

# Gastrointestinal nematodes in ostriches, *Struthio camelus*, in different regions of the state of Rio de Janeiro, Brazil

Nematoides gastrintestinais em avestruzes, *Struthio camelus*, de diferentes regiões do Estado do Rio de Janeiro, Brasil

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## Abstract

The ratite group is composed of ostriches, rheas, emus, cassowaries and kiwis. Little research has been done on parasitism in these birds. The aim of this study was to determine the distribution of infections by gastrointestinal nematodes in ostriches in the state of Rio de Janeiro. For this, fecal samples were collected from 192 on 13 farms. From each sample, four grams of feces were used to determine the eggs per gram of feces (EPG) count, by means of the McMaster technique. Part of the feces sample was used for fecal cultures, to identify 100 larvae per sample. The results were subjected to descriptive central trend and dispersion analysis, using confidence intervals at the 5% error probability level in accordance with the Student t distribution, and Tukey's test with a 95% confidence interval. The mean EPG in the state was 1,557, and the municipality of Três Rios had the lowest average (62). The city of Campos dos Goytacazes presented the highest mean EPG of all the municipalities analyzed. The northern region presented the highest mean EPG, followed by the southern, metropolitan, coastal lowland and central regions. *Libyostrongylus* species were observed on all the farms: *L. douglassii* predominated, followed by *L. dentatus* and *Codiostomum struthionis*.

**Keywords:** *Libyostrongylus douglassii*, *Libyostrongylus dentatus*, *Codiostomum struthionis*.

## Resumo

O grupo das ratitas é composto pelas avestruzes, emas, emús, cassuares e kiwi. São poucas as pesquisas sobre as parasitoses nessas aves. O objetivo deste estudo foi determinar a distribuição de infecções por nematóides gastrintestinais em avestruzes no Estado do Rio de Janeiro. Para tanto, foram coletadas amostras fecais de 192 avestruzes de 13 propriedades. De cada amostra, quatro gramas foram utilizados para a contagem de ovos por grama de fezes (OPG), pela da técnica de McMaster. Parte das fezes foi utilizada para cultivos fecais para identificação de 100 larvas por amostra. Os resultados foram submetidos à análise descritiva de tendência central e de dispersão, utilizando-se o intervalo de confiança ao nível de 5% de probabilidade de erro de acordo com a distribuição t de Student e teste de Tukey com intervalo de confiança de 95%. A média de OPG no Estado foi de 1.557, e o município de Três Rios obteve a menor média (62). A cidade de Campos dos Goytacazes teve a maior média de OPG de todos os municípios analisados. A Região Norte teve a maior média de OPG, seguida das regiões Sul, Metropolitana, Baixada Litorânea e Central. As espécies de *Libyostrongylus* foram observadas em todas as propriedades, predominando *L. douglassii*, seguido de *L. dentatus* e *Codiostomum struthionis*.

**Palavras-chave:** *Libyostrongylus douglassii*, *Libyostrongylus dentatus*, *Codiostomum struthionis*.

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## Introduction

Nematodes of the genera *Libyostrongylus* and *Codiotomum* were probably introduced into Brazil at the time when ostriches were also introduced, in the 1990s (AICHINGER et al., 2007). This will have been due to deficiencies in sanitary inspection, since there is no standardization by governmental agencies, regarding control over these birds' parasitic diseases, internal marketing or importation. Spreading of some helminth species is favored by their direct life cycle, lack of adequate knowledge among the producers regarding ostrich handling and parasitological control, and trade in these birds without verification of the presence of parasitism. The best procedure for avoiding infection and subsequent economic loss is to prevent introduction of the parasites into the flock (CRAIG & DIAMOND, 1996). As a biosafety measure, rearing other birds on the same farm needs to be avoided (COOPER, 2005), given that the potential for cross-transmission among other birds, especially among other ratites, has not been determined (HOBERG et al., 1995).

The clinical signs of infection by *Libyostrongylus* spp. include anorexia, weight loss, anemia (BARTON & SEWARD, 1993), generalized muscle weakness and lethargy (MUKARATIRWA et al., 2004). The pathogenicity caused by infection by *C. struthionis* is not well known. Oliveira et al. (2009) described an intestinal mucosa thickening where parasites were observed at high concentrations, with nodular areas in the distal third of the infected caeca, as well as hemorrhagic areas abutting small ulcers surrounded by edema. The concentration of *C. struthionis* found in infected birds was directly correlated with the severity of lesions observed in the caeca.

