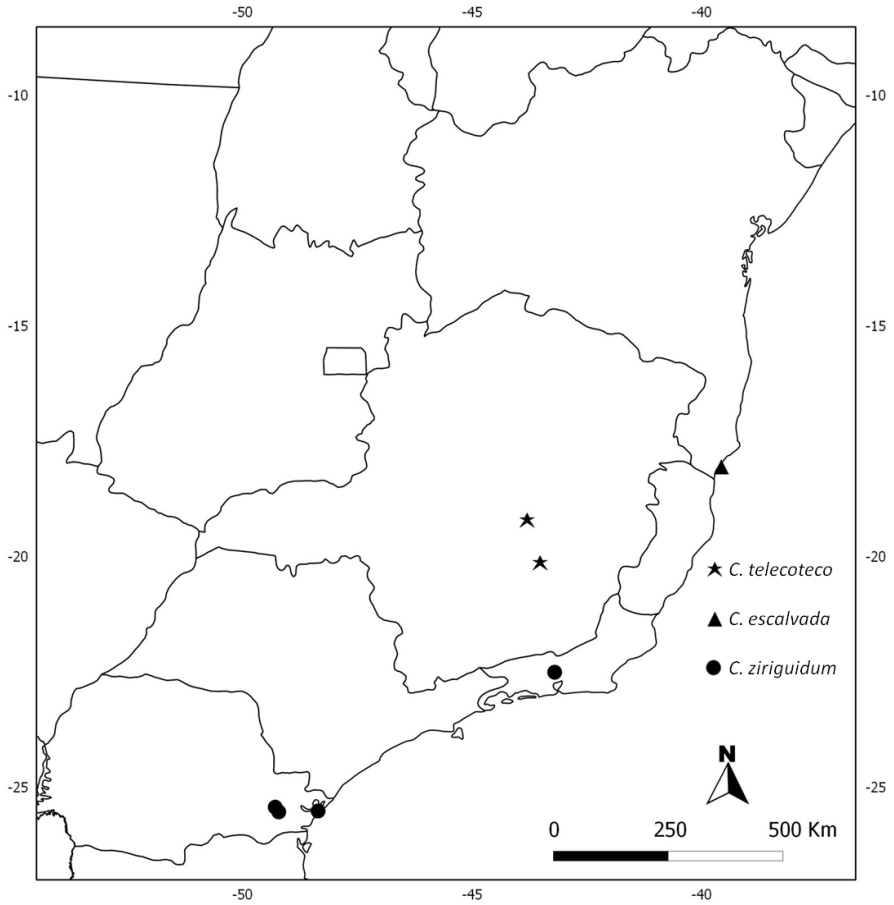


FIGURE 49. Known distribution records of *Corinna telecoteco* n. sp., *C. escalvada* n. sp. and *C. ziriguidum* n. sp. in Southeastern Brazil.

***Corinna aechmea* n. sp.**

Figs. 1–5, 11,–12, 18–23, 31–33, 50–53, 62

Type Material. Male holotype from Porto Rico, 22°44'38.78"S 53°12'59.85"W, Paraná, Brazil, 18.III.2010, F. Amadeo, deposited in MPEG (20248). Paratypes: Brazil. Minas Gerais: Belo Horizonte (Estação Ecológica da UFMG) [19°52'56"S 43°58'18"W], 1 male and 1 female, 30.VI.1993, H.R. Pimenta (UFMG); São Paulo: (Usina Hidrelétrica de Rosana), [22°35'57.72"S 52°52'29.32"W], 1 male, XII.1986 (IBSP 4513); Paraná: Porto Rico (Planície de Inundação do Alto Rio Paraná), 22°44'38.78"S 53°12'59.85"W, 1 female, 25.II.2010, F. Amadeo (MPEG 20249); 1 female (MPEG 20250).

Etymology. The specific name is a noun in apposition taken from the scientific name of the bromeliad genus in which the holotype was collected, *Aechmea distichantha* Lem.

Diagnosis. Males of *Corinna aechmea* n. sp. resemble those of *C. jecatatu* n.sp. and *C. zecarioca* n. sp. by presenting the RTA bifurcated, with a large median process in the prolateral suface, but it is readly recognized by the rounded apical tegular projection (Figs. 50–51); females differ from those of *C. zecarioca* n. sp. by presenting the secondary spermathecae closer to each other and from those of *C. jecatatu* n. sp. by the posterior margin of copulatory opening "u"- shaped and posterior vulval plate with lateral rounded protuberances in the internal margin (Figs. 52–53).

Description. Male (holotype). Carapace black with posterior end of thoracic region dark red, chelicerae black, endites red, posterior end paler; labium black, posterior end paler; sternum brown, legs with coxae, trochanters and femora dark brown, patellae, tibiae, metatarsi and tarsi dark yellow; dorsum of abdomen light gray, dorsal scutum with distal portion large tapering to the median region; abdomen ventrally light gray with large median brown band in longitudinal axis. Total length: 11.1. Carapace 5.0 long; 4.2 wide; 2.7 high. Clypeus 0.4 high. Abdomen 5.4 long; 3.3 wide. Eyes: anterior row 1.9, posterior row 2.2. MOQ: 0.68 long, 0.76 anterior wide, 0.73 posterior wide. Eyes diameters and interdistances: AME 0.27; ALE 0.21; PME 0.21; PLE 0.2; AME–AME 0.26; AME–ALE 0.37; PME–PME 0.33; PME–PLE 0.58; ALE–PLE 0.14. Chelicerae 3.0 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.6 long; 2.15 wide. Leg measurements: femur I 4.25/ patella 2.1/ tibia 3.6/ metatarsus 3.15/ tarsus 1.75/ total 14.85; II 3.65/ 1.9/ 3.05/ 2.95/ 1.6/ 13.15; III 3.2/ 1.6/ 2.25/ 2.75/ 1.2/ 11; IV 4.05/ 1.7/ 3.5/ 3.7/ 1.3/ 14.25. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr-1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-1-2, r0-0-1; tibia v2-2-0, r0-1-1; metatarsus

p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, vp1-2-0; metatarsus r0-1-0, v2-1p-1. Palp: femur with two posterior dorsal spines; TPC inserted far from base of embolus; E filiform, partly covered by the apical groove of conductor; C with large unsclerotized area in the distal region (Figs 50, 51).

Female (paratype MPEG 20249). Coloration as in male, except dorsum of abdomen gray, without dorsal scutum. Total length: 11.3. Carapace 4.7 long; 3.8 wide; 2.4 high. Clypeus 0.38 high. Abdomen 6.5 long; 4.1 wide. Eyes: anterior row 1.77, posterior row 2.05. OQ: 0.62 long, 0.71 anterior wide, 0.68 posterior wide. Eyes diameters and interdistances: AME 0.23; ALE 0.2; PME 0.18; PLE 0.18; AME–AME 0.28; AME–ALE 0.37; PME–PME 0.35; PME–PLE 0.44; ALE–PLE 0.14. Chelicerae 2.5 long. Sternum 2.3 long; 2.05 wide. Leg measurements: femur I 3.8/ patella 1.85/ tibia 3.25/ metatarsus 2.65/ tarsus 1.6/ total 13.15; II 3.45/ 1.7/ 2.8/ 2.55/ 1.45/ 11.95; III 2.85/ 1.4/ 2.2/ 2.4/ 1.1/ 9.95; IV 3.65/ 1.5/ 3.2/ 3.25/ 1.15/ 12.75. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr-1-1-1; metatarsus v2-3-0. III – femur d1-1-1, p0-1-1, r0-1-0; tibia v2-2-0, r0-1-1; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1; tibia r0-1-1, vp1-r1-p1; metatarsus r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO median, small and inconspicuous; CD not visible ventrally; PVP enveloping partly the PS and SS (Figs. 52–53).

Variation. Length (8 males): total 7.4–11.1; carapace 3.8–5.1; femur I 2.7–3.8; (6 females) total 8.0–12.2; carapace 3.8–5.1; femur I 3.2–4.1.

Distribution. Minas Gerais and Paraná, Brazil.

Other material examined. Brazil. **Minas Gerais**: Uberlândia [18°54'40.64"S 48°15'43.83"W] (Mata do Glória), 1 male, 24.VI.1993, P.C. Motta (UNB 464); Santa Bárbara (RPPN, Estação de Preservação e Desenvolvimento Ambiental de Peti),

19°53'S 43°22'W, 3 males and 1 female, 05–06.VI.2010, A.J. Santos (UFMG 4460); 1 female, 07.IV.2004, E.T. Rodrigues; Belo Horizonte (Parque Municipal das Mangabeiras), 19°57'14.86"S 43°54'19.15"W, 1 male, 5–12.XII.2008, H.H. Santos et. al. (UFMG 8508); **Paraná**: Porto Rico (Planície de Inundação do Alto Rio Paraná), 22°44'38.78"S 53°12'59.85"W, 1 female, 15.IV.2010, F. Amadeo (MPEG 20251); Vila Alta [currently Alto Paraíso] (Ilha Grande), [23°45'0.00"S 54°2'60.00"W], 2 males, 08.XII.1995, M.L. Fischer (MPEG); Altônia (Ilha do Cristo), [23°52'21.25"S 53°53'48.30"W], 1 female, 08.X.1989, R.Pinto da Rocha (MHCI).

***Corinna jecatatu* n. sp.**

Figs. 54–57, 62

Type Material. Male holotype from Barueri [23°30'40.69"S 46°52'36.39"W], São Paulo, Brazil, 06.XII.1964, K. Lenko, deposited in MZUSP (3821). Paratypes: Brazil. São Paulo: Barueri [23°30'40.69"S 46°52'36.39"W], 1 female, 16.I.1966, K. Lenko, (MZUSP 5591); 1 male, 06.XII.1964, K. Lenko (MZUSP 3821); Osasco [23°31'54.09"S 46°47'23.72"W], 1 male and 1 female, 15.IV.1938, F.Lane (MZUSP 12525); São Paulo [23°32'56.19"S 46°38'19.74"W], 1 female, 11.VII.1937, J.R. Mattos, (IBSP); 1 male, I.1960 (IBSP 1480).

Etymology. The specific name refers to classic hick character "Jeca Tatu", created by the Brazilian writer Monteiro Lobato for his masterpiece "Urupês", first published in 1918. The character was based in the countrymen from the state of São Paulo countryside and was intended to denounce the poor living conditions of those Brazilians.

Diagnosis. Males of *Corinna jecatatu* n. sp. resemble *C. aechmea* n. sp. and *C. zecarioca* n. sp by presenting the RTA bifurcated, with a large median process in the prolateral suface; differ from those of *C. aechmae* n. sp. by the absence of a rounded tegular apical projection and from those of *C. zecarioca* n. sp. by the longer RTA median process (Figs. 54–55); as in *C. aechmae* n. sp., females present the secondary spermathecae close to each other, but are readily recognized by posterior margin of copulatory opening straight and posterior vulval plate without internal protuberances (Figs. 56–57).

Description. Male (holotype). Carapace dark reddish brown with posterior end of thoracic region red, chelicerae black, endites red with posterior end paler; labium black with posterior end paler, sternum brown; legs I and II dark yellow except metatarsi and tarsi, dark red, leg III and IV yellow except metatarsi and tarsi, dark yellow; abdomen light yellow with dorsal scutum extending to the median region of abdomen. Total length: 10.2. Carapace 5.1 long; 4.3 wide; 3.1 high. Clypeus 0.5 high. Abdomen 4.7 long; 3.1 wide. Eyes: anterior row 1.9, posterior row 2.4. MOQ: 0.8 long, 0.87 anterior wide, 0.8 posterior wide. Eyes diameters and interdistances: AME 0.32; ALE 0.17; PME 0.2; PLE 0.22; AME–AME 0.22; AME–ALE 0.37; PME–PME 0.37; PME–PLE 0.6; ALE–PLE 0.15. Chelicerae 2.4 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.7 long; 2.1 wide. Leg measurements: femur I 4.3/ patella 2.05/ tibia 3.75/ metatarsus 3.3/ tarsus 1.7/ total 15.1; II 3.85/ 1.6/ 3.2/ 3.0/ 1.55/ 13.2; III 3.25/ 1.6/ 2.25/ 2.7/ 1.25/ 11.05; IV 4.1/ 1.8/ 3.5/ 3.75/ 1.35/ 14.5. Leg formula 1423. Leg spination: I – femur d1-0-1, p0-0-1; tibia v1p-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-1p-1r. III – femur d1-1-1, p0-1-0; tibia v2-2-0, r1-0-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v1p-1r-1p; metatarsus r0-1-0, v2-2-1. Palp: femur with one posterior dorsal spine; T without apical

projection; TPC placed far from base of embolus; E filiform, partially covered by distal fold of conductor; C with enlarged base and unsclerotized area in the distal region (Figs. 54–55).

Female (paratype MZSP 5591). Coloration as in male, except abdomen light gray, without dorsal scutum. Total length: 10.8. Carapace 4.8 long; 4.2 wide; 2.65 high. Clypeus 0.37 high. Abdomen 5.6 long; 3.9 wide. Eyes: anterior row 1.95, posterior row 2.35. MOQ: 0.8 long, 0.85 anterior wide, 0.8 posterior wide. Eyes diameter and interdistances: AME 0.3; ALE 0.2; PME 0.2; PLE 0.17; AME–AME 0.25; AME–ALE 0.35; PME–PME 0.37; PME–PLE 0.65; ALE–PLE 0.15. Chelicerae 2.75 long. Sternum 2.65 long; 2.3 wide. Leg measurements: femur I 3.65/ patella 2.0/ tibia 3.5/ metatarsus 3.0/ tarsus 1.6/ total 13.75; II 3.7/ 1.95/ 3.05/ 2.55/ 1.5/ 12.75; III 3.15/ 1.65/ 2.2/ 2.6/ 1.1/ 10.7; IV 4.15/ 1.85/ 3.55/ 3.6/ 1.3/ 14.45. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-1r-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-1-0; tibia v2-2-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v1p-1r-1p; metatarsus r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO median, small and inconspicuous; CD not visible ventrally; PVP without lateral protuberances enveloping partially PS and SS (Figs. 56–57).

Variation. Length (12 males): total 7.25–11.0; carapace 5.1–3.8; femur I 4.25–36; (4 females): total 8.6–12.0; carapace 5.9–4.8, femur I 4.5–3.65.

Distribution. São Paulo, Brazil.

Other material examined. Brazil. **São Paulo:** Guarulhos (Bonsucesso), [23°24'59.56"S 46°24'20.43"W], 1 male, 15–20.VII.1968, T.Heitgmann-Fontenelle (MZUSP 12524); Barueri, [23°30'40.69"S 46°52'36.39"W], 1 male, 20.III.1966, K.Lenko (MZUSP 5576); Carapicuiba, [23°31'9.20"S 46°50'12.26"W], 1 male,

V.1988, O.Boher (IBSP 56675); São Paulo, [23°32'56.19"S 46°38'19.74"W], 1 male, 1965, Biasi (MZUSP 4119); Mogi das Cruzes, [23°31'29.11"S 46°11'13.68"W], 1 male, V.1980, M. Vinicius (IBSP 28004); São Paulo (Ipiranga), [23°35'29.00"S 46°36'28.00"W], 2 males, 18.III.1942, M. Semedo (MZUSP 7257); 1 male and 1 female, IV.1942, B.A.M.Soares (MZUSP 6854); [Grajaú, Cocaia], Represa nova, [23°46'9.23"S 46°39'36.72"W], 1 female, 24.VIII.1947, H.Urban (MZUSP 12526); (Ilha da Queimada Grande), [24°29'4.99"S 46°40'31.97"W], 1 male, IV.1947, Exp. Butantan (IBSP).

***Corinna zecarioca* n. sp.**

Figs. 58–62

Type Material. Male from Gávea [22°59'10.35"S 43°14'42.60"W], Rio de Janeiro, Rio de Janeiro, Brazil, Wenskoski, E.H., deposited in MNRJ (06447). Paratype: Brazil. Rio de Janeiro: Petrópolis [22°30'16.70"S 43°10'56.38"W] (Estação Rio, 850m), 1 female, 2.XI.1945, H. Sick (AMNH).

Etymology. The specific name refers to Walt Disney's character "José Carioca" or, shortly, "Zé Carioca", created in 1942, in congruence with USA's World War II foreign police. The green parrot Zé Carioca is a friend of Donald Duck and his comic books still make success among Brazilian kids, including those born in the city of Rio de Janeiro, the Cariocas.

Diagnosis. Males of *Corinna zecarioca* n. sp. resemble those of *C. aechmea* n. sp. and *C. jecatatu* n. sp. by the bifurcated RTA, but can be recognized by the stout, sub-squared median process in the prolateral surface of RTA and by the tegular process positioned at the base of embolus (Figs. 58–59); females are readly reconized by the

globular secondary spermathecae separated from each other by more than one of its diameter (Figs. 60–61).

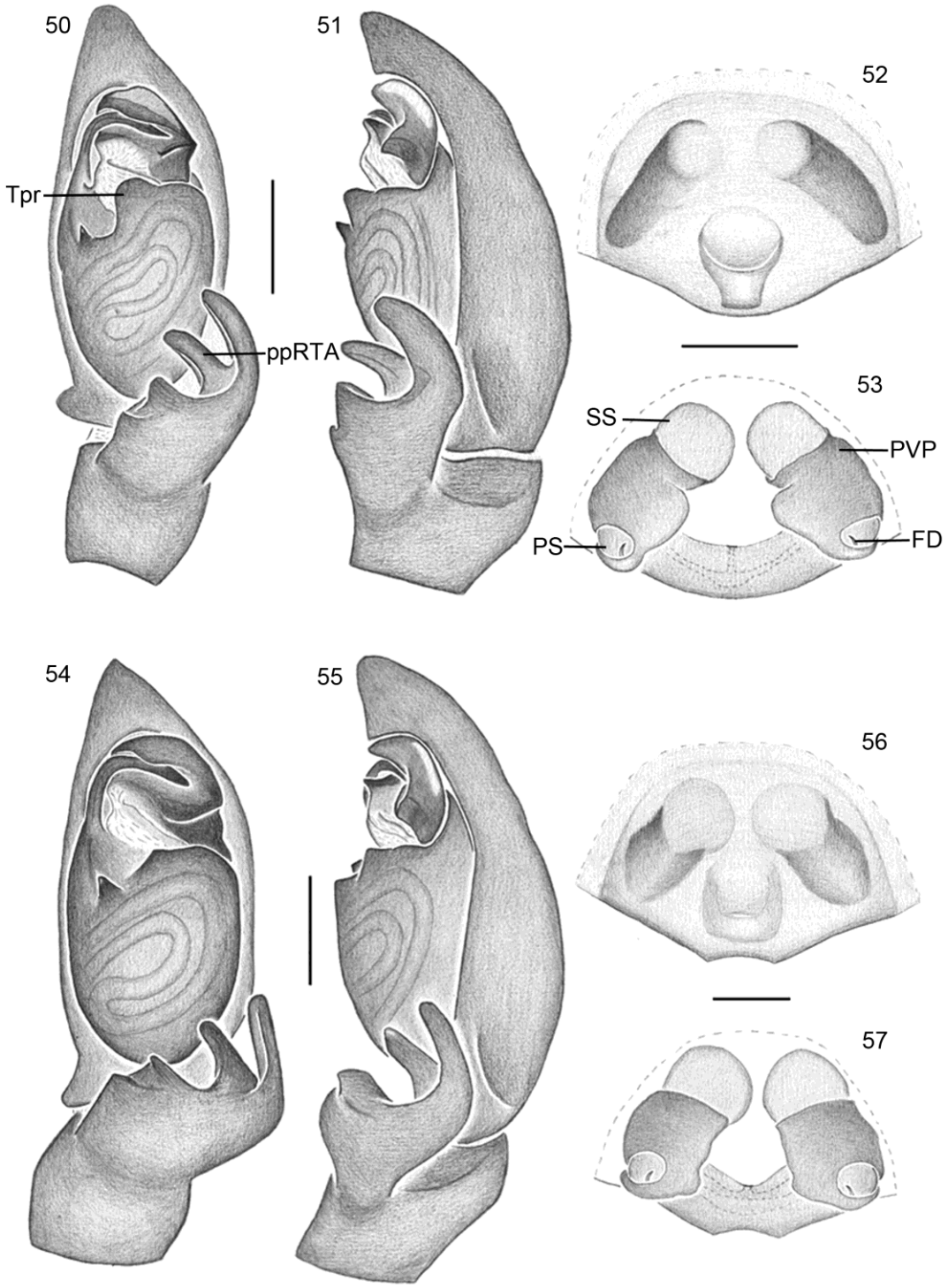
Description. Male holotype. Carapace black, with posterior end of thoracic region dark red; chelicerae black, endites red, with posterior end paler; labium black, with posterior end paler, sternum brown; leg I with coxae, trochanter, metatarsus and tarsus dark red, femur, patella and tibia dark yellow; legs II dark yellow, except metatarsus and tarsus, dark red, legs III and IV yellow except metatarsus and tarsus, dark yellow; dorsum of abdomen gray with dorsal scutum extending to the median region of abdomen, ventrally light yellow. Total length: 10.7. Carapace 5.3 long; 4.6 wide; 1.8 high. Clypeus 0.5 high. Abdomen 5.2 long; 3.4 wide. Eyes: anterior row 2.1, posterior row 2.5. MOQ: 0.81 long, 0.96 anterior wide, 0.9 posterior wide. Eye diameters and interdistances: AME 0.33; ALE 0.24; PME 0.24; PLE 0.24; AME–AME 0.27; AME–ALE 0.45; PME–PME 0.42; PME–PLE 0.69; ALE–PLE 0.18. Chelicerae 3.0 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.8 long; 2.4 wide. Leg measurements: femur I 4.8/ patella 2.3/ tibia 4.1/ metatarsus 3.6/ tarsus 2.0/ total 16.8; II 4.3/ 2.1/ 3.4/ 3.4/ 1.7/ 14.9; III 3.5/ 1.8/ 2.5/ 3.2/ 1.4/ 12.4; IV 4.5/ 1.9/ 3.9/ 4.3/ 1.5/ 16.1. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-1-1, r0-1-0; tibia v2-2-0, r0-1-1; metatarsus p1-0-0, r0-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1; tibia r0-1-1, v2-2-0; metatarsus r0-1-0, v2-2-1. Palp: femur with three posterior dorsal spines; E filiform, partially covered by distal groove of conductor; C without distal unsclerotized area (Figs. 58–59).

Female (paratype AMNH). Coloration as in male, except by abdomen light gray, without scutum. Total length 9.9. Carapace 5.0 long; 4.3 wide; 1.4 high. Clypeus 0.45 high. Abdomen 4.6 long; 3.4 wide. Eyes: anterior row 2.1, posterior row 2.4. MOQ:

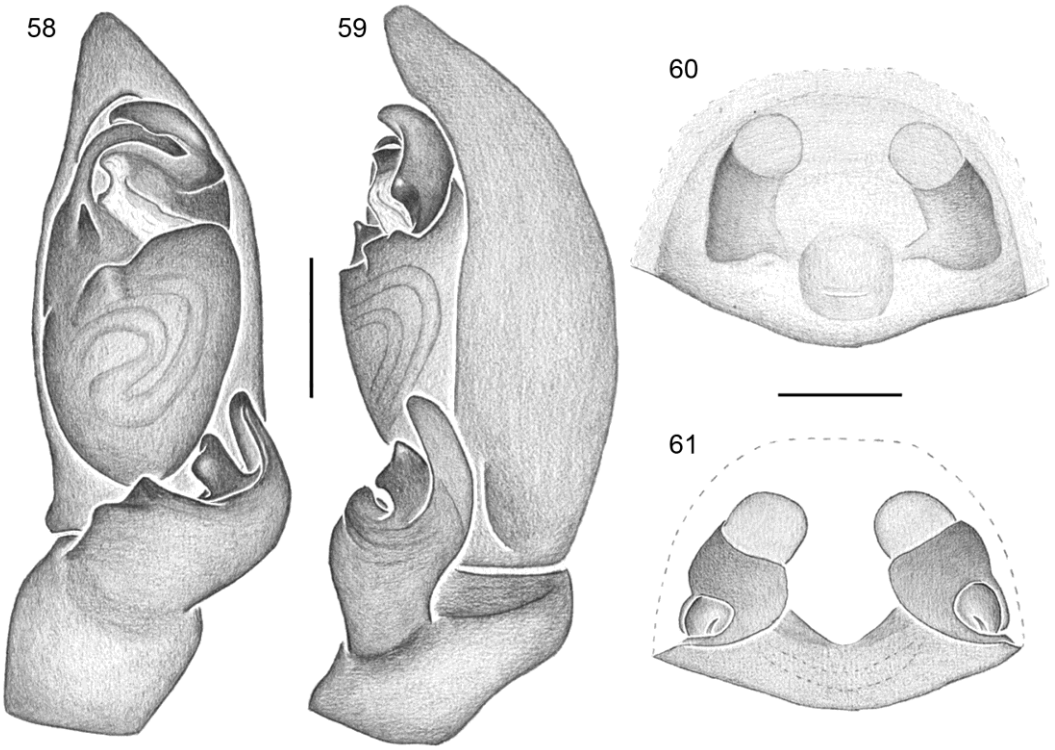
0.84 long, 0.87 anterior wide, 0.9 posterior wide. Eye diameters and interdistances: AME 0.3; ALE 0.25; PME 0.24; PLE 0.24; AME–AME 0.24; AME–ALE 0.37; PME–PME 0.39; PME–PLE 0.69; ALE–PLE 0.15. Chelicerae 3.0 long. Sternum 2.7 long; 2.3 wide. Leg measurements: femur I 4.4/ patella 2.0/ tibia 3.7/ metatarsus 3.0/ tarsus 1.6/ total 14.7; II 3.9/ 1.9/ 3.1/ 2.9/ 1.5/ 13.3; III 3.3/ 1.7/ 2.4/ 2.7/ 1.1/ 11.2; IV 4.1/ 1.9/ 3.7/ 3.7/ 1.2/ 14.6. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-2; tibia v1r-2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-1, r0-1-0, p0-0-1; tibia v2-2-0, r0-1-1; metatarsus p1-0-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v2-2-0; metatarsus r1-0-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO median, small and inconspicuous; CD not visible ventrally; PVP enveloping partially the PS and SS without lateral protuberances (Figs. 60–61).

Distribution. Rio de Janeiro, Brazil.

Other material examined. None.



FIGURES 50–57. *Corinna aechmea* n. sp., male: 50) palp, ventral; 51) same, retrolateral; female: 52) epigynum, ventral; 53) same, dorsal. *Corinna jecatatu* n. sp., male: 54) palp, ventral; 55) same, retrolateral; female: 56) epigynum, ventral; 57) same, dorsal. Scale bar = 0.5 mm. Abbreviations: FD–Fertilization ducts; ppRTA–Prolateral process of Retrolateral tibial apophysis; PS–Primary spermathecae; PVP–Posterior vulvar plate; SS–Secondary spermathecae; Tpr–Tegular projection.



FIGURES 58–61. *Corinna zecarioca* n. sp., male: 58) palp, ventral; 59) same, retrolateral; female: 60) epigynum, ventral; 61) same, dorsal. Scale bar = 0.5 mm.

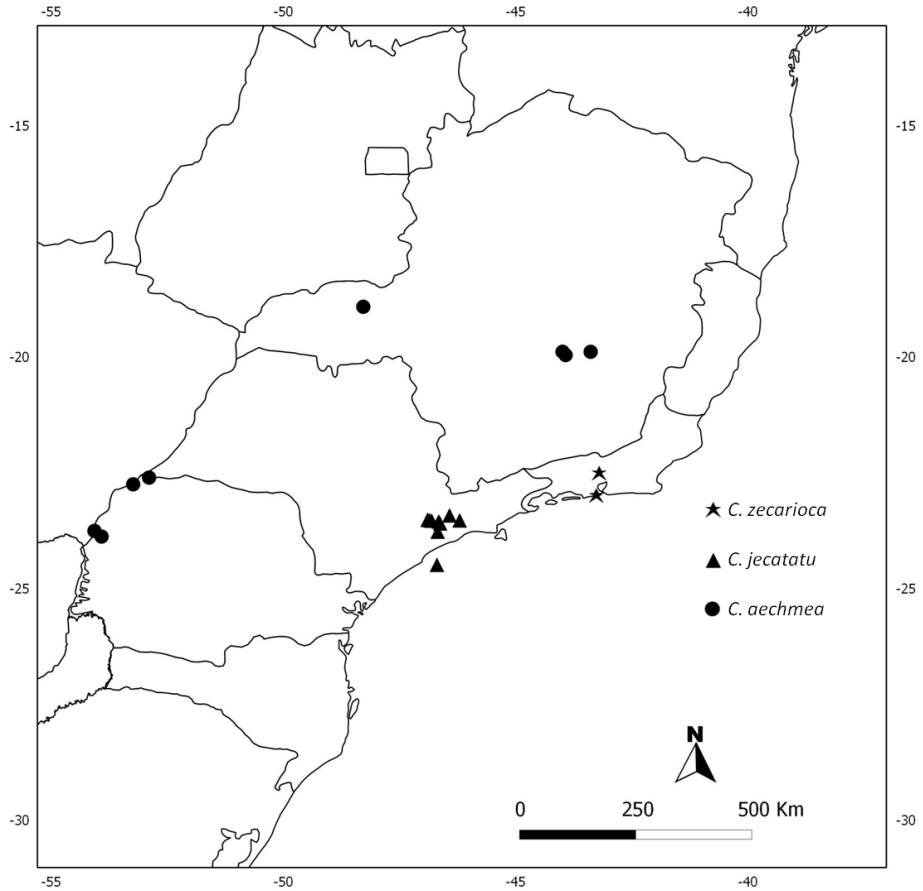


FIGURE 62. Known distribution records of *Corinna zecarioca* n. sp., *C. jecatatu* n. sp. and *C.aechmea* n. sp. in Southeastern and Southern Brazil.