*Libyostrongylus* Lane, 1923, includes three species: *L. douglassii* (COBBOLD, 1882) Lane, 1923; *L. dentatus* Hoberg, Lloyd and Omar, 1995; and *L. magnus* Gilbert, 1937. All of these can infect the proventriculus of ostriches. Two of these species can be distinguished based on the morphology and morphometry of the posterior end of the infective larvae: *L. douglassii* has a short sheath tail with an acute termination, while *L. dentatus* has a long and filamentous sheath tail (EDERLI et al., 2008a).

The infective larvae of *L. magnus* have not yet been described. The larvae of *Libyostrongylus* spp. are characterized by a knob on the tip of the tail, which is not observed in the infective larvae of *Codiotomum struthionis* (EDERLI et al., 2008b). Moreover, the latter species occurs in the caeca of ostriches. The infective larvae of this species are similar to those of *L. dentatus*, with a long and filamentous sheath tail, but can be distinguished based on the morphology of the larval tail tip, which does not present a knob but ends in a finger-like tip (EDERLI et al., 2008b).

Differentiation of these species has become possible because of studies conducted by Ederli et al. (2008a) and Ederli et al. (2008b), which describe the infective larvae of these nematodes. Prior to these studies, the observed low occurrence of *L. dentatus* and *C. struthionis* may have been due to similarities between the infective larvae in species of these genera. In addition, these two species of *Libyostrongylus* exhibit different site specificities in the ostrich proventriculus: *L. douglassii* is located under the koilin layer, whereas *L. dentatus* is found within the koilin layer (EDERLI & OLIVEIRA, 2009). Prior to the latter study, both

of these species had been reported to be located below the koilin layer. *Libyostrongylus douglassii* is distributed worldwide and has been found in several countries on five continents, excluding Asia and Antarctica (PONCE GORDO et al., 2002). On other hand, *L. dentatus* has only been described in the United States (HOBERG et al., 1995) and Brazil (EDERLI et al., 2008c), while *C. struthionis* has only been reported in Europe (PONCE GORDO et al., 2002) and Brazil (EDERLI et al., 2008b).

The aim of the present study was to determine the distribution of infection by gastrointestinal nematodes in different regions of the state of Rio de Janeiro, Brazil, by means of fecal examination.

## Materials and Methods

Convenience samples of feces from 192 adult ostriches on 13 farms in nine municipalities in the state of Rio de Janeiro were collected soon after the birds defecated. This was done with aid of plastic bags, taking care not to collect feces that had been in contact with the ground. The samplings was done on birds in different paddocks, and at least one sample per paddock was collected. Four grams of feces per bird were used to quantify the number of eggs per gram (EPG), in accordance with Gordon & Withlock (1939) and four grams of feces were used to perform fecal cultures as described by Bonadiman et al. (2006). From the positive samples, we identified 100 larvae per animal, according to the differences in the posterior end of the larvae as described by Ederli et al. (2008a) and Ederli et al. (2008b), in order to analyze the frequencies of nematode species shed by ostriches.

The sampling was performed in clusters representing the main regions of the state of Rio de Janeiro (municipalities and farms), by selecting farms according to convenience so as to have greater representation and accessibility. The data were analyzed using descriptive central trend and dispersion analysis, using the confidence interval at the 5% probability level in accordance with the Student t distribution. The Tukey test was used to compare the percentage values of each nematode species by means of analysis on the larvae, per farm, municipality and region. All calculations were performed using the SAEG program, version 9.1.

## Results

In 13 ostrich-breeding farms in the state of Rio de Janeiro, the observed frequency of infected ostriches was 93% (179 out of 192 samples) (Table 1). The eggs presented the typical characteristics of Strongylida. The mean EPG in the state of Rio de Janeiro was 1,557.46, ranging from zero to 19,600. The municipality of Três Rios (central region) had the lowest mean, of 61.76, with a range from zero to 200. Campos dos Goytacazes (northern region) had the highest mean EPG, of 3,825.00, with a range from zero to 6,100. The northern region of the state had the highest mean EPG (3,190, ranging from zero to 10,050), followed by the southern region (2,433.33, ranging from 800 to 8,400), metropolitan region (2,033.30, ranging from 100 to 19,600), coastal lowland region (1,239.38, ranging from zero to 16,900) and central region (61.76, ranging from zero to 200). The results per farm, municipality and region are presented in Table 1.