***Corinna caatinga* n. sp.**

Figs. 36, 40, 63–73

Type Material. Male holotype from ECB Rochas Ornamentais Ltda, Fazenda Bonito, 05º14'07.1"S 41º41'16.3"W, Castelo do Piauí, Piauí, Brazil, deposited in MPEG (20257). Partypes: Brazil. Piauí: Caracol (Parque Nacional da Serra das Confusões), 09º13'16"S 43º29'21"W, 5 male (MPEG 20259); 1 female (MPEG 20260); 1 female, X.2006, P.R.R Silva et. al. (CHNUFPI 0164); Castelo do Piauí (Fazenda Bonito, ECB Rochas Ornamentais Ltda,), 05°12'42"S 41º42'13"W, 3 males (MPEG 20258); 4 male, 06.VI.2012, F.M. Oliveira-Neto (CHNUFPI 0158); Rio Grande do Norte: João Câmara, 05º32'15"S 35º49'11"W, 1 male and 1 female, VI.2012, N. Sebastian et. al. (MPEG 19970); Natal [5°47'40.12"S 35°12'39.43"W], 3 males, 1951, M. Alvarenga, (MZUSP 12626); Paraíba: [7°14'23.86"S 36°46'55.02"W], 1 male, R. Ihering (MNRJ 42241); Bahia: Feira de Santana [12°15'19.67"S 38°57'15.43"W] (Chácara Promenade), 1 male, 19.I.2003, T. Aguzzoli and A. Carvalho (MCN 35064).

Etymology. The specific name, in apposition, refers to a typical xerophytic phytophysiognomy from Northeastern Brazil, the "caatinga".

Diagnosis. Males of *Corinna caatinga* n. sp. are readly distinguished by the extremely developed chelicerae and enlarged cephalic region (Fig. 40); in the male palp, the tapered basal cymbial projection, the enlarged conductor and extremely long and delicate embolus are also unique (Figs. 63–64, 68–69); females present the same carapace and chelicerae modifications and can be further identified by the epigynal plate projected posteriorly with a median, conspicuous copulatory opening (Figs. 65–66).

Description. Male (holotype). Carapace black, with posterior end of thoracic region dark red, chelicerae black, endites red with posterior end paler, labium black with

posterior end paler, sternum dark red, leg I and II dark brown, leg III and IV dark yellow; dorsum of abdomen gray, dorsal scutum with distal portion large tapering to the median region, venter of abdomen gray. Total length: 13.3. Carapace 6.9 long; 6.2 wide; 4.2 high. Clypeus 0.5 high. Abdomen 6.3 long; 4.2 wide. Eyes: anterior row 2.45, posterior row 3.0. MOQ: 0.9 long, 1.0 anterior wide, 1.05 posterior wide. Eye diameters and interdistances: AME 0.3; ALE 0.31; PME 0.24; PLE 0.22; AME–AME 0.44; AME–ALE 0.46; PME–PME 0.6; PME–PLE 0.84; ALE–PLE 0.2. Chelicerae 5.4 long; 6 retromarginal teeth and 3 promarginal teeth. Sternum 3.5 long; 3.1 wide. Leg measurements: femur I 6.88/ patella 2.5/ tibia 6.15/ metatarsus 5.7/ tarsus 2.88/ total 24.11; II 6.0/ 2.55/ 5.2/ 5.12/ 2.4/ 21.27; III 5.05/ 2.5/ 3.6/ 4.72/ 1.84/ 17.71; IV 6.6/ 2.56/ 6.0/ 7.12/ 2.6/ 24.88. Leg formula 4123. Leg supination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2-2; metatarsus v2-1p-1r. II – femur d1-1-0, p0-0-1; tibia v2-2-2-2-2; metatarsus v2-1p-1r. III – femur d1-1-1, p0-1-2, r0-1-1; tibia v2-1r-1p, r0-1-1, p0-1-1; metatarsus p0-1-0, r1-1-0, v2-2-1. IV – femur d1-1-1, r0-1-1, p0-0-1; tibia r0-1-1, v2-0-2; metatarsus p0-1-0, r1-1-0, v2-2-1. Palp: femur with two posterior dorsal spines; RTA strongly excavated prolaterally; T without projection; TPC retrolateral, far from base of embolus; E long and filiform partially covered by distal fold of conductor; C enlarged with wide unexclerotized base (Figs. 63–64, 67–71).

Female (paratype, MPEG 20260). Coloration as in male, except carapace dark red with posterior end of thoracic region red; sternum, endites and labium brown; abdomen without scutum dorsal. Total length: 12.6. Carapace 7.0 long; 6.1 wide; 4.3 high. Clypeus 0.65 high. Abdomen 5.0 long; 4.1 wide. Eyes: anterior row 2.7, posterior row 3.3. MOQ: 0.8 long, 1.0 anterior wide, 1.2 posterior wide. Eye diameters and interdistances: AME 0.3; ALE 0.34; PME 0.27; PLE 0.19; AME–AME 0.44; AME–ALE 0.42; PME–PME 0.68; PME–PLE 0.89; ALE–PLE 0.25. Chelicerae 4.85 long.

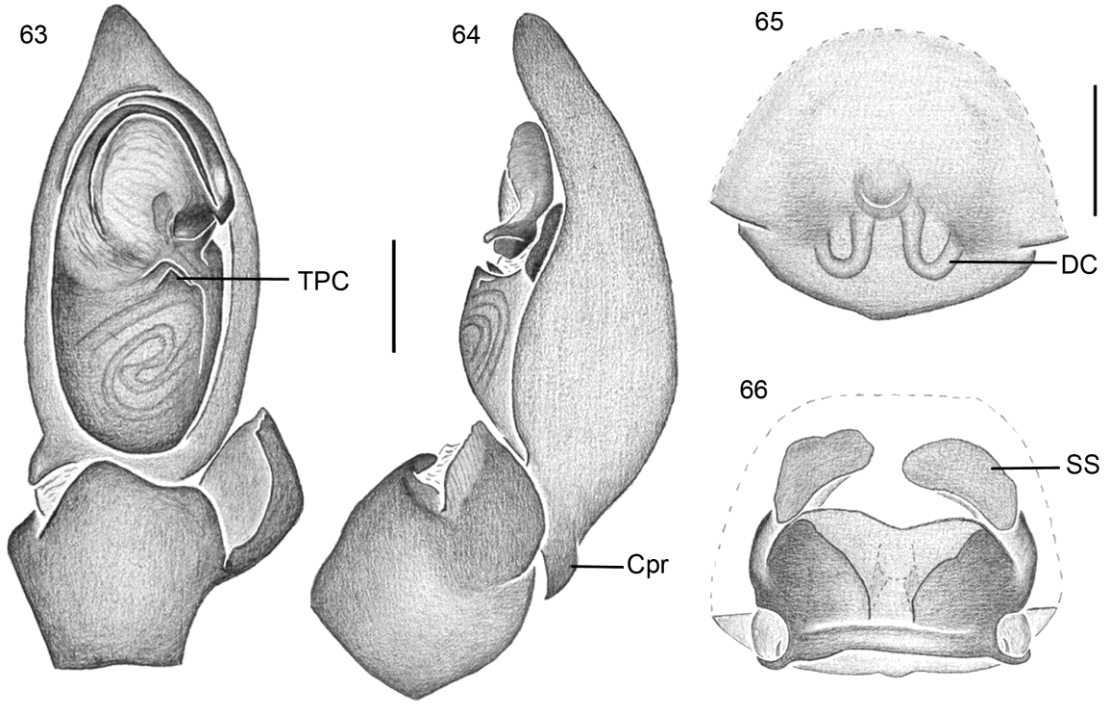
Sternum 3.55 long; 3.3 wide. Leg measurements: femur I 6.95/ patella 2.95/ tibia 5.7/ metatarsus 5.7/ tarsus 2.55/ total 23.85; II 6.3/ 2.7/ 5.12/ 5.3/ 2.3/ 21.72; III 5.05/ 2.4/ 3.6/ 4.95/2.0/ 18; IV 6.88/ 2.56/ 6.32/ 7.12/ 2.4/ 25.28. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-1r-1p-2; metatarsus v2-1p-1r. II – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-1p-1r. III – femur d1-1-1, p0-1-1, r0-1-1; tibia v2-1r-1p, r0-1-1, p0-1-1; metatarsus p0-1-0, r1-1-0, v2-1p-1r-1. IV – femur d1-1-1, r0-0-1, p0-0-1; tibia r0-1-1, v2-0-2; metatarsus p0-1-0, r1-1-0, v2-1p-1r-1.

Epigynum: epigynal plate projected posteriorlly; CO with posterior border recurved; CD visible ventrally, sinuous; SS irregular, collapsed in our preparations; PVP enveloping partially the PS but not enveloping SS (Figs. 65–66).

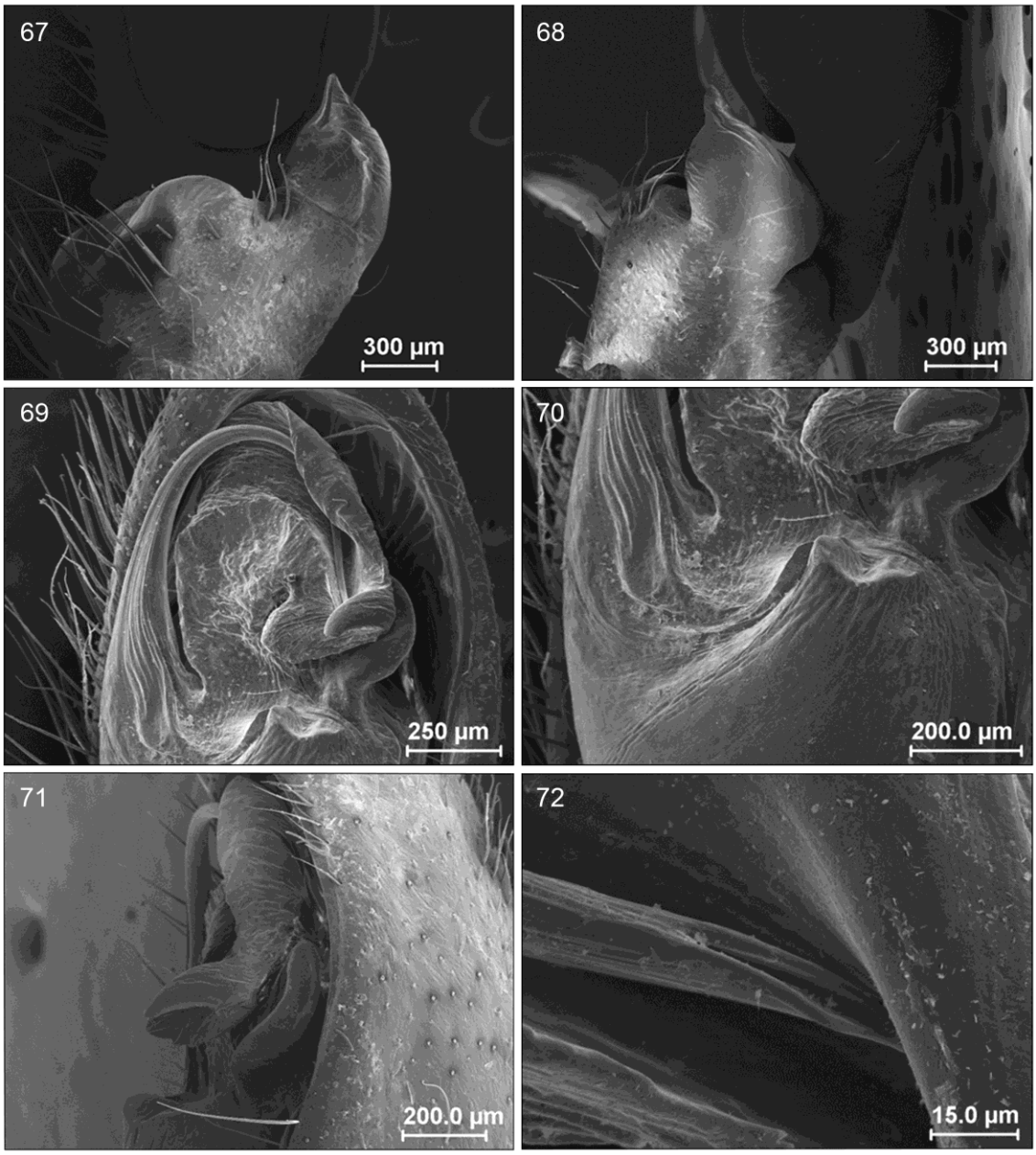
Variation. Length (10 males): total 11.4–14.5; carapace 5.6–6.9; femur I 5.5–6.88; (4 females) total 12.6–16.5; carapace 6.7–7.4, femur I 6.5–6.95. The specimens from Piauí are larger and darker than the ones from other locations.

Distribution. Piauí, Rio Grande do Norte, Paraíba and Bahia, Brazil.

Other material examined. BRAZIL. 4 males, R. Ihering MNRJ (41992). **Piauí:** Castelo do Piauí (ECB Rochas Ornamentais Ltda, Fazenda Bonito), [05º12'42"S 41º42'13"W], 1 male, F.M. Oliveira-Neto (CHNUFPI 0156); 1 male (CHNUFPI 0157); Caracol (Parque Nacional da Serra das Confusões), 09º13'16"S 43º29'21"W, 1 male, X.2006, P.R.R Silva et. al. (CHNUFPI 0165); 1 macho (CHNUFPI 0166);. **Rio Grande do Norte:** João Câmara, 05º32'15"S 35º49'11"W, 1 female, XI.2011, N. Sebastian et. al. (MPEG 19965); 1 male, VII.2012, N. Sebastian et. al. (MPEG 19966); 1 male, VI.2012, N. Sebastian et. al. (MPEG 19967); 1 male, VI.2012, N. Sebastian et. al. (MPEG 19968); 1 male, VI.2012, N. Sebastian et. al. (MPEG 19969); **Paraíba: [**7°14'23.86"S 36°46'55.02"W]**,** 1 male, R. Ihering (MNRJ 42241).



FIGURES 63–66. *Corinna caatinga* n. sp., male: 63) palp, ventral; 64) same, retrolateral; female: 65) epigynum, ventral; 66) same, dorsal. Scale bar = 0.5 mm. Abbreviations: CD–Copulatory ducts; Cpr–Cymbial projection; SS–Secondary spermathecae; TPC–Tegular process of *Corinna*.



FIGURES 67–72. *Corinna caatinga* n. sp., male: 67) Retrolateral tibial apophysis, ventral; 68) same, retrolateral; 69) bulb apex, ventral; 70) Tegular process, ventral; 71) conductor, retrolateral; 72) Apex of embolus.

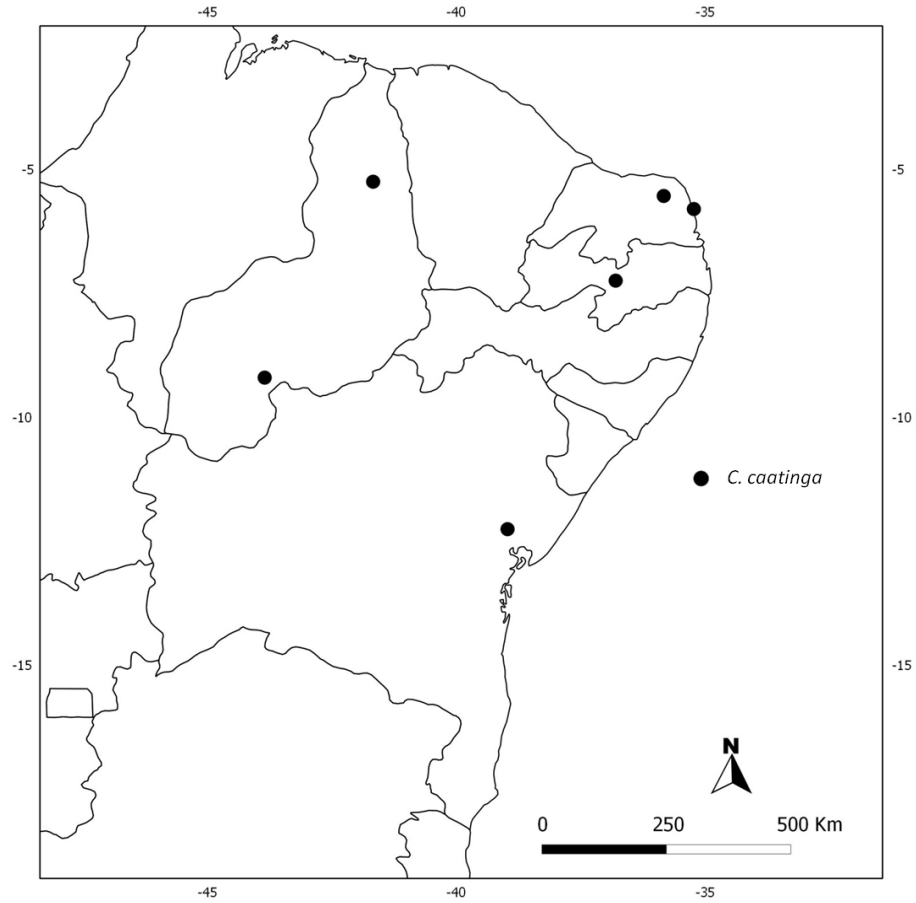


FIGURE 73. Known distribution records of *Corinna caatinga* n. sp. in Northeastern Brazil.

***Corinna vesperata* n. sp.**

Figs. 38, 74–77, 82

Type Material. Male holotype from Minas da Serrinha, Diamantina [18°14'17.05"S 43°36'39.64"W], Minas Gerais, Brazil, XII.1944, E. Cohn, deposited in AMNH. Paratypes: Brazil. Minas Gerais: Diamantina [18°14'17.05"S 43°36'39.64"W], (Minas da Serrinha), 1 female, XII.1944, E. Cohn (AMNH); 1 male, I-III.1945 (AMNH).

Etymology. The specific name refers to the cultural event called "vesperata", that occurs twice a month from March to October in Diamantina, the type locality. In a

typical vesperata presentation, the musicians perform from the balconies and windows of historic buildings and are conducted by a maestro in the middle of the street.

Diagnosis. Males of *Corinna vesperata* n. sp. resemble those of *C. hyalina* n. sp. by the retrolaterally enlarged RTA but differ by the wider than long RTA in retrolateral view (Figs. 74–75); females can be identified by the posteriorly placed copulatory opening, inserted in an epigynal excavation (Figs. 76–77).

Description. Male (holotype). Carapace dark reddish brown, chelicerae black, endites and labium dark red with posterior end paler, sternum brown, leg I with coxae, trochanter, femur, metatarsus and tarsus dark brown, patella and tibia dark yellow, legs II, III and IV dark yellow; dorsum of abdomen gray, with dorsal scutum extending to the median region of abdomen, ventrally pale gray. Total length: 11.7. Carapace 6.2 long; 5.0 wide; 3.3 high. Clypeus 0.6 high. Abdomen 5.6 long; 3.9 wide. Eyes: anterior row 2.1, posterior row 2.5. MOQ: 0.82 long, 0.87 anterior wide, 0.85 posterior wide. Eye diameters and interdistances: AME 0.28; ALE 0.22; PME 0.25; PLE 0.21; AME–AME 0.35; AME–ALE 0.41; PME–PME 0.42; PME–PLE 0.72; ALE–PLE 0.15. Chelicerae 3.4 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 3.1 long; 2.7 wide. Leg measurements: femur I 5.3/ patella 2.3/ tibia 5.1/ metatarsus 4.0/ tarsus 2.2/ total 18.9; II 4.9/ 2.4/ 4.3/ 3.6/ 2.1/ 17.3; III 4.1/ 2.0/ 3.2/ 3.5/ 1.8/ 14.6; IV 5.5/ 2.2/ 4.9/ 5.2/ 2.0/ 19.8. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-2; tibia v2-2-2; metatarsus v2-2-0. II – femur d1-1-1, p0-0-2; tibia v1-1-2; metatarsus v2-2-0. III – femur d1-1-1, p0-1-1, r0-1-1; tibia vp1-0-1, vr0-1-0, p0-1-0, r0-1-1; metatarsus v2-2-1, p0-1-0, r1-1-0. IV – femur d1-1-1, r0-0-1, p0-0-1; tibia v2-2-0, r1-0-1; metatarsus v2-2-1, p1-0-0, r1-1-0. Palp: femur with two posterior dorsal spines; TPr retroapical curved slightly prolateraly; TPC placed far from base of embolus; E filiform partially covered by distal fold of conductor; C with small distal unsclerotized area (Figs. 74–75).

Female (paratype AMNH). Coloration as in male. Total length: 12.7. Carapace 6.4 long; 5.1 wide; 3.25 high. Clypeus 0.5 high. Abdomen 5.9 long; 4.3 wide. Eyes: anterior row 2.3, posterior row 2.6. MOQ: 0.9 long, 1.0 anterior wide, 0.95 posterior wide. Eye diameters and interdistances: AME 0.34; ALE 0.25; PME 0.25; PLE 0.23; AME–AME 0.37; AME–ALE 0.46; PME–PME 0.45; PME–PLE 0.8; ALE–PLE 0.17. Chelicerae 3.6 long. Sternum 3.2 long; 2.6 wide. Leg measurements: femur I 4.8/ patella 2.5/ tibia 4.7/ metatarsus 3.9/ tarsus 2.2/ total 18.1; II 4.8/ 2.3/ 4.1/ 3.6/ 2.1/ 16.9; III 4.2/ 2.0/ 3.0/ 3.2/ 1.6/ 14.0; IV 4.3/ 2.2/ 4.5/ 4.2/ 2.0/ 17.2. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia v1-1-2; metatarsus v2-2-0. III – femur d1-1-1, r0-1-0, p0-1-1; tibia v2-2-0, r1-0-1, p0-1-0; metatarsus p1-1-0, r0-1-0, v2-2-0. IV – femur d1-1-0, r0-0-1; tibia r1-0-1, v2-1r-1p; metatarsus p1-1-0, r0-1-0, v1r-2-1. Epigynum: epigynal plate not projected posteriolly; CO inserted in an excavation, posterior border V-shaped; CO conspicuous; CD not visible ventrally; SS placed very far from each other; small, nearly same sized as PS; PVP enveloping partially the PS and SS (Figs. 76–77).

Variation. Length (2 males): total 11.7–12.1, carapace 6.2–6.3, femur I 5.3–5.6.

Distribution. Known only from the type locality.

Other material examined. None.

***Corinna hyalina* n. sp.**

Figs. 78–79, 82

Type Material. Male holotype from Parque Nacional de Itatiaia, Itatiaia, Rio de Janeiro, Brazil, 22º27'17"S 44º36'29.8"W, 15–22.II.2011, G.H.F. Azevedo et. al., deposited in UFMG (9701). Paratypes: Brazil. Minas Gerais: Santana do Riacho

(Parque Nacional da Serra do Cipó), 19°21'S 43°31'W, 1 male, 10–14.II.2001, E.S.S. Álvares & E.O. Machado (UFMG 1078).

Etymology. The specific name is an adjetive referring to the small hyaline structure in apex of the retrolateral tibial apophysis.

Diagnosis. Males of *Corinna hyalina* n. sp. resemble those of *C. vesperata* n. sp. by the retrolaterally enlarged RTA but differ by presenting a small hyaline process in the apex of the RTA; embolus very short and thin, conductor relatively small and tegular process very close to base of embolus (Figs. 78–79).

Description. Carapace and chelicerae black, endites red with posterior end paler; labium black with posterior end paler; sternum brown, leg I brown, legs II, III, IV yellow except metatarsus and tarsus, dark yellow; dorsum of abdomen gray with dorsal scutum occupying distal third, ventrally pale gray. Total length: 10.0. Carapace 4.3 long; 4.1 wide; 2.5 high. Clypeus 0.43 high. Abdomen 5.6 long; 4.0 wide. Eyes: anterior row 1.8, posterior row 2.2. MOQ: 0.66 long, 0.71 anterior wide, 0.73 posterior wide. Eye diameters and interdistances: AME 0.25; ALE 0.21; PME 0.21; PLE 0.23; AME–AME 0.26; AME–ALE 0.32; PME–PME 0.32; PME–PLE 0.56; ALE–PLE 0.14. Chelicerae 2.7 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.5 long; 2.15 wide. Leg measurements: femur I 4.55/ patella 2.0/ tibia 4.2/ metatarsus 3.7/ tarsus 2.05/ total 16.5; II 4.2/ 2.0/ 3.7/ 3.5/ 1.9/ 15.3; III 3.2/ 1.7/ 2.35/ 2.95/ 1.3/ 11.5; IV 4.2/ 2.0/ 3.9/ 3.9/ 1.4/ 15.4. Leg formula 1243. Leg spination: I – femur d1-0-1, p0-0-1; tibia v1p-2-2-2-2-2; metatarsus v2-0-0. II – femur d1-0-1, p0-0-1; tibia v2p-1p-2-2; metatarsus v2-2-0. III – femur d1-1-0, p0-0-2; tibia v2-2-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-0-1, v1p-0-1p; metatarsus v1r-1p-1. Palp: femur with two posterior dorsal spines; RTA with rounded prolateral region and tapered ventral projection; T without projection; TPC placed very close to base of embolus; E

filiform, short, partially covered by distal groove of conductor; C small and subquadrangular (Figs. 78–79).

Female: Unknown.

Variation. Length (2 males): total 9.1–10.0; carapace 4.3–4.4; femur I 4.1–4.55.

Distribution. Minas Gerais and Rio de Janeiro, Brazil.

Other material examined. None

***Corinna tranquilla* n. sp.**

Figs. 80–82

Type Material. Female holotype from RPPN Estação Biológica da Mata do Sossego, 20°04'25.5"S 42°04'13"W, Simonésia, Minas Gerais, Brazil, 28–30.XI.2010, M. T. T. Santos, deposited in UFMG (4741).

Etymology. The specific name is a Latin adjective meaning quiet and was taken from the type locality name (Mata do Sossego), Portuguese for Forest of Tranquility.

Diagnosis. Females of *Corinna tranquilla* n. sp. are similar to those of *C. zecarioca* n. sp., by the separated secondary spermathecae, but are readily recognized by these spermathecae being ovoid instead globular (Figs. 80–81).

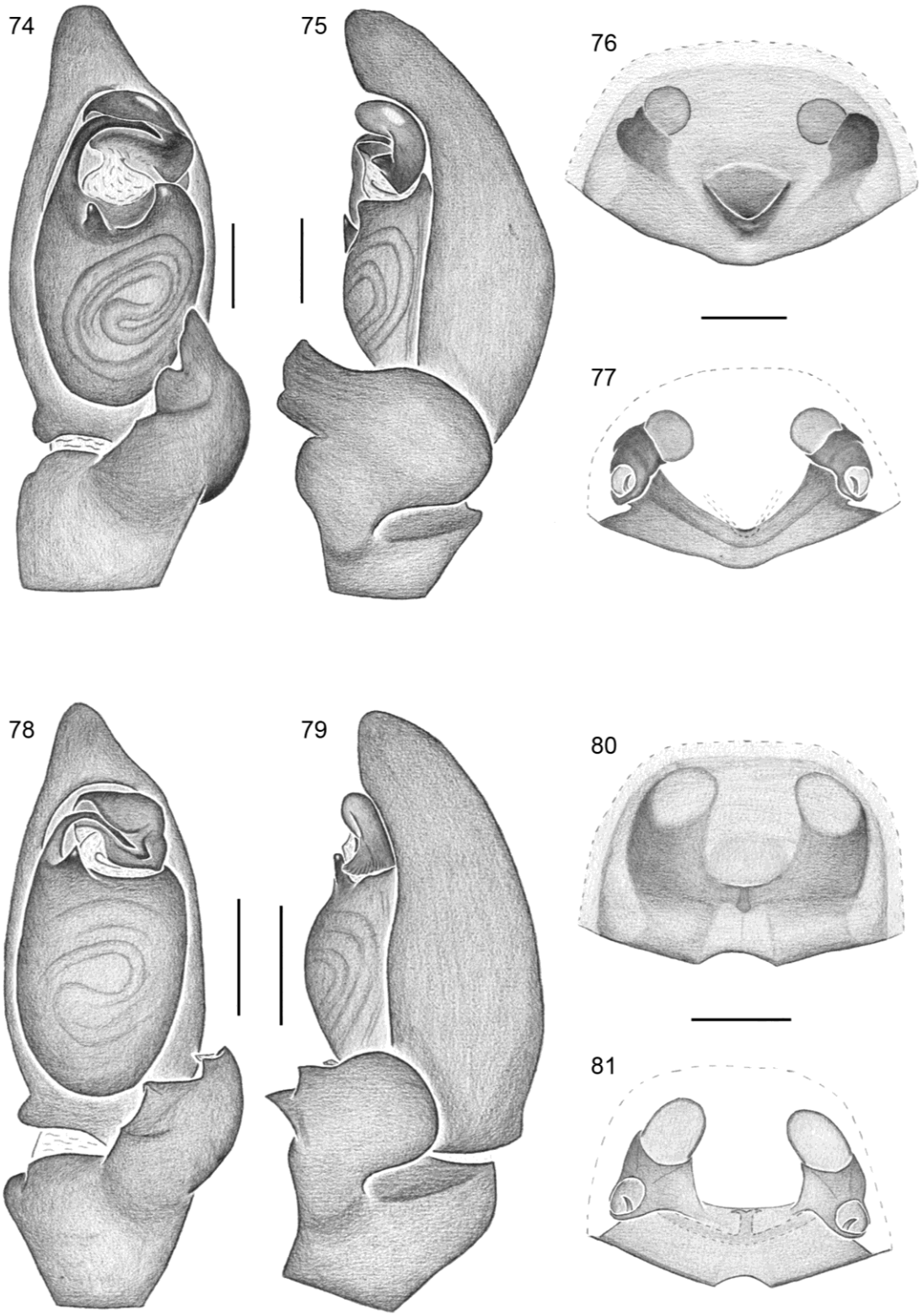
Description. Female (holotype). Carapace and chelicerae black, endites dark red and labium black with posterior end paler, sternum dark red, legs I and II dark red, legs III and IV dark yellow; dorsum of abdomen dark gray, ventrer gray. Total length: 11.6. Carapace 5.0 long; 3.9 wide; 2.7 high. Clypeus 0.55 high. Abdomen 6.3 long; 4.2 wide. Eyes: anterior row 2.05, posterior row 2.3. MOQ: 0.8 long, 0.85 anterior wide, 0.8 posterior wide. Eye diameters and interdistances: AME 0.33; ALE 0.25; PME 0.25; PLE 0.25; AME–AME 0.25; AME–ALE 0.38; PME–PME 0.3; PME–PLE 0.63; ALE–