**Table 1.** Eggs per gram of feces (EPG) counts and mean proportions of nematode species according to larvae identified from the feces of ostriches (*Struthio camelus*) in the state of Rio de Janeiro, Brazil. Mean  $\pm$  SE, followed by minimum and maximum values in parentheses and confidence interval (CI).

Regions/ Cities	Farm	Flock	N° of fecal samples	Positive samples	EPG	Infective larvae (%)		
						<i>L. douglassii</i>	<i>L. dentatus</i>	<i>C. struthionis</i>
<b>North</b>		<b>163</b>	<b>33</b>	<b>31</b>	<b>3,190.00<math>\pm</math>592.24 (50-10,050)</b> <b>CI = 1,211.26</b>	<b>86.28<math>\pm</math>2.37 (56-100)</b> <b>CI = 4.84<sup>A</sup></b>	<b>13.72<math>\pm</math>2.37 (0-44)</b> <b>CI = 4.84<sup>B</sup></b>	<b>0<sup>C</sup></b>
Campos dos Goytacazes		10	5	4	4,825.00 $\pm$ 566.24 (3,400-6,100) CI = 1,109.81	94.47 $\pm$ 3.87 (83-100) CI = 12.33 <sup>A</sup>	5.53 $\pm$ 3.87 (0-17) CI = 12.30 <sup>B</sup>	0 <sup>B</sup>
	1	10	5	4	4,825.00 $\pm$ 566.24 (3,400-6,100) CI = 1,109.81	94.47 $\pm$ 3.87 (83-100) CI = 12.33 <sup>A</sup>	5.53 $\pm$ 3.87 (0-17) CI = 12.30 <sup>B</sup>	0 <sup>B</sup>
São Francisco do Itabapoana		153	28	27	2,938.46 $\pm$ 666.75 (50-10,050) CI = 1,373.20	85.02 $\pm$ 2.60 (56-100) CI = 5.35 <sup>A</sup>	14.98 $\pm$ 2.60 (0-44) CI = 5.35 <sup>B</sup>	0 <sup>C</sup>
	2	98	15	15	4,728.57 $\pm$ 993.70 (100-10,050) CI = 2,146.76	81.36 $\pm$ 2.86 (60-97) CI = 6.17 <sup>A</sup>	18.64 $\pm$ 2.87 (3-40) CI = 6.17 <sup>B</sup>	0 <sup>C</sup>
	3	45	10	9	677.78 $\pm$ 237.77 (50-1,900) CI = 548.30	93.7 $\pm$ 4.75 (56-100) CI = 10.95 <sup>A</sup>	6.93 $\pm$ 4.75 (4-44) CI = 10.95 <sup>B</sup>	0 <sup>B</sup>
	4	10	3	3	1,366.67 $\pm$ 1,072.90 (100-3,500) CI = 4,616.31	78.00 $\pm$ 7.64 (68-93) CI = 32.86 <sup>A</sup>	22.00 $\pm$ 7.64 (7-32) CI = 32.86 <sup>B</sup>	0 <sup>B</sup>
<b>Littoral Lowland</b>		<b>776</b>	<b>121</b>	<b>111</b>	<b>1,239.38<math>\pm</math>231.18 (50-16,900)</b> <b>CI = 458.05</b>	<b>81.46<math>\pm</math>1.95 (0-100)</b> <b>CI = 3.87<sup>A</sup></b>	<b>16.69<math>\pm</math>1.89 (0-100)</b> <b>CI = 3.74<sup>B</sup></b>	<b>1.85<math>\pm</math>0.37 (0-20)</b> <b>CI = 0.72<sup>C</sup></b>
Araruama		32	18	15	640.00 $\pm$ 207.83 (50-2,600) CI = 445.76	81.64 $\pm$ 3.69 (50-99) CI = 7.92 <sup>A</sup>	16.55 $\pm$ 3.52 (1-48) CI = 7.54 <sup>B</sup>	1.81 $\pm$ 0.67 (0-8) CI = 1.44 <sup>C</sup>
	5	32	18	15	640.00 $\pm$ 207.83 (50-2,600) CI = 445.76	81.64 $\pm$ 3.69 (50-99) CI = 7.92 <sup>A</sup>	16.55 $\pm$ 3.52 (1-48) CI = 7.54 <sup>B</sup>	1.81 $\pm$ 0.67 (0-8) CI = 1.44 <sup>C</sup>
Cabo Frio		246	33	29	946.55 $\pm$ 460.90 (50-13,500) CI = 944.10	82.51 $\pm$ 3.15 (19-100) CI = 6.46 <sup>A</sup>	13.94 $\pm$ 3.03 (0-81) CI = 6.20 <sup>B</sup>	3.55 $\pm$ 1.03 (0-20) 2.11 <sup>C</sup>
	6	6	6	6	350.00 $\pm$ 102.47 (100-700) CI = 263.41	70.33 $\pm$ 11.32 (19-93) CI = 29.11 <sup>A</sup>	29.67 $\pm$ 11.32 (7-81) CI = 29.11 <sup>B</sup>	0 <sup>B</sup>