PLE 0.18. Chelicerae 2.75 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.4 long; 2.1 wide. Leg measurements: femur I 4.1/ patella 1.8/ tibia 3.55/ metatarsus 2.95/ tarsus 1.7/ total 14.1; II 3.65/ 1.7/ 3.0/ 2.7/ 1.5/ 12.55; III 2.95/ 1.45/ 2.1/ 2.5/ 1.1/ 10.1; IV 4.05/ 1.7/ 3.5/ 3.75/ 1.2/ 14.2. Leg formula 4123. Leg spination: I – femur d1-0-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-1; tibia v2-2-0, p1-0-0, r0-1-1; metatarsus p1-0-0, r0-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1; tibia r0-1-1, v1p-2-0; metatarsus r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO median, inconspicuous; CD not visible ventrally; PVP enveloping partially the PS and SS (Figs. 80–81).

Male: Unknown.

Distribution. Known only from the type locality.

Other material examined. None.



FIGURES 74–81. *Corinna vesperata* n. sp., male: 74) palp, ventral; 75) same, retrolateral; female: 76) epigynum, ventral; 77) same, dorsal. *Corinna hyalina* n. sp., male: 78) palp, ventral; 79) same, retrolateral. *Corinna tranquilla* n. sp., female: 80) epigynum, ventral; 81) same, dorsal. Scale bar = 0.5 mm.

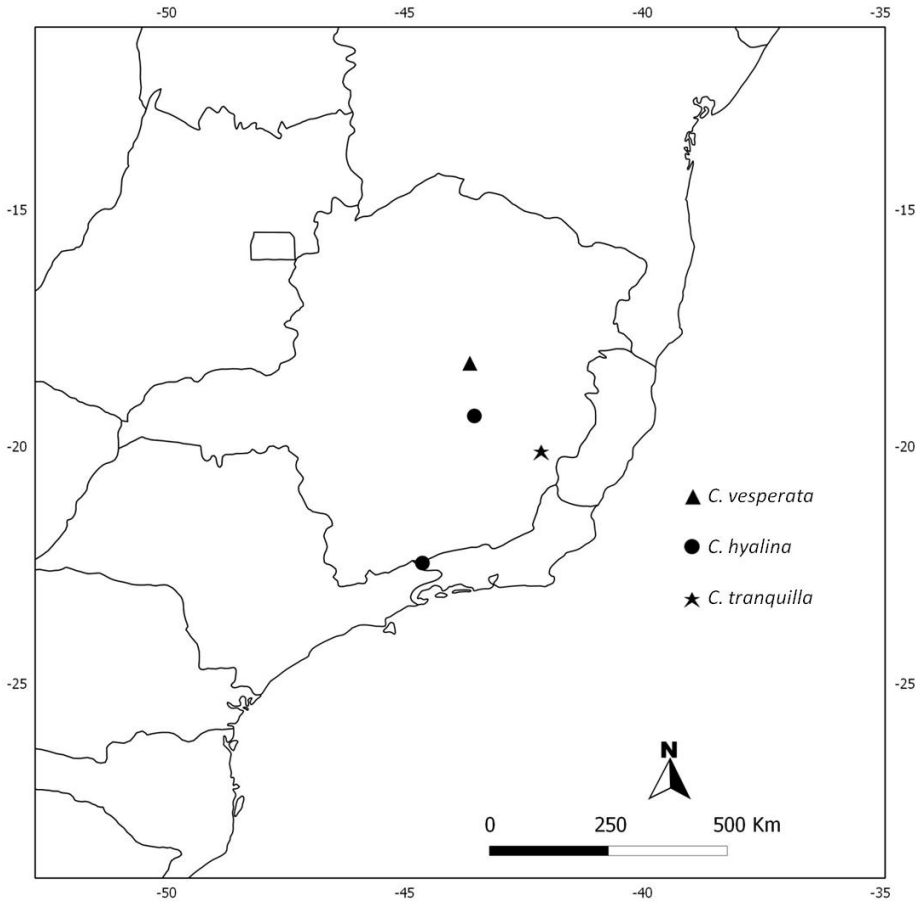


FIGURE 82. Known distribution records of *Corinna vesperata* n. sp., *C. hyalina* n. sp. and *C. tranquilla* n. sp. in Southeastern Brazil.

Corinna balacobaco **n. sp.**

Figs. 37, 83–86, 89

Type Material. Male holotype from Parque Estadual do Rio Doce, 19º39'31"S 42º34'32"W, Timóteo, Minas Gerais, Brazil, 20.VII.2010, B.T. Faleiro, deposited in UFMG (4931). Paratypes: Brazil. Minas Gerais: Timóteo **(**Parque Estadual do Rio Doce) 19º39'30.6"S 42º34'28.2"W, 1 female, 22.II.2011, B.T. Faleiro (UFMG 10695); 19º39'31"S 42º34'28"W, 1 female, 20.VII.2010, B.T. Faleiro (UFMG 4956).

Etymology. The specific name is a noun in apposition, from the Brazilian expression "do balacobaco". This expression means something excellent and problably had a Bantu origin.

Diagnosis. Males of *Corinna balacobaco* n. sp. differ from those of other species with a small median projection on prolateral surface of RTA by the tapered apex of RTA, embolus short, bent in right angle and by the strongly sclerotized conductor (Figs. 83–84); females are readly recognized by the copulatory opening inserted in a ventral triangular epigynal projection (Fig. 85).

Description. Carapace black with posterior end of thoracic region dark reddish brown; chelicerae black, endites and sternum red, labium black with posterior end paler, legs I and II dark yellow, except metatarsus and tarsus, brown, legs III and IV yellow, except metatarsus and tarsus, brown; abdomen pale gray with dorsal scutum extended to the median region of abdomen. Total length: 11.2. Carapace 5.6 long; 4.55 wide; 2.9 high. Clypeus 0.5 high. Abdomen 5.2 long; 3.7 wide. Eyes: anterior row 2.0, posterior row 2.4. MOQ: 0.74 long, 0.75 anterior wide, 0.75 posterior wide. Eye diameters and interdistances: AME 0.25; ALE 0.2; PME 0.2; PLE 0.18; AME–AME 0.27; AME–ALE 0.41; PME–PME 0.36; PME–PLE 0.71; ALE–PLE 0.17. Chelicerae 3.1 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.8 long; 2.2 wide. Leg measurements: femur I 4.7/ patella 2.1/ tibia 4.25/ metatarsus 3.8/ tarsus 1.85/ total 16.7; II 4.35/ 2.05/ 3.6/ 3.5/ 1.15/ 14.65; III 3.5/ 1.6/ 2.5/ 3.1/ 1.3/ 12; IV 4.6/ 1.65/ 3.9/ 4.3/ 1.5/ 15.95. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vp1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-0-1; tibia v2-2-0, r0-1-1; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v2-1r-1p; metatarsus p0-1-0, r0-1-0, v2-2-1. Palp:

femur with three posterior dorsal spines; T without projection; TPC placed far from base of embolus; E filiform partially covered by conductor (Figs. 83–84).

Female (paratype UFMG 4956). Coloration as in male. Total length 14.3. Carapace 6.0 long; 4.8 wide; 3.0 high. Clypeus 0.5 high. Abdomen 8.0 long; 5.2 wide. Eyes: anterior row 2.25, posterior row 2.7. MOQ: 0.72 long, 0.8 anterior wide, 0.85 posterior wide. Eye diameters and interdistances: AME 0.25; ALE 0.2; PME 0.2; PLE 0.18; AME–AME 0.3; AME–ALE 0.5; PME–PME 0.45; PME–PLE 0.82; ALE–PLE 0.15. Chelicerae 3.5 long. Sternum 2.8 long; 2.5 wide. Leg measurements: femur I 5.0/ patella 2.35/ tibia 4.2/ metatarsus 3.5/ tarsus 1.85/ total 16.9; II 4.25/ 2.15/ 3.5/ 3.3/ 1.75/ 14.95; III 3.5/ 1.65/ 2.55/ 3.0/ 1.5/ 12.2; IV 4.5/ 2.0/ 3.5/ 4.2/ 1.5/ 15.7. Leg formula 1423. Leg spination: I – femur d1-0-1, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-0, p0-0-1; tibia v2-2-0, r0-1-1; metatarsus p0-1-0, r1-0-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v2-2-0; metatarsus r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO anteriorly placed between SS; anterior border of copulatory opening conspicuous; CD not visible ventrally; SS closely placed in relation to each other; PVP enveloping partially PS and SS (Figs. 85–86).

Variation. Length (2 females): total 10.9–14.3; carapace 5.3–6.0; femur I 4.7–5.0.

Distribution. Known only from the type locality.

Other material examined. None.

***Corinna vilanovae* n. sp.**

Figs. 87–89

Type Material. Male holotype from Parque Nacional da Serra dos Orgãos, 22°27'23.4"S 42°59'41.4"W, Teresópolis, Rio de Janeiro, Brazil, 22–25.I.2011, G.H.F. Azevedo, deposited in UFMG (10189).

Etymology. The specific name is a patronim in honor of first author's friend Ellen Caroline Couto Vilanova.

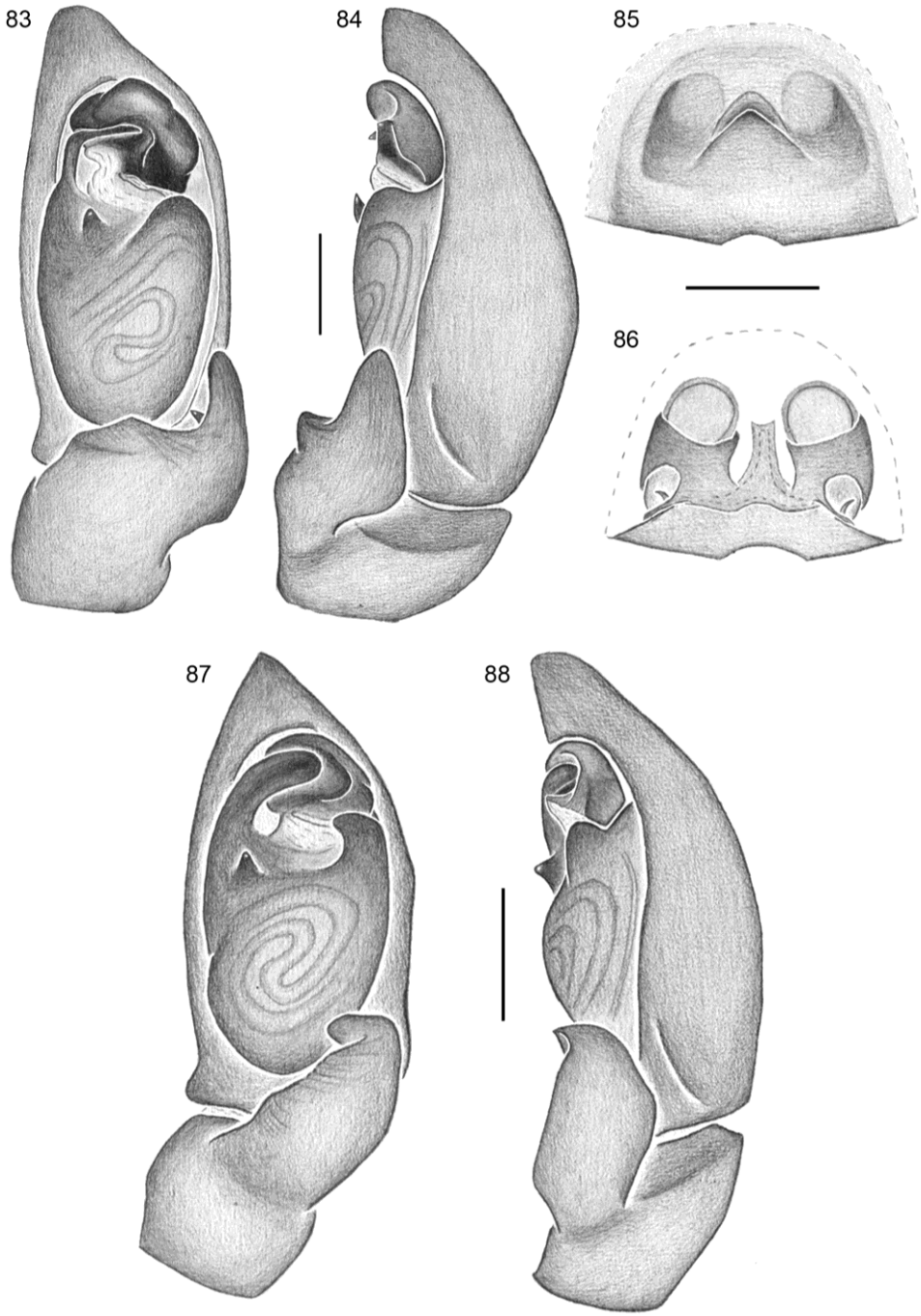
Diagnosis. Males of *Corinna vilanovae* n. sp. are readily recognized by the stout, spatulated embolus, tegulum with well-developed retroapical projection and apex of retrolateral tibial apophysis directed prolaterally (Figs. 87–88).