	7	240	27	23	1,102.17 $\pm$ 578.74 (50-13,500) CI = 1,200.24	85.69 $\pm$ 2.47 (59-100) CI = 5.13 <sup>A</sup>	9.84 $\pm$ 1.84 (0-31) CI = 3.82 <sup>B</sup>	4.48 $\pm$ 1.23 (0-20) CI = 2.55 <sup>B</sup>
Casimiro de Abreu		168	35	33	408.82 $\pm$ 125.73 (50-3,500) CI = 255.80	69.50 $\pm$ 4.86 (0-100) CI = 9.89 <sup>A</sup>	28.34 $\pm$ 4.78 (0-100) 9.73 <sup>B</sup>	2.15 $\pm$ 0.68 (0-15) 1.39 <sup>C</sup>
	8	120	21	20	554.76 $\pm$ 195.85 (50-3,300) CI = 408.53	71.80 $\pm$ 7.05 (0-100) CI = 14.7 <sup>A</sup>	28.20 $\pm$ 7.05 (0-100) CI = 14.7 <sup>B</sup>	0 <sup>C</sup>
	9	48	14	13	173.08 $\pm$ 54.19 (50-600) CI = 118.07	65.79 $\pm$ 5.87 (34-96) CI = 12.80 <sup>A</sup>	28.57 $\pm$ 5.57 (2-65) CI = 12.14 <sup>B</sup>	5.63 $\pm$ 1.32 (0-15) CI = 2.87 <sup>C</sup>
Saquarema		330	35	34	2,545.71 $\pm$ 569.55 (50-16,900) CI = 1,157.47	92.11 $\pm$ 1.27 (69-100) CI = 2.58 <sup>A</sup>	7.71 $\pm$ 1.20 (0-26) CI = 2.44 <sup>B</sup>	0.17 $\pm$ 0.14 (0-5) CI = 0.29 <sup>C</sup>
	10	330	35	34	2,545.71 $\pm$ 569.55 (50-16,900) CI = 1,157.47	92.11 $\pm$ 1.27 (69-100) CI = 2.58 <sup>A</sup>	7.71 $\pm$ 1.20 (0-26) CI = 2.44 <sup>B</sup>	0.17 $\pm$ 0.14 (0-5) CI = 0.29 <sup>C</sup>
<b>Central</b>		<b>180</b>	<b>17</b>	<b>16</b>	<b>61.76<math>\pm</math>9.12 (50-200)</b> <b>CI = 19.34</b>	<b>11.21<math>\pm</math>3.88 (0-61)</b> <b>CI = 8.23<sup>A</sup></b>	<b>88.79<math>\pm</math>3.88 (39-100)</b> <b>CI = 8.23<sup>B</sup></b>	<b>0<sup>C</sup></b>
Três Rios		180	17	16	61.76 $\pm$ 9.12 (50-200) CI = 19.34	11.21 $\pm$ 3.88 (0-61) CI = 8.23 <sup>A</sup>	88.79 $\pm$ 3.88 (39-100) CI = 8.23 <sup>B</sup>	0 <sup>C</sup>
	11	180	17	16	61.76 $\pm$ 9.12 (50-200) CI = 19.34	11.21 $\pm$ 3.88 (0-61) CI = 8.23 <sup>A</sup>	88.79 $\pm$ 3.88 (39-100) CI = 8.23 <sup>B</sup>	0 <sup>C</sup>
<b>South</b>		<b>10</b>	<b>6</b>	<b>6</b>	<b>2,433.33<math>\pm</math>1,199.07 (800-8,400)</b> <b>CI = 3,082.32</b>	<b>92.08<math>\pm</math>2.07 (86-100)</b> <b>CI = 5.32<sup>A</sup></b>	<b>6.43<math>\pm</math>1.78 (0-13)</b> <b>CI = 4.57<sup>B</sup></b>	<b>1.50<math>\pm</math>0.96 (0-6)</b> <b>CI = 2.46<sup>B</sup></b>
Valença		10	6	6	2,433.33 $\pm$ 1,199.07 (800-8,400) CI = 3,082.32	92.08 $\pm$ 2.07 (86-100) CI = 5.32 <sup>A</sup>	6.43 $\pm$ 1.78 (0-13) CI = 4.57 <sup>B</sup>	1.50 $\pm$ 0.96 (0-6) CI = 2.46 <sup>B</sup>
	12	10	6	6	2,433.33 $\pm$ 1,199.07 (800-8,400) CI = 3,082.32	92.08 $\pm$ 2.07 (86-100) CI = 5.32 <sup>A</sup>	6.43 $\pm$ 1.78 (0-13) CI = 4.57 <sup>B</sup>	1.50 $\pm$ 0.96 (0-6) CI = 2.46 <sup>B</sup>
<b>Metropolitan</b>		<b>20</b>	<b>15</b>	<b>15</b>	<b>78.05<math>\pm</math>3.61 (46-100)</b> <b>CI = 2,719.15</b>	<b>14.47<math>\pm</math>2.92 (0-38)</b> <b>CI = 7.74<sup>A</sup></b>	<b>7.48<math>\pm</math>1.84 (0-20)</b> <b>CI = 6.26<sup>B</sup></b>	<b>7.48<math>\pm</math>1.84 (0-20)</b> <b>CI = 3.95<sup>B</sup></b>

Proportions mean in the lines with the same superscript letters do not differ significantly by Tukey test with a confidence interval of 95%. P value < 0.05.

Table 1. Continued...

Regions/ Cities	Farm	Flock	Nº of fecal samples	Positive samples	EPG	Infective larvae (%)		
						<i>L. douglassii</i>	<i>L. dentatus</i>	<i>C. struthionis</i>
Miguel Pereira		20	15	15	2,033.33±1,267.79 (100-19,600)	78.05±3.61 (46-100)	14.47±2.92 (0-38)	7.48±1.84 (0-20)
					CI = 2,719.15	CI = 7.74 <sup>A</sup>	CI = 6.26 <sup>B</sup>	CI = 3.95 <sup>B</sup>
	13	20	15	15	2,033.33±1,267.79 (100-19,600)	78.05±3.61 (46-100)	14.47±2.92 (0-38)	7.48±1.84 (0-20)
					CI = 2,719.15	CI = 7.74 <sup>A</sup>	CI = 6.26 <sup>B</sup>	CI = 3.95 <sup>B</sup>
<b>TOTAL</b>	<b>13</b>	<b>1,149</b>	<b>192</b>	<b>179</b>	<b>1,557.46±241.54 (50-19,600)</b>	<b>75.73±2.07 (0-100)</b>	<b>22.45±2.07 (0-100)</b>	<b>1.83±0.31 (0-20)</b>
					CI = 423.35	CI = 4.08 <sup>A</sup>	CI = 4.08 <sup>B</sup>	CI = 0.61 <sup>C</sup>

Proportions mean in the lines with the same superscript letters do not differ significantly by Tukey test with a confidence interval of 95%. P value < 0.05.

Municipalities with mean EPG greater than 2,500 (Campos dos Goytacazes, São Francisco do Itabapoana and Saquarema) differed statistically from those with EPG less than 650 (Araruama, Casimiro de Abreu and Três Rios). Those with mean EPG near the median, ranging from 940 to 2400 (Cabo Frio, Valença and Miguel Pereira) did not differ statistically from other municipalities.