Description. Male (holotype). Carapace black, with posterior end of thoracic region dark red, chelicerae black, endites red and labium black, with posterior end paler, sternum brown, leg I dark red, legs II, III and IV dark yellow; dorsum of abdomen dark gray, ventrally gray. Total length 7.5. Carapace 4.0 long; 3.25 wide; 2.3 high. Clypeus 0.27 high. Abdomen 3.4 long; 2.46 wide. Eyes: anterior row 1.5, posterior row 1.8. MOQ: 0.57 long, 0.66 anterior wide, 0.63 posterior wide. Eye diameters and interdistances: AME 0.24; ALE 0.12; PME 0.21; PLE 0.15; AME–AME 0.18; AME–ALE 0.24; PME–PME 0.27; PME–PLE 0.45; ALE–PLE 0.12. Chelicerae 2.2 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.0 long; 1.75 wide. Leg measurements: femur I 3.4/ patella 1.5/ tibia 3.0/ metatarsus 2.6/ tarsus 1.6/ total 12.1; II 3.1/ 1.4/ 2.5/ 2.3/ 1.5/ 10.8; III 2.6/ 1.3/ 1.7/ 2.2/ 1.2/ 9.0; IV 3.4/ 1.4/ 2.7/ 2.9/ 1.3/ 11.7. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v1p-2-1r; metatarsus v2-0-2. II – femur d1-1-0; tibia v0-1r-1r; metatarsus v2-0-2. III – femur d1-1-1; tibia v0-2-0, r1-0-0, p0-1-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-1; tibia r0-1-1, v1p-1p-0; metatarsus r0-1-0, v2-2-1. Palp: femur with two posterior dorsal spines; TPC placed far from base of embolus; E stout and spatulated, not covered by folds of conductor (Figs. 87–88).

Female: Unknown.

Distribution. Known only from the type locality.

Other material examined. None.



FIGURES 83–88. *Corinna balacobaco* n. sp., male: 83) palp, ventral; 84) same, retrolateral; female: 85) epigynum, ventral; 86) same, dorsal. *Corinna vilanovae* n. sp., male: 87) palp, ventral; 88) same, retrolateral. Scale bar = 0.5 mm.

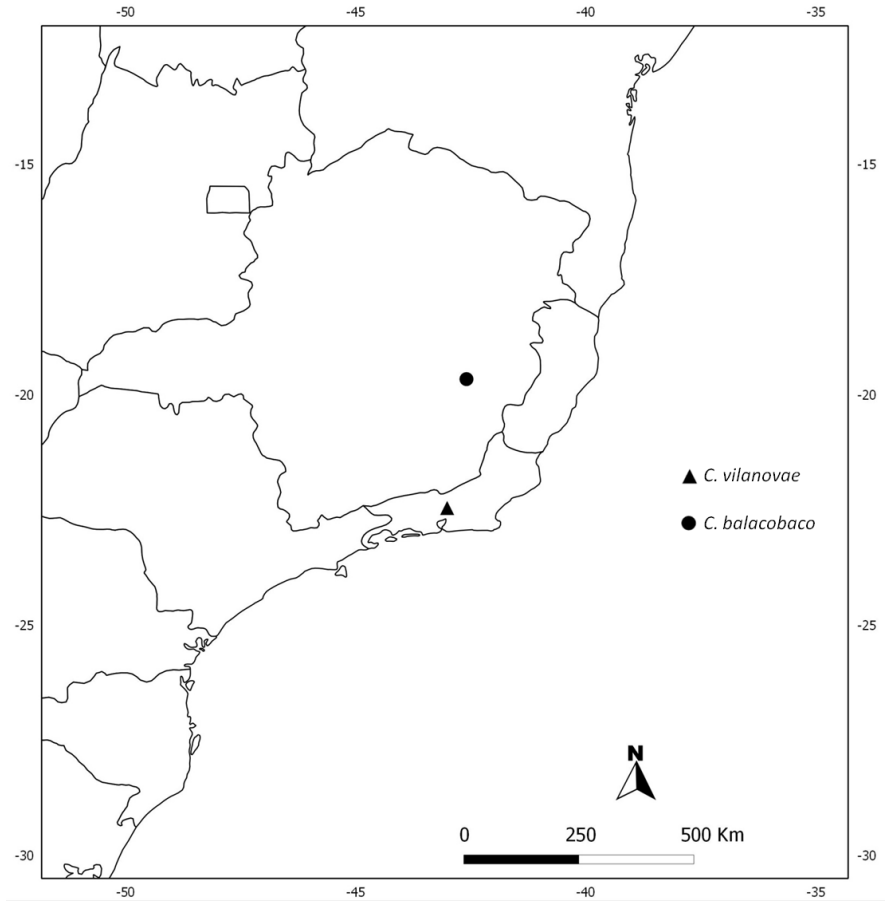


FIGURE 89. Known distribution records of *Corinna balacobaco* n. sp., and *C. vilanovae* n. sp. in Southeastern Brazil.

***Corinna demersa* n. sp.**

Figs. 17, 28–30, 39, 90–93, 98–104

Type Material. Male holotype from Ilha do Cardoso [25°7'54.00"S 47°57'59.00"W], Cananéia, São Paulo, Brazil, II.2008, G.C.O. Piccoli & G.Q. Romero, deposited in (MPEG 20254). Paratypes: Brazil. Bahia: Maracás [13°25'41.94"S 40°26'16.03"W] (Fazenda do Mato), 1 female, 16.II.1967, U. Caramachi et. al. (MNRJ 06435); Espírito Santo: Guarapari (Restinga de Setiba), 20°36'0"S 40°25'0"W, 1 male and 2 females, 28–04.IX–X.2009, T. Gonçalves Souza (UFMG 7188); Rio de Janeiro: Cabo Frio

[22°52'43.26"S 42° 1'11.55"W] (Figueira), 1 male, 2.I.2003, (MNRJ 06436); São Paulo: São Paulo (Ilhabela, Ilha do Búzios) [23°48'21.84"S 45° 8'41.18"W], 1 male and 1 female, X.1963, exped. DZ (MZUSP 12620); Cananéia (Ilha do Cardoso) [25°7'54.00"S 47°57'59.00"W], 1 females, II.2008, G.C.O. Piccoli & G.Q. Romero (MPEG 20255); 4 females (MPEG 20256); Rio Grande do Sul: Torres [29°20'20.81"S 49°43'37.32"W], 1 female, 09.X.1972, G. Carvalho (MCN 748).

Etymology. The specific name is a Latin adjective meaning submerged and refers to the microhabitat in which the holotype was collected, immersed in the water reservoir of a bromelid.

Diagnosis. Males of *Corinna demersa* n. sp. resemble those of *C. maracas* n. sp. by the RTA with a ventro-apical incision, absence of tegular process and presence of a well-delimited unsclerotized area in the distal retrolateral margin of conductor; differ by the RTA's ventro-apical incision deeper and by the thin, long embolus, partially covered by distal fold of conductor (Figs. 90–91, 99, 101–102); females resemble those of *C. maracas* n. sp. by the triangular copulatory opening, arising between the closely placed secondary spermathecae, but differ by the long, inverted T-shaped copulatory ducts visible ventrally and by the epigynal surface not grooved (Figs. 92–93, 103).

Description. Male (holotype). Carapace and chelicerae black, endites and labium black with posterior end paler, sternum dark red, leg with coxae, trochanter and femur dark brown, patella, tibia, metatarsus and tarsus dark yellow; dorsum of abdomen dark gray with scutum dorsal in distal third, ventrally brown, with large median gray band in longitudinal axis. Total length 11.1. Carapace 5.4 long; 5.1 wide; 2.2 high. Clypeus 0.4 high. Abdomen 5.6 long; 3.6 wide. Eyes: anterior row 2.0, posterior row 2.4. MOQ: 0.77 long, 0.92 anterior wide, 0.9 posterior wide. Eye diameters and interdistances: AME 0.3; ALE 0.22; PME 0.26; PLE 0.24; AME–AME 0.24; AME–ALE 0.38; PME–

PME 0.34; PME–PLE 0.56; ALE–PLE 0.18. Chelicerae 3.0 long; 5 retromarginal teeth and 3 promarginal teeth. Sternum 2.9 long; 2.5 wide. Leg measurements: femur I 5.1/ patella 2.4/ tibia 5.1/ metatarsus 4.1/ tarsus 1.8/ total 18.5; II 4.6/ 2.3/ 4.1/ 3.9/ 1.5/ 16.4; III 4.0/ 1.9/ 3.4/ 3.7/ 1.5/ 14.5; IV 5.2/ 2.0/ 4.9/ 5.5/ 1.8/ 19.4. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia v2-1p-2-2-0; metatarsus v2-2-0. III – femur d1-1-1, p0-1-2, r0-1-0; tibia v2-2-0, r0-1-1, p0-1-0; metatarsus p0-1-0, r1-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1, p0-0-1; tibia r0-1-1, v2-2-0; metatarsus p0-1-0, r1-1-0, v2-2-1. Palp: femur with two posterior dorsal spines; T without projection; TPC absent; E thin and long, partially covered by conductor (Figs. 90–91, 99, 101–102).

Female (paratype MPEG 20255). Coloration as in male. Total length 16.0. Carapace 6.3 long; 5.8 wide; 3.3 high. Clypeus 0.4 high. Abdomen 9.3 long; 5.9 wide. Eyes: anterior row 2.45, posterior row 2.8. MOQ: 0.88 long, 0.92 anterior wide, 0.94 posterior wide. Eye diameters and interdistances: AME 0.32; ALE 0.24; PME 0.26; PLE 0.26; AME–AME 0.3; AME–ALE 0.44; PME–PME 0.42; PME–PLE 0.7; ALE–PLE 0.18. Chelicerae 3.9 long. Sternum 3.4 long; 2.9 wide. Leg measurements: femur I 5.3/ patella 2.5/ tibia 5.2/ metatarsus 4.4/ tarsus 1.9/ total 19.3; II 4.8/ 2.3/ 4.2/ 4.0/ 1.6/ 16.9; III 4.3/ 2.1/ 4.0/ 3.7/ 1.6/ 15.7; IV 5.4/ 2.1/ 4.9/ 5.7/ 1.9/ 20.0. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-2-1p-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia v1p-2-2-2; metatarsus v2-2-0. III – femur d1-1-1, r0-1-0, p0-1-1; tibia v2-2-0, r0-1-1, p0-1-0; metatarsus p1-1-0, r1-1-0, v2-2-1. IV – femur d1-1-1, r0-0-1, p0-0-1; tibia r0-1-1, v2-2-0; metatarsus p1-1-0, r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO anterior, leveled with apices of SS; CO conspicuous; SS big, close to each other; PVP poor developed, linked at basal third of SS and enveloping partially the PS (Figs. 92–93, 103).

Variation. Length (7 males): total 11.1–14.1; carapace 5.3–6.4; femur I 4.6–5,6; (10 females) total 13.0–16.1; carapace 5.3–6.6; femur I 4.5–5.5.

Distribution. Bahia, Espírito Santo, Rio de Janeiro, São Paulo and Rio Grande do Sul, Brazil.

Other material examined. Brazil. **Bahia:** Nova Viçosa, [17°52'19.76"S 39°23'8.12"W], 1 male and 1 female, 23.III.1993, C.B. Castro & P.S. Young (MNRJ); **Espírito Santo:** Santa Tereza (Estrada Santa Tereza - Fundão), [19°56'13.23"S 40°28'44.03"W], 1 female, N.D. Santos and L.F. Reis (MNRJ); Presidente Kennedy (Praia das Neves), 21°14'10,3"S 40°58'33"W, 1 female, 5–8.X.2009, T. Gonçalves Souza (UFMG 8182); **Rio de Janeiro:** Vila da Penha, [22°50'39.08"S 43°18'45.23"W], 2 males and 1 female, N.O. dos Santos (MNRJ); Distrito Federal [actual Rio de Janeiro city 22°54'12.74"S 43°12'34.51"W], 2 females, 22.VI. 1939, A. Passarelli and A. Carvalho (MNRJ); Maricá, 1 male and 1 female, (MNRJ); Rio de Janeiro, (Recreio dos Bandeirantes), [23°1'36.74"S 43°28'9.81"W], 2 female (MNRJ 06446); Itauna, 1 female, 1.I.2003 (MNRJ 06437); **São Paulo:** Cananéia (Ilha do Cardoso), [25°7'54.00"S 47°57'59.00"W], 1 male, IV.2010, G.C. Piccoli (MPEG 20252); 1 male, G.Q. Romero (MPEG 20253).

***Corinna maracas* n. sp.**

Figs. 8, 13–15, 25, 94–97, 104

Type Material. Male holotype from Maracás [13°25'41.94"S 40°26'16.03"W], Bahia, Brazil, 22–25.VI.2009, A. Chagas Jr, A. Kury, D. Pedroso, A. Giupponi and V. Dill, deposited in MNRJ (6441). Paratypes: Brazil. Bahia: Maracás [13°25'41.94"S

40°26'16.03"W], 4 females, 22–25.VI.2009, A. Chagas Jr, A. Kury, D. Pedroso, A. Giupponi and V. Dill (MNRJ 6441); 1 female (MNRJ 6119).

Etymology. The specific name is a noun in apposition taken from the type locality.

Diagnosis. Males of *Corinna maracas* n. sp. differ from those of *C. demersa* n. sp. by the presence of a pointed retrolateral projection in the tegular apical margin and by the embolus not covered by folds of conductor (Figs. 94–95); females differ by the copulatory ducts not visible ventrally and by the epigynal surface with a wide longitudinal median groove (Fig. 96).