An analysis on the infective larvae recovered from fecal cultures confirmed the presence of *L. douglassii*, *L. dentatus* and *C. struthionis*, which infect ostrich flocks in the state of Rio de Janeiro, Brazil (Figure 1). Species of the genus *Libyostrongylus* were observed on all 13 farms analyzed. However, *C. struthionis* was observed only on six farms, with low proportions that never exceeded 7.5%. Infections of *C. struthionis* were not present in the northern region, where four farms in two municipalities (Campos dos Goytacazes and São Francisco do Itabapoana) were examined, or in the central region, where fecal samples from a single farm in the municipality of Três Rios were examined. In addition, *C. struthionis* was not identified on one of the two farms that we analyzed in the municipality of Cabo Frio, which is in the coastal lowland region, where ostriches on six farms were analyzed. On the single farm analyzed in the municipality of Saquarema, *C. struthionis* was observed at an extremely low proportion (0.17%), such that only two ostriches out of 35 were positive for this species, with counts of one and five larvae out of 100. In all the birds in which *C. struthionis* was diagnosed, infection by *L. douglassii* and *L. dentatus* was also present. Among the positive samples, only two birds were positive only for *L. dentatus*, and both of these were in Três Rios (central region).

Species of *Libyostrongylus* were found on all the 13 farms analyzed, with proportions ranging from 0 to 100% (Table 1). There were significant differences in the proportions of *L. douglassii*, *L. dentatus* and *C. struthionis*, such that *L. douglassii* was the most frequent species and *C. struthionis* the least frequent in the state of Rio de Janeiro. However, in the municipality of Três Rios (central region), there were unusual proportions of these three species, such that we observed a proportion of *L. dentatus* that was statistically greater than that of *L. douglassii* (Table 1). In this municipality, no *C. struthionis* larvae were present in ostrich feces.

With the exception of the farm located in the municipality of Três Rios (central Region), which had a mean *L. douglassii* proportion of 11.21, all the other farms presented mean proportions ranging from 65.7 to 94.4. The low proportion of *L. douglassii* and high

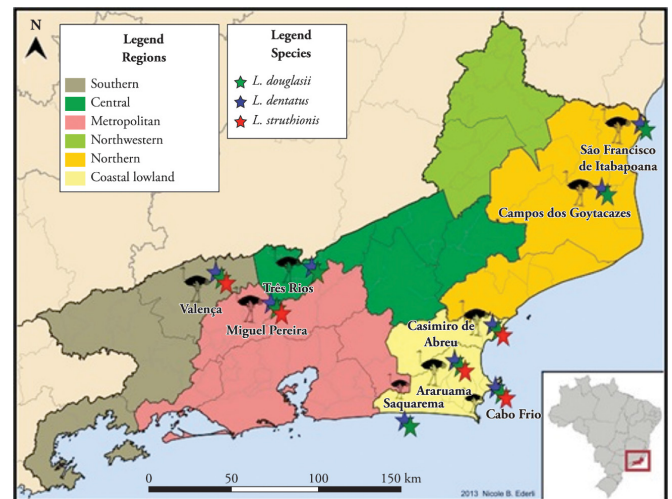


Figure 1. Map of the state of Rio de Janeiro, Brazil, including regions and cities, showing the distribution of gastrointestinal nematode infections.

proportion of *L. dentatus* observed on the farm in the municipality of Três Rios was statistically different from all other farms.

The metropolitan and southern regions of the state of Rio de Janeiro did not show any statistical difference between the proportions of *L. dentatus* and *C. struthionis* (Table 1). Although there was a statistical difference between the proportions of these three species in the northern region, some farms (one in Campos dos Goytacazes and two in São Francisco do Itabapoana) did not present any statistical difference in the proportions of *C. struthionis* and *L. dentatus*. The same was observed on one farm in the municipality of Cabo Frio (coastal lowland region).

## Discussion

The results presented here describe occurrences of gastrointestinal nematodes in ostriches in the state of Rio de Janeiro, Brazil. These infections are present on ostrich farms in the state of Rio de Janeiro, with predominance of *L. douglassii*, followed by *L. dentatus* and *C. struthionis* (Table 1). This pattern was also observed by Andrade et al. (2011), based on a study in which fecal examination was performed on ostriches on 17 farms distributed across different Brazilian states, including 42 samples from only three farms in the state of Rio de Janeiro. They did not observe any larvae in the feces

from four farms, including one in the state of Ceará (where only one farm analyzed), one in the state of Rio de Janeiro (three farms analyzed) and two in the state of Paraná (four farms analyzed). However, the authors did not identify the municipalities where the samples were collected. Andrade et al. (2011) did not perform EPG counts, because they received fecal samples sent by express mail, thus constraining this procedure. In our study, we analyzed the proportions of infective larvae of *L. douglassii*, *L. dentatus* and *C. struthionis*. Furthermore, we determined the parasite load in the Rio de Janeiro flock, thus showing the actual situation of this parasitosis in this state.

In our study, *Libyostrongylus* species were present on all the farms examined, while *C. struthionis* was not found only in the northern or central regions of the state of Rio de Janeiro. The frequency of *L. dentatus* infection statistically exceeded that of *L. douglassii* on one farm in the municipality of Três Rios. Andrade et al. (2011) also observed higher frequency of *L. dentatus* than of *L. douglassii* only on one farm, in the state of São Paulo, with mean percentages of infective larvae of *L. dentatus* and *L. douglassii* of 57% and 13%, respectively. In addition, on this same farm, the mean percentage of *C. struthionis* was also higher than that of *L. douglassii*. This was not observed on any farm in our study. The standard-deviations relating to the EPG analyses were higher than the means. This shows that there was great variability in the EPG counts from the flocks in the state of Rio de Janeiro.

These parasites were introduced into Brazil through the importation of ostriches in 1995, with the first imports of birds from the United States and Spain (AICHINGER et al., 2007). Today, these parasites are widely distributed and display a high level of adaptation within this country, with prevalence rates of 100%. However, *Libyostrongylus dentatus* and *C. struthionis* are only infrequently reported, probably because until recently they could only be properly identified via necropsy on the birds, in order to collect and identify adult parasites. *Libyostrongylus dentatus* has exclusively been reported in the United States (HOBERG et al., 1995) and Brazil (EDERLI et al., 2008c), while *C. struthionis* has only been reported in Europe (PONCE GORDO et al., 2002) and Brazil (EDERLI et al., 2008b). In contrast, *L. douglassii* is distributed worldwide and has been found in South Africa (MALAN et al., 1988; FOCKEMA et al., 1985; REINECKE, 1983), Australia (BARTON & SEWARD, 1993; BUTTON et al., 1993; MORE, 1996), the United States (HOBERG et al., 1995), Italy (PINTORI et al., 2000), Scotland (PENNYCOTT & PATTERSON, 2001), Spain, Belgium, Portugal and the Netherlands (PONCE GORDO et al., 2002), Sweden (JANSSON et al., 2002), New Zealand (MACKERETH, 2004; MCKENNA, 2005), Zimbabwe (MUKARATIRWA et al., 2004) and Brazil (BONADIMAN et al., 2006; EDERLI et al., 2008a, c).

Recently, Fagundes et al. (2012) described the seasonality and occurrence of *C. struthionis* on two farms in the municipalities of Areal and Itaboraí in the state of Rio de Janeiro, in which they analyzed 237 and 195 samples, respectively. However, the images of the posterior end of the infective larvae reported by Fagundes et al. (2012) are clearly from *L. dentatus*, since the larvae have a knob on the tip of the tail and a long, filamentous tail sheath. Thus, the seasonality described by Fagundes et al. (2012) for *C. struthionis* is likely based on mixed infection, since we too have identified mixed

infections of *Libyostrongylus* spp. and *C. struthionis* in the state of Rio de Janeiro, including these same regions (metropolitan and central). In these regions, we identified farms that were negative for infection and that displayed low frequencies of *C. struthionis*, as observed in other regions of the state of Rio de Janeiro (Table 1). This observation was also made by Andrade et al. (2011) in relation to several regions of Brazil, in which they observed *C. struthionis* on only three of the 17 farms analyzed.

In Brazil, commercial ostrich breeding is becoming common, but not widely distributed throughout the country, because the Brazilian Institute for the Environment and Natural Resources (IBAMA) regulates breeding.

Breeding of emus (*Dromaius novaehollandiae*), another ratite bird, especially in the central-western region of Brazil, has recently been started in this country. The potential for cross-transmission of these parasites to other birds, including ratites in particular, has not yet been investigated. There is a report describing these parasites in emus in Sweden (JANSSON & CHRISTENSSON, 2000), but this observation has not been confirmed by identification of infective larvae or adult nematodes because these authors reported eggs in bird feces. Finally, it is also important to examine possible infection by these nematodes in the rhea, a native bird of South America.

Thus, the present study demonstrates that gastrointestinal nematodes infecting ostriches are widely distributed in the state of Rio de Janeiro, occurring in an enzootic form in the flock with a high parasite load, in which *L. douglassii* predominates, followed by *L. dentatus* and *C. struthionis*. These results demonstrate that these exotic parasites have attained good adaptation to the region studied and possibly to the whole country.

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