Description. Male (holotype). Carapace dark red, chelicerae black, endites red with posterior end paler, labium black with posterior end paler, sternum brown, leg I with coxae, trochanter and femur brown, patella, tibia, metatarsus and tarsus dark yellow, legs II with coxae and trochanter dark yellow, femur, patella, tibia, metatarsus and tarsus yellow, legs III and IV yellow; dorsum of abdomen gray, with dorsal scutum in distal third, ventrally pale gray, with large median brown band in longitudinal axis. Total length 15.2. Carapace 6.6 long; 5.6 wide; 1.45 high. Clypeus 0.55 high. Abdomen 6.8 long; 3.6 wide. Eyes: anterior row 2.25, posterior row 2.65. MOQ: 0.87 long, 0.82 anterior wide, 0.9 posterior wide. Eyediameters and interdistances: AME 0.3; ALE 0.25; PME 0.27; PLE 0.25; AME–AME 0.27; AME–ALE 0.45; PME–PME 0.37; PME–PLE 0.7; ALE–PLE 0.17. Chelicerae 3.4 long; 5 (6) retromarginal teeth and 3 promarginal teeth. Sternum 3.6 long; 2.9 wide. Leg measurements: femur I 6.1 / patella 2.9/ tibia 6.0/ metatarsus 5.5/ tarsus 2.0/ total 22.5; II 5.6/ 2.7/ 4.9/ 5.0/ 2.0/ 20.2; III 4.3/ 1.7/ 3.8/ 4.1/ 1.7/ 15.6; IV 5.1/ 2.3/ 5.3/ 5.7/ 1.9/ 20.3. Leg formula 1243. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia v2-1p-2-1r-1p; metatarsus v2-2-0. III – femur d0-1-0; tibia v0-0-2, r0-1-1, p0-0-1; metatarsus p1-1-0, r0-1-0, v2-2-2-1. IV – femur d0-1-0; tibia r0-1-1, v0-0-2; metatarsus

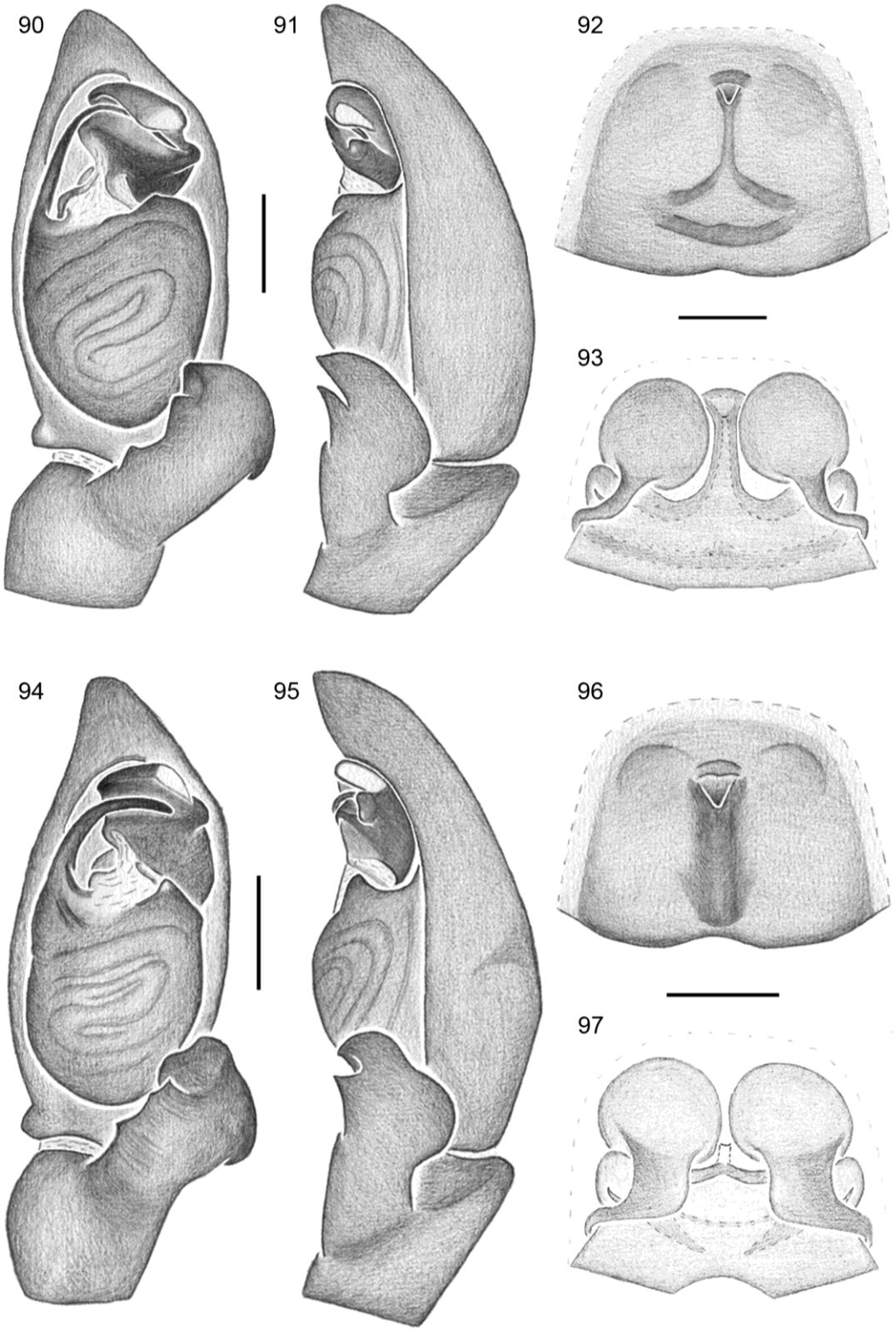
p1-3-0, r1-1-0, v2-0-2. Palp: femur with two posterior dorsal spines; RTA with ventro-apical incision; TPC absent; E filiform, not covered by a fold of conductor; C with unsclerotized area in the distal retrolateral margin (Figs. 94–95).

Female (paratype MNRJ 6441). Coloration as in male, except leg III and IV, brown.. Total length 13.7. Carapace 5.5 long; 5.0 wide; 2.4 high. Clypeus 0.32 high. Abdomen 7.8 long; 4.1 wide. Eyes: anterior row 2.1, posterior row 2.5. MOQ: 0.8 long, 0.8 anterior wide, 0.9 posterior wide. Eye diameters and interdistances: AME 0.3; ALE 0.27; PME 0.27; PLE 0.25; AME–AME 0.25; AME–ALE 0.4; PME–PME 0.35; PME–PLE 0.6; ALE–PLE 0.15. Chelicerae 3.4 long; 5 retromarginal teeth and 3 promarginal teeth. Sternum 3.1 long; 2.55 wide. Leg measurements: femur I 5.2/ patella 2.4/ tibia 4.9/ metatarsus 4.4/ tarsus 1.7/ total 18.6; II 4.7/ 2.3/ 4.0/ 4.0/ 1.6/ 16.6; III 4.1/ 1.7/ 3.4/ 3.9/ 1.4/ 14.5; IV 5.3/ 2.0/ 4.8/ 5.6/ 1.7/ 19.4. Leg formula 4123. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-1p-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia v2-1p-2-2; metatarsus v2-2-0. III – femur d1-1-1, r0-1-1, p0-0-1; tibia v2-0-2, r0-1-2, p0-1-0; metatarsus p2-1-0, r1-1-0, v2-2-2. IV – femur d1-1-1, r0-0-1, p0-1-1; tibia r0-1-1, v2-2-1; metatarsus p1-1-0, r1-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO anterior, leveled with base of SS; wide longitudinal groove from posterior border of copulatory opening to posterior end of epigynal plate; CO conspicuous; SS big, close to each other; PVP widely linked at basal third of SS enveloping partially the PS (Figs. 96–97).

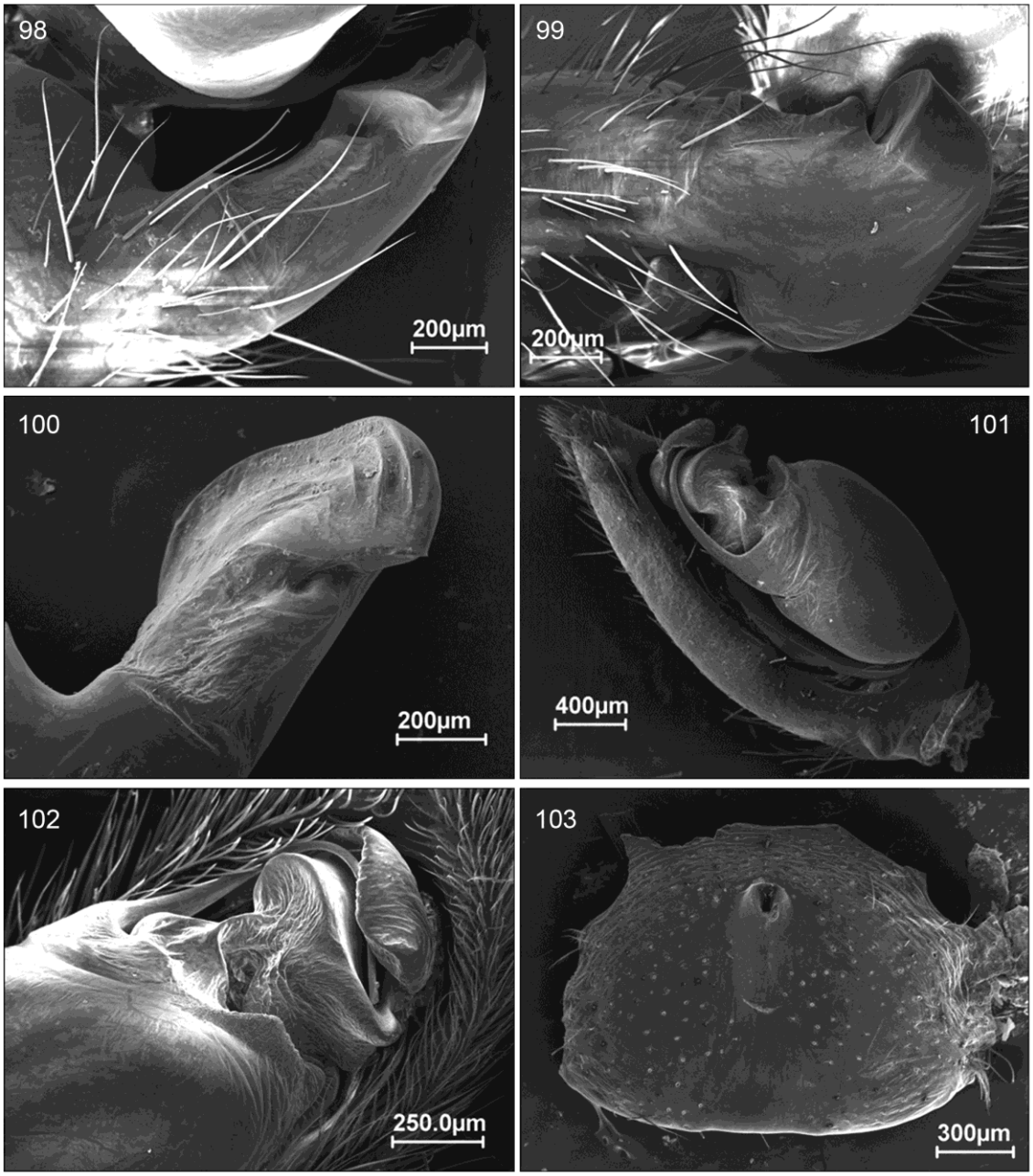
Variation. Length (5 females): total 13.7–16.5; carapace 5.5–6.2; femur I 5.0–5.3.

Distribution. Known only from the type locality.

Other material examined. None.



FIGURES 90–97. *Corinna demersa* n. sp., male: 90) palp, ventral; 91) same, retrolateral; female: 92) epigynum, ventral; 93) same, dorsal. *Corinna maracas* n. sp., male: 94) palp, ventral; 95) same, retrolateral; female: 96) epigynum, ventral; 97) same, dorsal. Scale bar = 0.5 mm.



FIGURES 98–103. *Corinna demersa* n. sp., male: 98) Retrolateral tibial apophysis, ventral; 99) same, retrolateral; 100) same, ventroprolateral; 101) palp, prolateral; 102) same, ventroretrolateral; female: 103) epigynum, ventral.

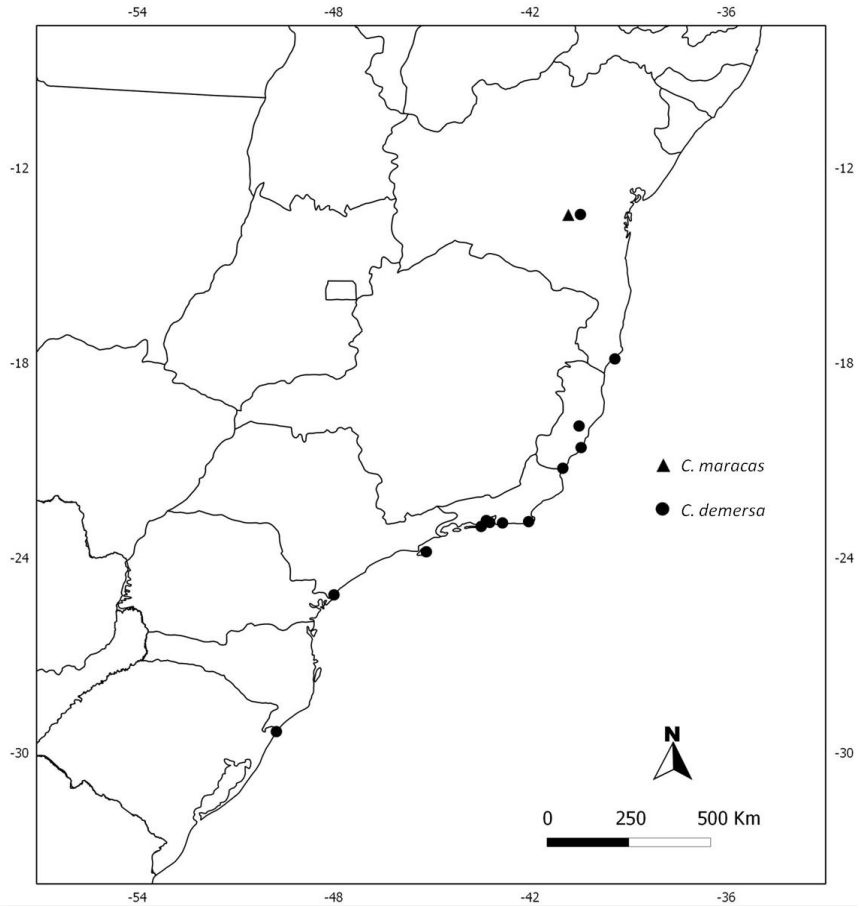


FIGURE 104. Known distribution records of *Corinna maracas* n. sp. and *C. demersa* n. sp. in Noertheastern, Southeastern and Southern Brazil.

***Corinna loiolai* n. sp.**

Figs. 105–106, 111

Type Material. Female holotype from Rio Grande, São Paulo, Brazil, 30.V.1951, V. Saar, deposited in IBSP (4083).

Etymology. The specific name is in honor of the first author's friend geologist Ignácio de Loiola Alvares Nogueira Neto. The holotype was collected in May, 30, the geologist day in Brazil.

Diagnosis. Females of *Corinna loiolai* n. sp. are similar to those of *C. regi* n. sp. by the copulatory opening delimited both anteriorly and posteriorly by well defined margins, but differ by the secondary spermatecae separated from each other and by the posterior margin of copulatory opening nearly straight (Fig. 105).

Description. Female (holotype). Carapace black with posterior end of thoracic region dark red; chelicerae black, endites red, labium red with posterior end paler, sternum brown, leg I with coxae, trochanter, femur and patella brown, tibia, metatarsus and tarsus dark red, legs II, III and IV dark yellow, except metatarsus and tarsus, brown; abdomen gray. Total length 11.1. Carapace 5.2 long; 4.0 wide; 3.0 high. Clypeus 0.4 high. Abdomen 5.9 long; 4.2 wide. Eyes: anterior row 2.1, posterior row 2.5. MOQ: 0.75 long, 0.83 anterior wide, 0.8 posterior wide. Eye diameters and interdistances: AME 0.18; ALE 0.18; PME 0.2; PLE 0.2; AME–AME 0.28; AME–ALE 0.45; PME–PME 0.4; PME–PLE 0.7; ALE–PLE 0.18. Chelicerae 2.95 long; 5 retromarginal teeth and 3 promarginal teeth. Sternum 2.5 long; 2.15 wide. Leg measurements: femur I 4.0/ patella 1.8/ tibia 3.4/ metatarsus 2.8/ tarsus 1.65/ total 13.65; II 3.6/ 1.7/ 3.0/ 2.7/ 1.55/ 12.55; III 3.0/ 1.5/ 2.15/ 2.5/ 1.0/ 10.15; IV 3.65/ 1.55/ 3.4/ 3.5/ 1.25/ 13.35. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v2-2-2-2; metatarsus v2-2-0. II – femur d1-1-0, p0-0-1; tibia vr1-1-1; metatarsus v2-2-0. III – femur d1-1-1, p0-1-0, r0-0-1; tibia v2-2-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0, r0-0-1; tibia r0-1-1, v1p-1r-1p; metatarsus r0-1-0, v2-1p-1. Epigynum: epigynal plate not projected posteriolly; CO medially placed, conspicuous; CD not visible ventrally; PVP with lateral folds projected laterally enveloping partially the PS and SS (Figs. 105–106).

Male: Unknown.

Distribution. Known only from the type locality.

Other material examined. None.

***Corinna regi* n. sp.**

Figs. 107–108, 111

Type Material. Female holotype from Parque Nacional da Serra do Cipó [19°12'31.53"S 43°46'59.87"W], Minas Gerais, Brazil, II.1987, deposited in MNRJ (1775). Paratype. Brazil. Minas Gerais: (Parque Nacional da Serra do Cipó), [19°12'31.53"S 43°46'59.87"W], 1 female, II.1987 (MNRJ 1775).

Etymology. The specific name is in honor to our friend and colleague, arachnologist Regiane Saturnino, nicknamed "Regi".

Diagnosis. Females of *Corinna regi* n. sp. differ from those of *C. loiolai* n. sp. by the secondary spermatecae touching each other and by the posterior margin of copulatory opening procurve (Fig. 107).

Description. Female (holotype). Carapace dark red with posterior end of thoracic region red, chelicerae black, endites and labium red with posterior end paler, sternum brown, leg I brown, leg II dark yellow, legs III and IV yellow; dorsum of abdomen gray, venter light gray. Total length 9.75. Carapace 4.4 long; 3.8 wide; 2.7 high. Clypeus 0.5 high. Abdomen 4.8 long; 3.2 wide. Eyes: anterior row 1.75, posterior row 2.12. MOQ: 0.75 long, 0.7 anterior wide, 0.72 posterior wide. Eye diameters and interdistances: AME 0.27; ALE 0.22; PME 0.2; PLE 0.17; AME–AME 0.22; AME–ALE 0.35; PME–PME 0.35; PME–PLE 0.6; ALE–PLE 0.13. Chelicerae 2.75 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.35 long; 2.1 wide. Leg measurements: femur I 4.0/ patella 1.75/ tibia 3.45/ metatarsus 2.85/ tarsus 1.65/ total 13.7; II 3.5/ 1.7/ 2.9/ 2.6/ 1.5/ 12.2; III 2.95/ 1.5/ 2.05/ 2.2/ 1.1/ 9.8; IV 3.9/ 1.65/ 3.25/ 3.4/ 1.15/ 13.35. Leg formula 1423. Leg spination: I – femur d1-1-0, p0-0-1; tibia v1p-2-2-2; metatarsus v2-1p-1r. II – femur d1-0-1; tibia vr1-1-1; metatarsus v2-1p-1r. III –

femur d1-1-1, p0-0-1, r0-1-0; tibia v2-2-0, r0-1-0; metatarsus p0-1-0, r0-1-0, v2-2-1. IV – femur d1-1-0; tibia r0-1-1, v1p-1r-1p; metatarsus r0-1-0, v2-2-1. Epigynum: epigynal plate not projected posteriolly; CO conspicuous medialy placed; CD not visible ventrally; PVP enveloping partially the PS and SS (Figs. 107–108).

Male. Unknown.

Variation. Length (2 females) total 9.65–9.75; carapace 4.4–4.55; femur I 4.0–4.05.

Distribution. Known only from the type locality.

Other material examined. None.

***Corinna kuryi* n. sp.**

Figs. 109–111

Type Material. Female holotype from RPPN Serra Bonita [15°23'S 39°33'W], Camacan, Bahia, Brazil, 11–13.VI.2009, A. Chagas Jr, A. Kury, D. Pedroso, A. Giupponi and V. Dill, deposited in MNRJ (6120).

Etymology. The specific name is a patronymic in honor of arachnologist Adriano Kury, one of the collectors of the holotype.

Diagnosis. Females of *Corinna kuryi* n. sp. resemble those of *C. rubripes* and *C. nitens* by the copulatory opening placed posteriorly to primary spermatecae and posterior margin of epigynum projected beyond the epigastric groove, but differ by the copulatory ducts not visible ventrally, by the copulatory opening placed in undifferentiated area and by the posterior epigynal margin medially truncated (Fig. 109).

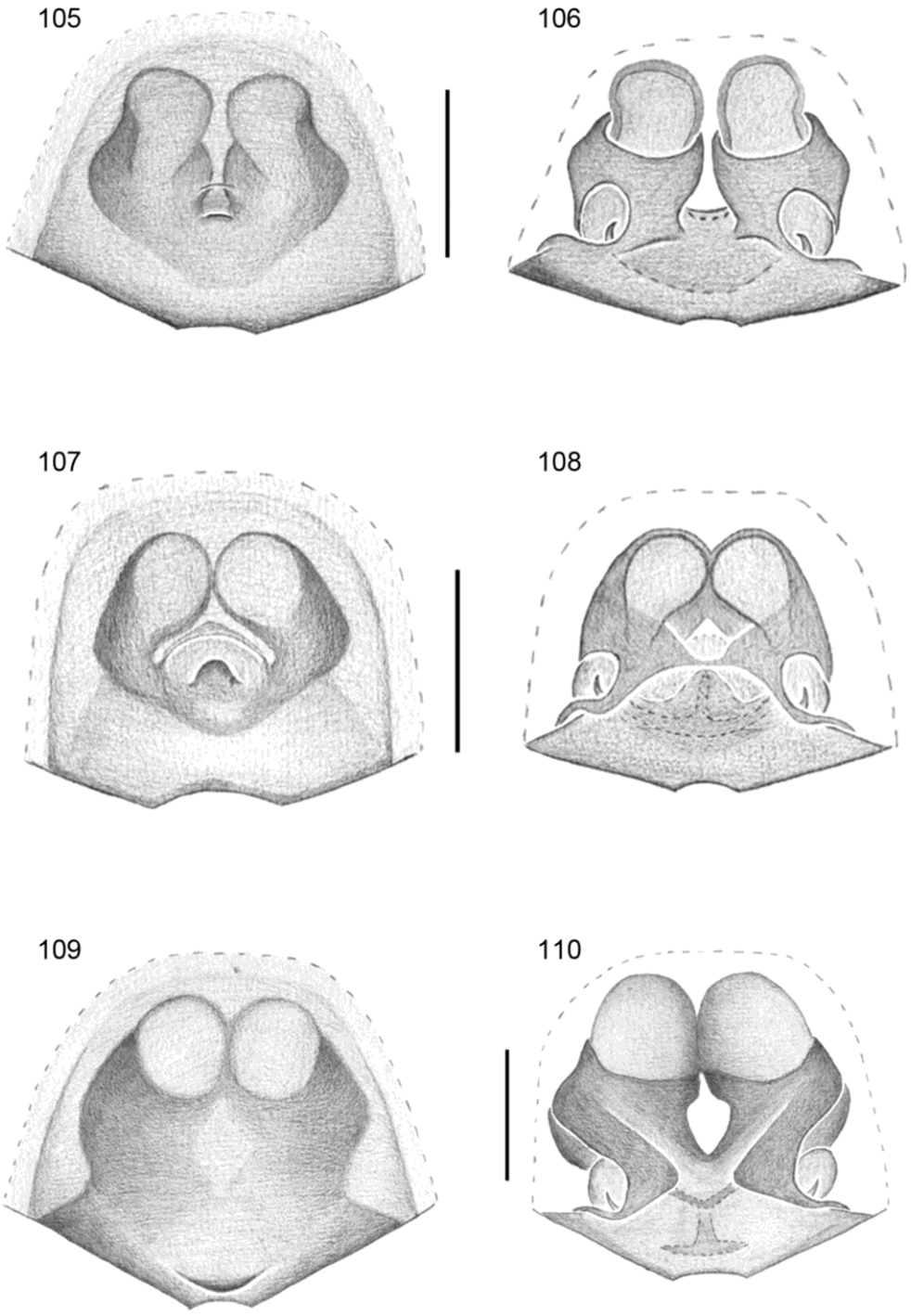
Description. Female (holotype). Carapace dark red with posterior end of thoracic region red, chelicerae black, endites red, labium red with posterior end paler, sternum

brown, legs yellow, with metatarsus and tarsus dark yellow; dorsum of abdomen gray, ventrally ligth gray with large median gray band in longitudinal axis. Total length: 11.4. Carapace 5.9 long; 4.8 wide; 3.3 high. Clypeus 0.53 high. Abdomen 5.2 long; 3.6 wide. Eyes: anterior row 2.1, posterior row 2.55. MOQ: 0.85 long, 0.88 anterior wide, 0.85 posterior wide. Eye diameters and interdistances: AME 0.33; ALE 0.25; PME 0.23; PLE 0.25; AME–AME 0.25; AME–ALE 0.28; PME–PME 0.35; PME–PLE 0.7; ALE–PLE 0.2. Chelicerae 3.1 long; 4 retromarginal teeth and 3 promarginal teeth. Sternum 2.9 long; 2.55 wide. Leg measurements: femur I 5.5/ patella 2.3/ tibia 5.4/ metatarsus 4.25/ tarsus 2.35/ total 19.8; II 5.1/ 2.2/ 4.25/ 3.9/ 2.05/ 17.5; III 4.1/ 1.9/ 2.95/ 3.45/ 1.55/ 13.95; IV 5.5/ 2.0/ 4.75/ 5.2/ 1.8/ 19.25. Leg formula 1423. Leg spination: I – femur d1-0-1, p0-0-1; tibia v2-1p-2-2; metatarsus v2-0-2. II – femur d1-0-1, p0-0-1; tibia v1r-2-2; metatarsus v2-1r-2. III – femur d1-1-1, p0-1-1, r0-1-1; tibia v2-0-2, p0-1-1, r0-1-1; metatarsus p1-1-0, r1-1-0, v2-1p-1r-1. IV – femur d1-1-1; tibia r0-1-1, v2-0-2; metatarsus r1-1-0, p0-1-0, v2-1p-1r-1. Epigynum: epigynal plate slightly projected posteriorlly; CO posterior conspicuous; SS touching each other; PVP well developed enveloping partially the PS and SS (Figs. 109–110).

Male: Unknown.

Distribution. Known only from the type locality.

Other material examined. None.



FIGURES 105–110. *Corinna* spp. female epigynum. *Corinna loiolai* n. sp.: 5), ventral; 106) dorsal. *Corinna regi* n. sp.: 107) ventral; 108) dorsal. *Corinna kuryi* n. sp.: 109) ventral; 110) dorsal. Scale bar = 0.5 mm.

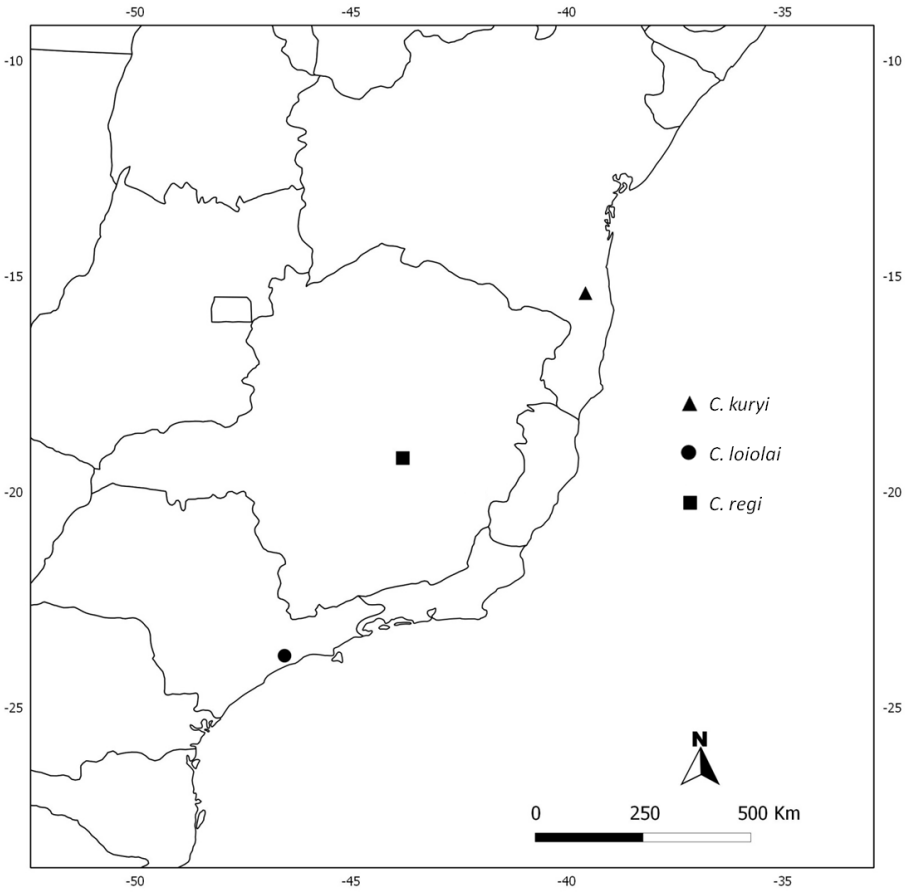


FIGURE 111. Known distribution records of *Corinna kuryi* n. sp., *C. loiolai* n. sp. and *C. regi* n. sp. in Northeastern and Southeastern Brazil.

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